

1 Stepwise linear discriminant analysis to  
2 differentiate Spanish red wines by their  
3 Protected Designation of Origin or category  
4 using physico-chemical parameters  
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## 11 Abstract

12 **Aim:** The aim of this work was to determine the physico-chemical variables that differentiating red wines from  
13 the “Castilla y León” Spanish region by their Protected Designation of Origin (PDO) and wine category  
14 (“young”, “oak”, “crianza”, or “reserve”).

15 **Methods and results:** A total of 135 commercial red wines from four Spanish PDOs in the region of Castilla and  
16 León were analysed. Forty physico-chemical parameters, related to classical enological parameters, phenolic and  
17 polysaccharidic composition, and content of higher alcohols were evaluated. Differences in physico-chemical  
18 composition were found in red wines from different PDOs and different categories. Stepwise linear discriminant  
19 analysis (SLDA) was applied to find a linear combination of the physico-chemical variables that separate and  
20 classify the red wines according to the PDO or category. One SLDA model selected 15 physico-chemical  
21 variables that allowed for good discrimination and classification of the wines from different PDOs. The SLDA  
22 model selected seven variables for wine category differentiation, but only allowed for good discrimination  
23 between young wines and aged wines (“crianza” and “reserve”).

24 **Conclusions:** The variables that contributed most to the separation of *Tempranillo* red wines were total  
25 polyphenols, total tannins, and absorbance values at 230 nm and 280 nm. The polysaccharides with an average  
26 molecular weight of 10 kDa, flavanols, stilbenes and 2-methyl-1-butanol were those most associated with the  
27 differentiation of the wines elaborated with the *Mencía* grape variety. The percentage of polymeric anthocyanins  
28 and absorbance at 230 nm could be proposed as good indicators for aged wines, and total tannins for young  
29 wines.

30 **Significance and impact of the study:** This study provides improved knowledge of the physico-chemical  
31 variables that could be used as markers of the origin of wines and/or the grape variety (*Tempranillo* and *Mencía*)  
32 and that allow differentiating young wines from those aged for a long time.

33 **Keywords:** higher alcohols; organic acids; phenolic compounds; polysaccharides; Protected Designation of  
34 Origin; red wine categories; stepwise linear discriminant analysis

35

## 36 Introduction

37 Wine is widely known as a complex matrix composed mainly of water and ethanol and, to a lesser extent, a  
38 large number of chemical compounds such as phenolic compounds, polysaccharides, non-fermentable  
39 sugars, organic acids, glycerol, volatile compounds, etc. The content of these compounds has an important  
40 role in the quality of wines, which can vary depending on the grape variety and the enological technology  
41 used in the winemaking process, and several environmental aspects such as soil, geographical location, and  
42 weather conditions (Riu-Aumatell *et al.*, 2002; Monagas *et al.*, 2005a; Robinson *et al.*, 2012; Serrano-  
43 Lourido *et al.*, 2012).

44 Wine phenolic compounds play an important role in the sensory properties and have been proposed as  
45 chemical markers of the geographical origin of grapes (Makris *et al.*, 2006). Anthocyanins are directly  
46 responsible for color in grapes and young wines (Glories, 1984). Monomeric anthocyanins are  
47 progressively transformed into more stable oligomeric and polymeric pigments during wine aging  
48 (Monagas *et al.*, 2005b) due to several chemical reactions producing changes in the color of the wine.  
49 Other phenolic compounds such as flavanols, flavonols, hydroxycinnamic and hydroxybenzoic acid  
50 derivatives can contribute to modifying the different sensory properties of red wines, such as astringency,  
51 bitterness, structure and color (Gawel, 1998; Schwarz *et al.*, 2005; Hufnagel and Hofmann, 2008; Sáenz-  
52 Navajas *et al.*, 2010).

53 Wine polysaccharides are compounds that come from the grapes and yeasts involved in the fermentation  
54 process and they can have a positive effect on the technological and sensory characteristics of wines. They  
55 can interact with phenolic compounds, reducing wine astringency and bitterness (del Barrio-Galán *et al.*,  
56 2011; González-Royo *et al.*, 2013; del Barrio-Galán *et al.*, 2015), and improving mouthfeel (Vidal *et al.*,  
57 2004) and stabilization (Poncet-Legrand *et al.*, 2007) of wines. However, some types of wine  
58 polysaccharides can also have several negative effects on the technological process of winemaking, such as  
59 the filtration process (Belleville *et al.*, 1991).

60 Other wine compounds, such as organic acids, ethanol, glycerol and higher alcohols, can make an  
61 important contribution to wine characteristics. The main organic acids present in red wines are tartaric acid  
62 and lactic acid which are mainly responsible for acidity (Mato *et al.*, 2005), and have an important role in  
63 physico-chemical stabilization, and the balance and sensory perception of wines (Silva *et al.*, 2015). The  
64 ethanol content depends on the sugar content of grapes and influences the sensory characteristics of wines,  
65 increasing the bitterness and reducing the astringency (Fontoin *et al.*, 2008; Rinaldi *et al.*, 2012). Glycerol  
66 can also have an influence on several sensory properties of wines, such as sweetness and body (Noble and  
67 Bursick, 1984), and is mainly produced by yeast during alcoholic fermentation (Remize *et al.*, 2001).  
68 Finally, higher alcohols are formed during fermentation, and could have a positive or negative effect on  
69 wine sensory properties depending on their concentration (de la Fuente-Blanco *et al.*, 2017).

70 The identification of a wine's geographical origin has an important commercial role in the wine industry  
71 (Urvieta *et al.*, 2018). This is an important factor that consumers consider (Famularo *et al.*, 2010), and  
72 many are highly oriented to the consumption of high-quality wines (Urvieta *et al.*, 2018). Wine category  
73 (young or aged) also influences consumer choice; during oak barrel and bottle aging, the structure of the  
74 phenolic compounds changes and the physical, chemical and sensory properties of the wines can modify  
75 the quality.

76 Various studies have focused on differentiating wines geographical regions and country, according to their  
77 physico-chemical and sensory parameters and through using multivariate statistical tools (Pérez-Magariño  
78 and González-San José, 2001; Cliff *et al.*, 2007; Riovanto *et al.*, 2011; Serrano-Lourido *et al.*, 2012). Other  
79 studies have focused on monitoring the aging time of red wines using different physico-chemical  
80 parameters (Agazzi *et al.*, 2018; Astray *et al.*, 2019). However, to our knowledge this is the first study that  
81 uses wines from several important Spanish Protected Designations of Origin (PDOs) that are very close

82geographically. Therefore, the aim of this work was to determine the physico-chemical parameters that  
83differentiate the red wines from four PDOs in Castilla y León and those PDO red wines by their category  
84(“young”, “oak”, “crianza” and “reserve”).

## 85Material and methods

### 86 1. 1. Wines

87A total of 135 commercial red wines from four Spanish PDOs in the region of Castilla y León (north  
88Spain), were analyzed: 76 wines from Ribera del Duero (RD), 21 wines from Bierzo (BI), 22 wines  
89from Toro (TO) and 16 wines from Cigales (CI). These wines were from different vintages (2003 to  
902016).

91There are certain specifications from the Regulatory Councils of each PDO. The wines from RD and  
92TO must be elaborated with at least 75 % *Tempranillo* grape variety; the wines from RD can be  
93blended with other varieties, such as *Cabernet Sauvignon*, *red Grenache*, *Malbec*, *Merlot* and *Albillo*,  
94and those from TO can only be blended with the *red Grenache* grape variety. The wines from CI must  
95be elaborated with at least 50 % *Tempranillo* and/or *red and gray Grenache* and can then be blended  
96with other authorized grape varieties (*Cabernet Sauvignon*, *Merlot* and *Syrah*). Finally, the wines from  
97BI must be elaborated with at least 70 % *Mencía* grape variety and can be blended with *Tintorera*  
98*Grenache*.

99The wines from each PDO were classified into four categories: 1) “young” - not aged in oak barrels; 2)  
100“oak” - aged in oak barrels for more than than 3 months; 3) “crianza” - a minimum aging period of 24  
101months, with at least 12 of these months in oak barrels; 4) “reserve” - a minimum aging period of 36  
102months, with at least 12 of these months in oak barrels. The remaining aging time for “crianza” and  
103“reserve” wines must be done in the bottle. Table 1 shows the number of wines in each category for  
104each PDO.

105Table 1. Number of wines analyzed in each category for each PDO.

	Ribera del Duero	Bierzo	Toro	Cigales	Total
Young	14	7	7	4	32
Oak	21	5	7	4	37
Crianza	21	7	4	4	36
Reserve	20	2	4	4	30
Total	76	21	22	16	135

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### 107 2. 2. Reagents and standards

108Phenolic compound standards were supplied by Sigma-Aldrich (Steinheim, Germany), Fluka (Buchs,  
109Switzerland), and Extrasynthèse (Lyon, France). Polysaccharide standards were supplied by Sigma-  
110Aldrich.

111Major volatile compound standards were supplied by Fluka, Sigma-Aldrich and Alfa Aesar  
112(Lancashire, UK). Organic acid standards were supplied by Panreac (Madrid, Spain). Glucose,  
113fructose and glycerol standards were supplied by Sigma-Aldrich.

114The ethanol for high performance liquid chromatography (HPLC) analyses was provided by Panreac,  
115and the ethanol 96 % was from Labkem (Spain). Acetonitrile and methanol for HPLC analyses were

116supplied by Carlo Erba (Sabadell, Spain) and the remaining reagents by Panreac. Water Milli-Q was  
117obtained through a Millipore system (Bedford, MA).

118Helium BIP (99.9997 %), air zero (99.998 %) and premier plus hydrogen (99.9998 %) for gas  
119chromatography (GC) were provided by Carbueros Metálicos S.A. (Valladolid, Spain).

### 120 3. 3. Analytical methods

121Ethanol, total and free SO<sub>2</sub>, total acidity (TA) and pH were determined according to the official  
122methods of OIV (2015).

123The color intensity, tonality and percentage of blue tones (% blue) were evaluated as indicates in  
124Glories (1984). The total polyphenols (TP) (expressed in mg/L of gallic acid) and total anthocyanins  
125(expressed in mg/L of malvidin-3-glucoside) were analyzed according to the methods described in  
126Pérez-Magariño *et al.* (2009). Total tannins (TT) (expressed in mg/L of catechin) were analyzed  
127according to methods in Mercurio *et al.* (2007) and polymeric anthocyanins (polymeric ACY)  
128(expressed in percentage) according to Levenson and Boulton (2004). Absorbances at 230 nm and  
129280 nm (A230 and A280) were also measured because of their correlation with the phenolic content of  
130the wines. These absorbances were measured with a quartz cuvette (1 cm of path length) after sample  
131dilution 1:400 with Milli-Q water. These physico-chemical parameters were all measured using an  
132UV/Vis Agilent Cary 60 spectrophotometer (Santa Clara, California, USA).

133Low molecular weight phenolic compounds were analyzed using High Performance Liquid  
134Chromatography coupled to a diode array detector (Agilent Technologies 1100 Series, HPLC-DAD  
135system). The samples were directly injected following the chromatographic conditions described in  
136Pérez-Magariño *et al.* (2008). A reverse-phase Zorbax SB-C18 column was used, provided by Agilent  
137(250 mm × 4.6 mm i.d., 3.5 µm particle size). The different compounds determined and quantified  
138(expressed in mg/L of different phenolic standards) were grouped as follows: hydroxybenzoic acids  
139(gallic, protocatechuic, vanillic, syringic, ellagic acids, and ethyl gallate); hydroxycinnamic acids  
140(*trans*-caffeic and *trans*-*p*-coumaric acids); tartaric esters of hydroxycinnamic acids (*trans*-caftaric,  
141*cis*-coutaric, *trans*-coutaric and *trans*-fertaric acids; and two *trans*-*p*-coumaric acid hexose esters);  
142flavanols (catechin and epicatechin); flavonols and derivatives (myricetin and myricetin-3-glycosides,  
143quercetin and quercetin-3-glycosides, kaempferol, isorhamnetin and syringetin-3-glucoside); phenolic  
144alcohols (tyrosol and tryptophol); and stilbenes (*trans*-resveratrol-3-glucoside, *cis*-resveratrol-3-  
145glucoside, and *trans*-resveratrol).

146The extraction and quantification of soluble polysaccharides (expressed in mg/L of dextrans) was  
147carried out following the methodology described by Guadalupe *et al.* (2012). These compounds were  
148analyzed using high performance size exclusion chromatography coupled to a refractive index detector  
149(Agilent Technologies 1100 Series, HPSEC-RID system), using two Shodex columns in serie: (OHpak  
150SB-803 HQ and OHpak SB-804 HQ; 300 mm × 8 mm i.d.; Showa Denko, Tokio, Japan). Seven  
151analytical standards of dextran from *Leuconostoc mesenteroides* were used for the molecular weight  
152calibration. Dextran with a 270 kDa molecular weight and one pectin (esterified potassium salt from  
153citrus fruit) were used as external standards for quantification. This methodology makes it possible to  
154separate four polysaccharide fractions according to their molecular weight: PS1 (polysaccharides with  
155a molecular weight average of 150 kDa); PS2 (polysaccharides with a molecular weight average of 60  
156kDa); PS3 (polysaccharides with a molecular weight average of 10 kDa); and PS4 (polysaccharides  
157with a molecular weight average of 7 kDa).

158Organic acids (tartaric, malic, lactic and acetic acids), glucose, fructose and glycerol were analyzed  
159according to the methodology described in Monteiro-Coelho *et al.* (2018), using HPLC coupled to a

160DAD and a RID, with some modifications. Briefly, 2 mL of wine was filtered through an 0.45  $\mu\text{m}$   
161PVDF filter and a volume of 20  $\mu\text{L}$  was injected. The column was an AMINEX HPX-87H (300  $\times$  7.8  
162mm) with internal particles of 9.0  $\mu\text{m}$  (Bio Rad, California, USA). The flow rate applied was 0.6  
163mL/min using 4.0 mM  $\text{H}_2\text{SO}_4$  as a mobile phase. The temperature of the column oven was maintained  
164at 65  $^\circ\text{C}$  and the RID flow cell was kept at 30  $^\circ\text{C}$ . The quantification was carried out at 205 nm for  
165organic acids and by RID for glucose, fructose and glycerol.

166Higher alcohols (acetaldehyde, ethyl acetate, methanol, diacetyl, 1-propanol, isobutanol, 1-butanol, 2-  
167methyl-1-butanol, 3-methyl-1-butanol, expressed in mg/L of the corresponding standard), were  
168analyzed following the method described in Pérez-Magariño *et al.* (2019), using a gas chromatograph  
169with flame ionization detector (GC-FID).

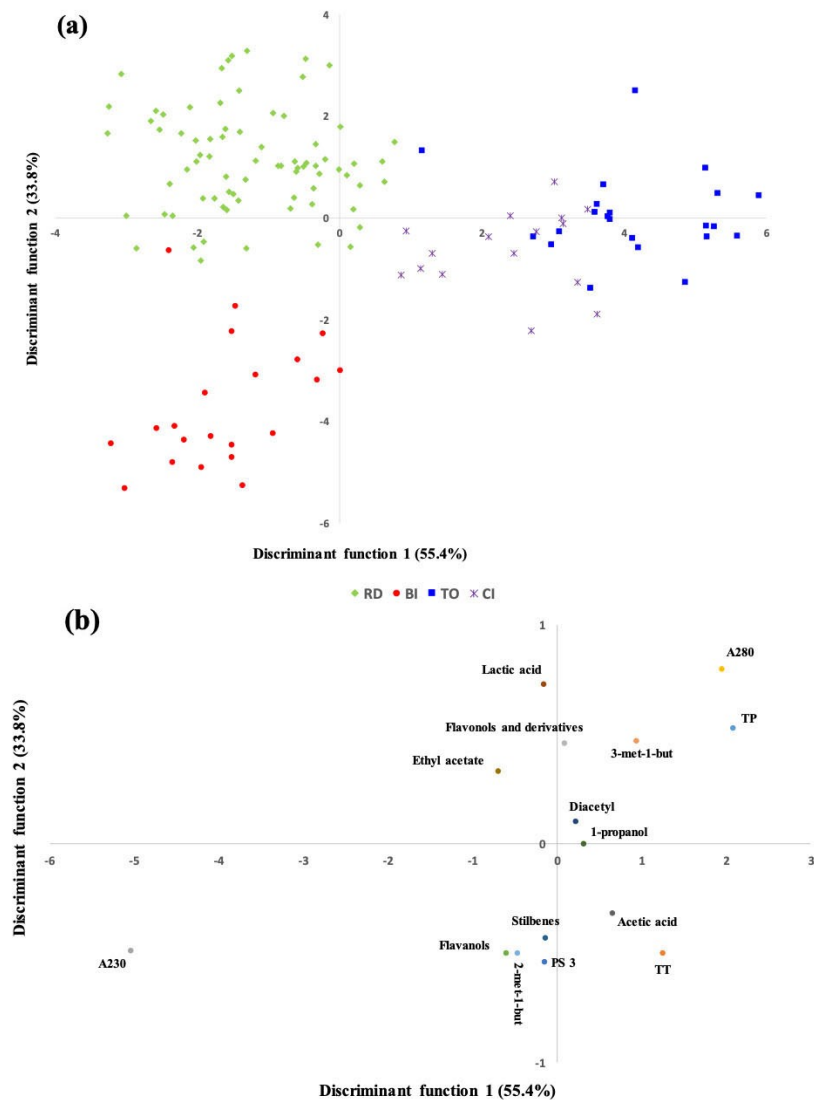
#### 170 4. 4. Statistical analyses

171Stepwise linear discriminant analysis (SLDA) was applied to find a linear combination of the variables  
172that characterize or separate two or more classes of objects. In this study, the forward method was used  
173to select the variables most useful for differentiating the wines according to PDO or wine category.  
174This procedure begins with no variables in the model and adds the variables with the highest  
175discriminant power. The selection of variables carried out by the model was done using the  $F$  statistic  
176(minimum  $F$  value = 4). The goodness of the prediction capacity of the SLDA discriminant models  
177was evaluated by cross-validation in four steps, and in each step excluding 25 % of the cases. An  
178analysis of variance (ANOVA) and a least significant difference test (LSD) at a significance level of  $p$   
179< 0.05 was performed, with the physico-chemical variables selected by the SLDA models according to  
180PDO and category criteria for explaining significant differences in the content of the different wines.  
181The statistical analyses were carried out with standardized data, using the Statgraphics Centurion XVII  
182statistical package.

### 183Results and discussion

184Forty physico-chemical variables were used in the SLDA to determine which ones allow to separating  
185the red wines according to their PDO and their category. The variables included were as follows: three  
186color parameters (color intensity, tonality and percentage of blue tones); four phenolic groups (TP, TT,  
187total anthocyanins, polymeric ACY); seven groups of low molecular weight phenolic compounds  
188(hydroxybenzoic and hydroxycinnamic acids, tartaric esters of hydroxycinnamic acids, flavanols,  
189flavonols and derivatives, phenolic alcohols and stilbenes); two absorbance values related to the  
190phenolic content (A230 and A280); four polysaccharide fractions (PS1, PS2, PS3 and PS4) and total  
191polysaccharides content; four organic acids (tartaric, malic, lactic and acetic) and their total content;  
192nine higher alcohols (acetaldehyde, ethyl acetate, methanol, diacetyl, 1-propanol, isobutanol, 1-  
193butanol, 2-methyl-1-butanol, 3-methyl-1-butanol); four classic oenological parameters (pH, ethanol,  
194total acidity and glucose plus fructose) and glycerol.

195The best model obtained by the SLDA selected the 15 physico-chemical variables that most  
196contributed to the differentiation of wines from different PDOs. The variables selected were as follows  
197(from highest to lowest discriminant power according to the  $F$  statistic values): flavonols and  
198derivatives, flavanols, PS3, ethyl acetate, lactic acid, stilbenes, acetic acid, 3-methyl-1-butanol, A230,  
199total tannins, A280, total polyphenols, 1-propanol, diacetyl, and 2-methyl-1-butanol. The highest  
200discriminant power of flavanols and flavonols are in agreement with the results reported by Makris *et*  
201*al.* (2006) as good indicators for differentiating red wines from Greece according to their geographical  
202origin.



203

204 **Figure 1.** Plot of scores (a) and loadings (b) for PDO wine differentiation using  
 205 discriminant functions 1 and 2.

206 *TP: total polyphenols. TT, total tannins; PS3 (polysaccharides with a molecular*  
 207 *weight average of 10 kDa) 2-met-1-but, 2-methyl-1-butanol; 3-met-1-but, 3-*  
 208 *methyl-1-butanol.*

209

210 The SLDA selected three discriminant functions that explained the total variance. By using the three  
 211 discriminant functions it was possible to separate the wines according to their PDO. Figures 1a and 1b  
 212 show the scores and loadings represented in the plane of the two first discriminant functions, which  
 213 explained 89.2 % of the total variance. As can be seen in Figure 1a, the first discriminant function,  
 214 which explained 55.4 % of the total variance, allowed a separation between the wines from RD and BI  
 215 (located on the left of the plane) and the wines from TO and CI (located on the right of the plane).  
 216 Thus, the physico-chemical variables on the left of the plane were more associated with the separation  
 217 of the wines from RD and BI, as A230 contributed most to this separation (Figure 1b). The ANOVA  
 218 results show that the value of A230 was significantly higher in the wines from RD than in the others  
 219 (Table 2). The variables on the right of the plane (A280, TP, TT, 3-methyl-1-butanol and acetic acid)  
 220 were more associated with the separation of the wines from TO and CI. The A230 and A280  
 221 measurements have been commonly used for a rapid determination of the total phenolic content in

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222wines (Kennedy *et al.*, 2006; Boulet *et al.*, 2016) due to their high correlation with these wine  
 223compounds. Nevertheless, the relative contribution of A230 with tannins, other polyphenols, or other  
 224wine compounds contents, are not clear yet (Boulet *et al.*, 2016).

225Table 2. Mean values  $\pm$  standard deviation of the variables selected by the SLDA  
 226for wine PDO discrimination.

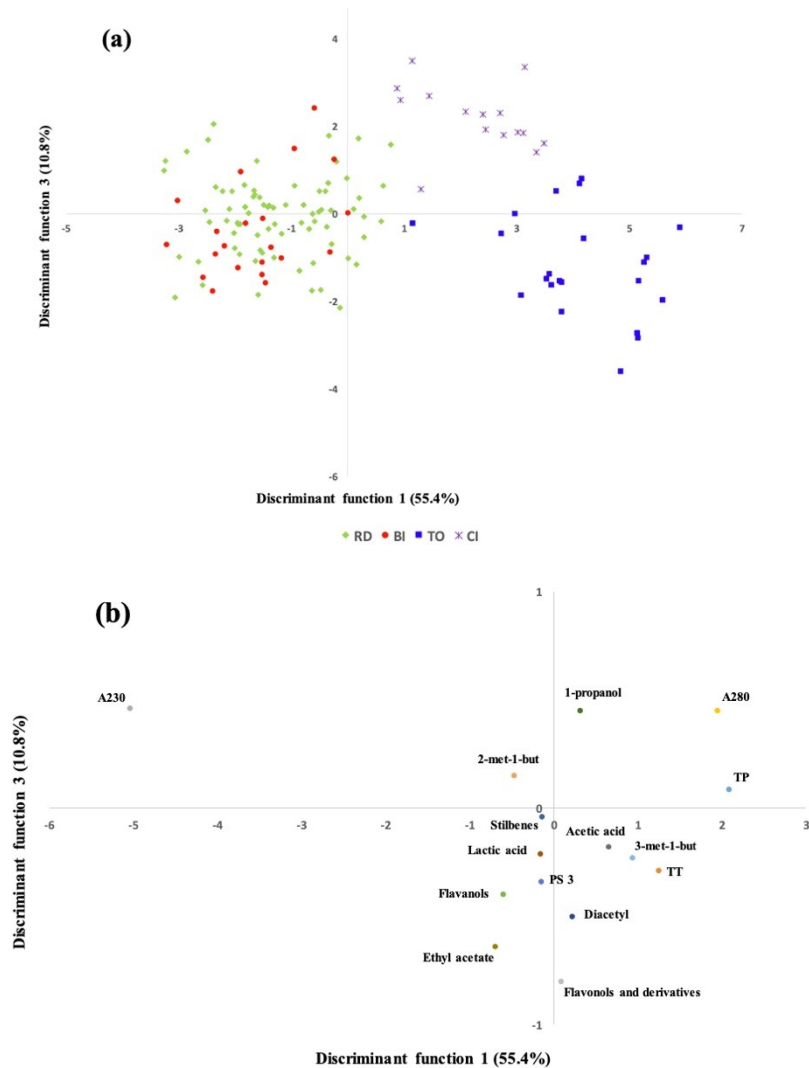
	RD	BI	TO	CI
Total polyphenols	2626 $\pm$ 321 <sup>b</sup>	2353 $\pm$ 293 <sup>a</sup>	2520 $\pm$ 324 <sup>ab</sup>	2450 $\pm$ 276 <sup>a</sup>
Total tannins	2362 $\pm$ 385.4 <sup>a</sup>	2189 $\pm$ 367 <sup>a</sup>	2566 $\pm$ 522.5 <sup>b</sup>	2394 $\pm$ 480.2 <sup>ab</sup>
A230	0.653 $\pm$ 0.09 <sup>b</sup>	0.560 $\pm$ 0.08 <sup>a</sup>	0.578 $\pm$ 0.09 <sup>a</sup>	0.579 $\pm$ 0.09 <sup>a</sup>
A280	0.173 $\pm$ 0.02 <sup>b</sup>	0.154 $\pm$ 0.02 <sup>a</sup>	0.163 $\pm$ 0.02 <sup>ab</sup>	0.158 $\pm$ 0.02 <sup>a</sup>
PS3*	153 $\pm$ 26.8 <sup>a</sup>	207 $\pm$ 46.2 <sup>b</sup>	199 $\pm$ 36.1 <sup>b</sup>	168 $\pm$ 26.9 <sup>a</sup>
Flavanols	46.9 $\pm$ 17.5 <sup>c</sup>	59.2 $\pm$ 22.6 <sup>d</sup>	36.4 $\pm$ 11.8 <sup>b</sup>	17.7 $\pm$ 6.1 <sup>a</sup>
Stilbenes	1.86 $\pm$ 0.85 <sup>b</sup>	2.84 $\pm$ 1.00 <sup>c</sup>	1.37 $\pm$ 0.50 <sup>a</sup>	1.25 $\pm$ 0.38 <sup>a</sup>
Flavonols and derivatives	40.2 $\pm$ 14.3 <sup>c</sup>	23.0 $\pm$ 9.5 <sup>b</sup>	40.7 $\pm$ 8.7 <sup>c</sup>	17.4 $\pm$ 8.2 <sup>a</sup>
Lactic acid	1.76 $\pm$ 0.43 <sup>c</sup>	1.21 $\pm$ 0.22 <sup>a</sup>	1.60 $\pm$ 0.42 <sup>bc</sup>	1.35 $\pm$ 0.25 <sup>ab</sup>
Acetic acid	0.470 $\pm$ 0.09 <sup>a</sup>	0.518 $\pm$ 0.14 <sup>ab</sup>	0.563 $\pm$ 0.17 <sup>b</sup>	0.553 $\pm$ 0.14 <sup>b</sup>
Ethyl acetate	98.2 $\pm$ 25.5 <sup>b</sup>	85.1 $\pm$ 20.9 <sup>a</sup>	73.6 $\pm$ 28.1 <sup>a</sup>	71.2 $\pm$ 36.0 <sup>a</sup>
Diacyl	2.06 $\pm$ 1.52 <sup>b</sup>	2.24 $\pm$ 1.07 <sup>b</sup>	3.87 $\pm$ 3.28 <sup>c</sup>	0.705 $\pm$ 1.96 <sup>a</sup>
1-propanol	29.2 $\pm$ 10.3 <sup>b</sup>	21.7 $\pm$ 6.4 <sup>a</sup>	28.9 $\pm$ 8.23 <sup>b</sup>	31.8 $\pm$ 8.35 <sup>b</sup>
2-methyl-1-butanol	48.6 $\pm$ 9.9 <sup>ab</sup>	51.4 $\pm$ 9.9 <sup>b</sup>	50.0 $\pm$ 9.2 <sup>ab</sup>	44.0 $\pm$ 10.7 <sup>a</sup>
3-methyl-1-butanol	186 $\pm$ 27.4 <sup>b</sup>	165 $\pm$ 23.7 <sup>a</sup>	213 $\pm$ 36.2 <sup>c</sup>	180 $\pm$ 33.2 <sup>ab</sup>

227<sup>a-c</sup>Superscript letters for each compound or parameter indicate statistically  
 228significant differences at  $p < 0.05$ .

229\*PS3: polysaccharides with a molecular weight average of 10 kDa.

230The second discriminant function, which explained the 33.8 % of total variance, made it possible to  
 231distinguish the wines from BI from the other PDOs. The physico-chemical parameters that most  
 232contributed to the separation of these wines were PS3, TT, flavanols, 2-methyl-1-butanol, A230 and  
 233stilbenes. This separation could be associated with the grape variety used, because the red wines from  
 234BI were elaborated with cv. *Mencía* and those from the rest PDOs with cv. *Tempranillo*. The content of  
 235PS3 was significantly higher in the wines from BI than in the others, with the exception of wines from  
 236TO where the PS3 content was similar. Conversely, the BI wines showed the highest content of  
 237flavanols and stilbenes. Low differences were found in the other variables.





238

239 **Figure 2.** Plot of scores (a) and loadings (b) for PDO wine differentiation using  
 240 discriminant functions 1 and 3.

241 TP, total polyphenols; TT, total tannins; PS3 (polysaccharides with a molecular  
 242 weight average of 10 kDa) 2-met-1-but, 2-methyl-1-butanol; 3-met-1-but, 3-  
 243 methyl-1-butanol.

244

245 Figures 2a and 2b show the scores and loadings defined in the first and the third discriminant functions  
 246 in the plane and explained 66.2 % of the total variance. The third discriminant function explained 10.8  
 247 % total variance, and allowed a good separation between the wines from TO and CI. The wines from  
 248 CI were located at the top of the plane, and the physico-chemical variables that most contributed to  
 249 their separation, from highest to lowest weight of their loadings, were A230, A280, 1-propanol, 2-  
 250 methyl-1-butanol and TP. The rest of the physico-chemical variables were most associated with the  
 251 separation of the wines from TO, which were located at the bottom of the plane, and the variables with  
 252 the highest loading weights were flavonols and their derivatives, ethyl acetate, diacetyl, flavanols and  
 253 PS3.

254 The classification matrix of the model indicated that, in total, 97.8 % of the wines studied were  
 255 correctly classified using the variables selected by the SLDA model: 98.7 % of the wines from RD,  
 256 95.2 % from BI, 95.5 % from TO and 100 % from CI. The wines that were misclassified might be due

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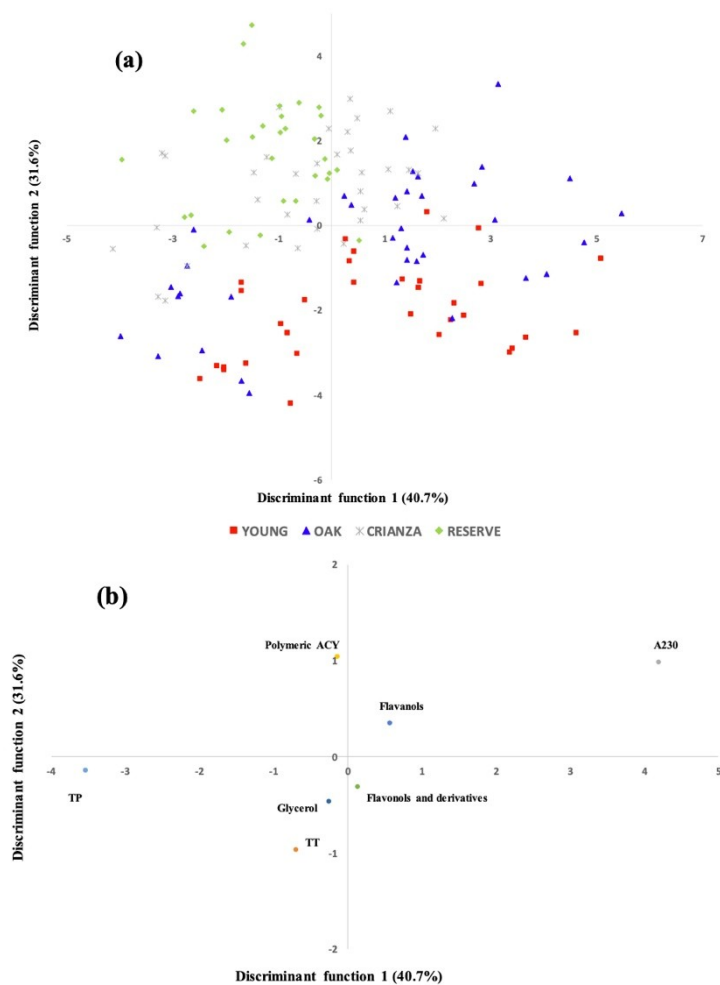
257to the geographical proximity between PDOs, as described in a study by Pérez-Magariño and  
258González-San José (2001) with wines from different Spanish PDOs. The validation of the model using  
259four cross-validation steps showed that 91.7 % of the wines were correctly classified. Therefore, the  
260model can be considered good and stable.

261The results obtained in our study are in agreement with other studies carried out with wines from  
262different regions, which some of them were close geographically but others not, with different  
263enviromental conditions and with different grape varieties. The phenolic composition is the most  
264useful for characterizing and discriminating wines from different regions. However, the phenolic  
265compounds selected for discriminating the wines were different in each study. Thus, Buscema and  
266Boulton (2015) reported good differentiation in *Malbec* wines from four Mendoza regions, selecting  
267different phenolic compounds that included anthocyanins and non-anthocyanin compounds. Another  
268study carried out by Rodríguez-Delgado *et al.* (2002) showed good differentiation of wines from  
269different production areas in the Canary Islands (Spain) using five non-anthocyanin phenolic  
270compounds. Peña-Neira *et al.* (2000) discriminated wines from four Spanish PDOs (La Mancha,  
271Valdepeñas, Rioja and Cariñena) using their phenolic composition, mainly related to some  
272hydroxybenzoic (gallic acid) and hydroxycinnamic (caffeic acid) acids and phenolic alcohols (tyrosol).  
273Di Paola-Naranjo *et al.* (2011) obtained good classifications by variety and the origin of wines from  
274three provinces of Argentina, using the phenolic profile and other compounds. They concluded that  
275*trans*-resveratrol was the phenolic compound that discriminated between these three regions. Another  
276study showed the important role of phenolic compounds (anthocyanins and non-anthocyanins) in the  
277classification of wines from three Spanish PDOs (Penedes, Rioja and Ribera del Duero) (Serrano-  
278Lourido *et al.*, 2012). However, they concluded that the discrimination of wines from Rioja and Ribera  
279del Duero was more difficult, probably because production areas are geographically quite close. In  
280another study carried out by Pérez-Magariño and González-San José (2001), the wines from five  
281different Spanish PDOs (Ribera del Duero, Rioja, Valdepeñas, La Mancha and Madrid) were correctly  
282differentiated, with anthocyanic pigments as the most discriminant variables.

283As mentioned above, the content of polysaccharide with low molecular weight (PS3) also contributed  
284to the separation of the wines according to the PDO. Cejudo-Bastante *et al.* (2018) also showed an  
285effect of the polysaccharide composition in *Carignan* red wines from six different areas in Chile: they  
286found that polysaccharide fractions with a high molecular weight had a greater influence on  
287differentiating the wines from these areas than polysaccharide fractions with a low molecular weight.

288In our study, several higher alcohols contributed to the separation of the wines by PDO. Sagratini *et al.*  
289(2012) found that 3-methyl-1-butanol (one isoamyl alcohol) was one of the most discriminant  
290compounds (together with ethyl decanoate and ethyl octanoate) for separating *Montepulciano* red  
291wines from two different Italian regions.

292Another SLDA was carried out to discriminate the wines according to their category, using the same  
29340 physico-chemical variables. In this case, the best model selected seven physico-chemical variables  
294that most contributed to differentiating the wines according to the category. The variables selected  
295were as follows, from highest to lowest discriminant power: polymeric ACY, glycerol, flavanols,  
296flavonols and their derivatives, A230, TP and TT. The final model selected three discriminant  
297functions that explained 83.1 % of the total variance: the first discriminant function explained 40.7 %,  
298the second 31.6 %, and the third 10.8 %.



299  
 300 **Figure 3.** Plot of scores (a) and loadings (b) for wine category differentiation  
 301 using discriminant functions 1 and 2.  
 302 TP, total polyphenols; TT, total tannins; Polymeric ACY (polymeric anthocyanins).  
 303

304 Figures 3a and 3b show the scores and loadings in the plane of the two first discriminant functions.  
 305 Only a good separation between “young” and “reserve” wines was observed, mainly associated with  
 306 the second discriminant function. In general, the “reserve” wines were located at the top of the plane  
 307 and the “young” wines at the bottom. According to the weight of the physico-chemical variables in  
 308 this discriminant function, polymeric ACY, A230 and flavanols were positively correlated with the  
 309 separation of “reserve” wines. Table 3 shows that the percentage of polymeric ACY increased with  
 310 aging of the wines: the longer the aging of the wines, the higher the content of polymeric ACY. This  
 311 result was expected and is in agreement with other studies reported in the literature (Burin *et al.*, 2011;  
 312 Chira *et al.*, 2011; del Barrio-Galán *et al.*, 2011; McRae *et al.*, 2012; Dipalmo *et al.*, 2016; Agazzi *et*  
 313 *al.*, 2018). These compounds are mainly formed during the aging of wines (in oak barrel and/or bottle)  
 314 because chemical reactions between the monomeric anthocyanins and other phenolic compounds and  
 315 metabolites (Monagas *et al.*, 2005b; De Rosso *et al.*, 2009) having an important role on the long-term  
 316 color stability of aged red wines (Boulton, 2001). The value of A230 was significantly higher in the  
 317 aged wines than in the “young” wines, and could be proposed as an indicator of wines that have  
 318 experienced some type of aging.

319Table 3. Mean values  $\pm$  standard deviation of the variables selected by the SLDA  
320for wine category discrimination.

	Young	Oak	Crianza	Reserve
Total polyphenols	2294 $\pm$ 255 <sup>a</sup>	2645 $\pm$ 352 <sup>b</sup>	2608 $\pm$ 290 <sup>b</sup>	2616 $\pm$ 271 <sup>b</sup>
Total tannins	2129 $\pm$ 356 <sup>a</sup>	2512 $\pm$ 470 <sup>b</sup>	2360 $\pm$ 361 <sup>b</sup>	2474 $\pm$ 424 <sup>b</sup>
A230	0.550 $\pm$ 0.08 <sup>a</sup>	0.653 $\pm$ 0.11 <sup>c</sup>	0.626 $\pm$ 0.09 <sup>b</sup>	0.634 $\pm$ 0.08 <sup>b</sup>
% polymeric ACY	40.5 $\pm$ 6.21 <sup>a</sup>	49.9 $\pm$ 9.56 <sup>b</sup>	56.9 $\pm$ 7.57 <sup>c</sup>	78.7 $\pm$ 8.76 <sup>d</sup>
Flavanols	51.9 $\pm$ 21.0 <sup>b</sup>	49.7 $\pm$ 21.4 <sup>b</sup>	37.9 $\pm$ 16.3 <sup>a</sup>	29.2 $\pm$ 10.8 <sup>a</sup>
Flavonols and derivatives	40.4 $\pm$ 14.9 <sup>b</sup>	40.8 $\pm$ 12.3 <sup>b</sup>	28.6 $\pm$ 14.9 <sup>a</sup>	29.3 $\pm$ 14.1 <sup>a</sup>
Glycerol	8.89 $\pm$ 1.34 <sup>b</sup>	8.41 $\pm$ 1.78 <sup>ab</sup>	7.81 $\pm$ 1.19 <sup>a</sup>	8.71 $\pm$ 1.14 <sup>b</sup>

321<sup>a-d</sup>Superscript letters for each compound or parameter indicate statistically  
322significant differences at  $p < 0.05$ .

323Conversely, the variables at the bottom of the plane (TT, glycerol, flavonols and their derivatives and  
324TP) allowed the separation of “young” wines, and the TT and flavonols and their derivatives were the  
325most significant variables. The “young” wines had significantly lower content of TT and TP than the  
326aged wines, possibly because, in general, the winemakers selected wines with a high phenolic content  
327to be aged for a longer time. However, the “young” and the “oak” wines presented higher content of  
328flavanols and flavonols and their derivatives than the wines aged for a longer time.

329The classification matrix reported by the SLDA model classified correctly 80 % of wines according to  
330their category. As was previously presented, the best separation was found between “reserve” and  
331“young” wines, as these wines showed the best classification percentages (90 % for “reserve” and 87.5  
332% for “young”). Next, 80.6 % of “crianza” wines were classified correctly. Finally, “oak” wines  
333presented the lowest classification percentage at 64.9 %, as these wines were confused with “young”  
334or “crianza” wines. This result was expected because the aging time of “oak” wines is less stringent  
335than “crianza” and “reserve”, which must comply with a minimum aging time.

336According to the cross-validation, the percentage of the wines classified correctly was lower but  
337acceptable for “reserve” and “young” wines (83 % and 81 %, respectively). However, the results for  
338“crianza” and “oak” wines were poorer (70.5 % and 55 %, respectively).

339Various studies have evaluated the discriminant power of several physico-chemical variables between  
340young and aged wines. Agazzi *et al.* (2018) discriminated *Malbec* red wines from Mendoza and  
341California at the beginning of aging and after five years. They observed that aging time affects  
342significantly the total polyphenols and total monomeric anthocyanins, flavanols, flavonols, and  
343hydroxycinnamic acids content. The aged wines presented higher concentrations of *p*-coumaric acid  
344and lower concentrations of monomeric anthocyanins than young wines. Dipalmo *et al.* (2016)  
345observed that red wines from the *Primitivo* grape variety aged for two years showed an increase of  
346polymeric anthocyanins (pyranoanthocyanins). Finally, Astray *et al.* (2019) determined the aging time  
347of red wines from PDO Toro using classical enological parameters and total polyphenol index.

## 348Conclusions

349Differences in physico-chemical composition were found in red wines from  
350different Spanish PDOs and different categories. Phenolic composition and  
351absorbance values associated with total phenolic content allowed good  
352discrimination and classification of wines from different PDOs. It was posible to

353discriminate the wines mainly elaborated with the *Tempranillo* grape variety (RD,  
354TO and CI) and those elaborated with the *Mencía* grape variety (BI), and the  
355absorbance values and phenolic groups such as TP and TT were the most  
356important variables for this differentiation. Other variables such as  
357polysaccharides with low molecular weight, isoamyl alcohols and low molecular  
358weight phenolic compounds (mainly flavanols and flavonols), were relatively  
359important in this differentiation, mainly associated with those wines elaborated  
360with the *Mencía* grape variety.

361Although there are clear differences between the wines elaborated with different  
362grape varieties, the winemaking techniques used in each region, and even in  
363each winery, could also have an effect.

364The discrimination of wines according to their category was more difficult, and  
365only an acceptable classification of “young” wines and those with longer aging  
366periods (“crianza” and “reserve”) was achieved. The percentage of polymeric ACY  
367and A230 could be proposed as good indicators for aged wines, and TT for young  
368wines.

369Further studies should be carried out that analyse other physico-chemical  
370variables that could differentiate the red wines by their origin and/or category,  
371such as volatile compounds, and sensory attributes.

## 372Acknowledgements

373This study was supported by a project 2017/721 from “Rural Development  
374Program (PDR) of Castilla y León 2014-2020” (financed with FEADER funds). The  
375authors are grateful to the different Regulatory Councils for providing the wines  
376for the study.

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