



REVIEW ARTICLE

Grape, wine and pomace anthocyanins: winemaking biochemical transformations, application and potential benefits

Katarina Delić^{1,2,3}, Danijel D. Milinčić¹, Mirjana B. Pešić¹, Steva Lević², Viktor A. Nedović², Anne-Laure Gancel³, Michael Jourdes³, Pierre-Louis Teissedre^{3,*}

¹ Laboratory of Food Chemistry and Biochemistry, Institute of Food Technology and Biochemistry, Faculty of Agriculture, University of Belgrade, Nemanjina 6, 11080 Belgrade, Serbia.

² Laboratory of Food Biotechnology, Institute of Food Technology and Biochemistry, Faculty of Agriculture, University of Belgrade, Nemanjina 6, 11080 Belgrade, Serbia.

³ Université de Bordeaux, Bordeaux INP, Bordeaux Sciences Agro, UMR 1366 OENOLOGIE, ISW, 33140 Villenave-d'Ornon, France.



*correspondence:

pierre-louis.teissedre@u-bordeaux.fr

Associate editor:

Luca Rolle



Received:

6 March 2024

Accepted:

26 September 2024

Published:

26 November 2024

ABSTRACT

Anthocyanins are polyphenols found in red grapes, wines and their by-products. This scientific paper reviews their physiological significance in grape tissues, the biochemical transformations they undergo during winemaking and wine aging, their potential application in the food industry, and the health benefits associated with them. The paper sheds light on the factors influencing their extraction, stability and composition in wines, and explores their applications in various food products and the possibility of sustainable winemaking using pomace. The paper focuses on the broad impact of anthocyanins on product quality and consumer preferences, and highlights potential preventive and therapeutic applications for human health conditions. Overall, this comprehensive overview provides valuable insights into the multifaceted roles of anthocyanins, paving the way to future research on the application of anthocyanin in agriculture, food science and medicine.

KEYWORDS: grape, wine, anthocyanins, polyphenols, winemaking, biochemical transformations, application, health benefits



This article is published under the **Creative Commons licence** (CC BY 4.0).

Use of all or part of the content of this article must mention the authors, the year of publication, the title, the name of the journal, the volume, the pages and the DOI in compliance with the information given above.

INTRODUCTION

Since ancient times, grape growing has been closely intertwined with human culture, making it one of the world's oldest and most significant agricultural traditions. The cultivation of grapes and the production of wine have played pivotal roles in various civilisations, spanning continents and centuries. Till now, a total of about 10,000 grapevine varieties are known according to OIV reports (International Organisation of Vine and Wine); however, approximately 50 % of the vineyard area worldwide contains 33 grapevine varieties (OIV, 2017). These findings underscore the significant dominance and widespread cultivation of a relatively small number of grapevine varieties (known as “international varieties”) on a global scale (OIV, 2017). On the other hand, local indigenous varieties are cultivated in specific regions of numerous winemaking countries. These local, autochthonous or native varieties are currently increasing in importance for the wine industry due to their enhanced adaptability to the escalating challenges posed by climate change (Otto *et al.*, 2023). A large percentage of global grape production is dedicated to wine production; the wine industry plays a major role in world economy with global grape production estimated to be 7.2 million hectares in 2023, and global wine production to be 237 million hectoliters (OIV, 2024). As a result of numerous chemical and biochemical reactions and mechanisms, wine production is a synergy of several different elements, such as grape variety, terroir and winemaking techniques (Razungles, 2022).

The quality of red wine is directly proportional to its content in aromatic compounds, proanthocyanins, and anthocyanins. Moreover, the colour of red wine is one of the prime sensory characteristics in which anthocyanins have a crucial role (Paissoni *et al.*, 2018). Anthocyanins are natural water-soluble pigments that are found in a wide range of plants and appear red, blue or purple (Winefield *et al.*, 2009). Grapevine anthocyanins have an important role in winemaking, since they are responsible for the red colour of wine. The stimulation of anthocyanin molecules by visible light influences their colours (Li *et al.*, 2021). Anthocyanins also have a special physiological role in the grapevine linked to, for example, free radical scavenging and antioxidative capacity, defence against pathogens, seed dispersal, and the proposed modulation of signaling cascades (He *et al.*, 2010) which has to date not been thoroughly explained and requires further investigation (Roubelakis-Angelakis, 2009). Anthocyanins are found in the skin of the black and purple varieties of grape berries; white grape varieties do not usually synthesise anthocyanins, although some studies have shown that synthesis can occur in white berries due to mutations of the genes that are responsible for the synthesis of anthocyanins (He *et al.*, 2010; Ivanova *et al.*, 2011a). ‘Teinturier’ grape varieties are known as dyer grapes, because anthocyanins are contained in the flesh of their berries (Körösi *et al.*, 2022). Biochemically, anthocyanins are highly reactive and unstable bioactive compounds, which undergo degradative changes, depending on, for example,

the pH of the medium, temperature, light, presence of oxygen, presence of dry matter and water content (Moreno-Arribas & Polo, 2009). Due to their structure and instability, anthocyanins undergo transformation processes during red winemaking, with changes to their chemical structure and various complexes being formed with other compounds (sugars, organic acids, phenolic compounds and halogens). Anthocyanins often exhibit positive effects on human health (Zia ul Haq *et al.*, 2016); as a result, their presence in winemaking waste (*i.e.*, pomace and its constituents) is being increasingly valorised and they are further used in different food industry sectors (García-Lomillo & González-SanJosé, 2017). For example, different types of encapsulations have been developed to increase the use of anthocyanins in food technology, as well as their stability and bioavailability (Zuidam & Nedovic, 2010; Popović *et al.*, 2019; Milinčić *et al.*, 2019; Milinčić *et al.*, 2022b).

This review manuscript aims to consolidate all the research related to grape anthocyanins in several subsections, which address the 1) behaviour of anthocyanins during physiological processes in grapevine, 2) presence of anthocyanins in different international and indigenous black grape varieties, 3) biochemical transformations of anthocyanins during the winemaking process and wine aging, 4) effect of anthocyanins on wine sensory properties, 5) valorisation of anthocyanins in winemaking waste and further applications in the food industry, and 6) health benefits of anthocyanins and *in vivo/in vitro* transformation after their consumption.

OVERVIEW AND CHEMISTRY OF ANTHOCYANINS

Anthocyanins are the most important group of water-soluble plant pigments. Plant pigments have different roles, such as attracting pollinators and fruit dispersers, tracking the time of the day and photoprotection. In epidermal plant cells, anthocyanins are accumulated in vacuoles (Li *et al.*, 2021; Crang *et al.*, 2018). The anatomy, morphology and structure of the berries and clusters are key parameters in ampelography for grapevine variety distinction. The development and composition of mature berries and anthocyanin transformation is mostly genetically influenced, but environmental conditions may also have an influence (Ribéreau-Gayon *et al.*, 2006).

In chemical and biochemical terms, anthocyanins are bioactive compounds belonging to polyphenols, and more specifically to a subclass of secondary metabolites known as flavonoids (Li *et al.*, 2021). Anthocyanins have amphoteric properties due to their structure. They change colour with a change in pH of the medium. The flavylium cation consists of two benzene rings (A, B) connected by an unsaturated cationic oxygenated heterocycle (C), which originates from the 2-phenyl-benzopyrylium nucleus, thus giving anthocyanins amphoteric properties (Ribéreau-Gayon *et al.*, 2006). Anthocyanins exist in many different forms, which affect their colour and stability. Glycosylation of anthocyanidins usually occurs at the C₃ position of the 2-phenyl-benzopyrylium nucleus (C),

giving 3-monoglucoside. In the case of glycosylation with the second sugar, it is mostly a 5-hydroxyl position. The most common sugar is glucose, but there can also be other monosaccharides, such as galactose, arabinose and xylose (Vermerris & Nicholson, 2006). Furthermore, acylation also increases anthocyanin stability and occurs at the C₆ position of the sugar moieties by esterification with acetic, *p*-coumaric and caffeic acids (Moreno-Arribas & Polo, 2009).

1. Anthocyanins-biosynthesis

Anthocyanins are synthesised *via* the flavonoid pathways (He *et al.*, 2010), encoded by two groups of genes: structural and regulatory. The biosynthesis of anthocyanins is intersected by many different products that act as precursors for other related compounds, involving a considerable number of enzymes (Ivanova *et al.*, 2011b) [Figure 1].

Enzymes that are responsible for anthocyanin biosynthesis are encoded by structural genes (Yang *et al.*, 2023). Regulatory genes, also known as transcription factors, control the expression of structural genes, regulating the synthesis of anthocyanins (Holton & Cornish, 1995; Moreno-Arribas & Polo, 2009). Anthocyanin synthesis occurs during the phenological ripening phase, and is controlled by the VvMYBA1 anthocyanin biosynthesis transcription factor, which controls the expression of anthocyanin-specific biosynthetic gene 3-O-glucosyltransferase [UGFT] (Cutanda-Perez *et al.*, 2009). Anthocyanin accumulation occurs in most cases after veraison, but recent research has shown that it depends largely on the grapevine variety (Wang *et al.*, 2024). Other than genetic factors, environmental factors, such as low temperature and light, also affect gene expression during anthocyanin biosynthesis (Azuma, 2018). However, some research has shown that anthocyanin

biosynthesis may not be dependent on light, since, in some varieties, flesh coloration occurred after skin coloration (Lu *et al.*, 2023). Recent studies have given new insights into the metabolic pathway and biosynthesis of anthocyanins, and the genes involved. Some have suggested that the pathways of anthocyanin components have large effects on different levels of grape berry development (Zia ul Haq *et al.*, 2016). This implies that some levels of development of the grape berry may, to a greater or lesser extent, depend on anthocyanin precursor compounds and the genes and enzymes responsible for their biosynthesis. Regarding ‘teinturiers’, these varieties have shown a high tendency to direct the methylation of cyanidin but a low readiness for its hydroxylation (Papoušková *et al.*, 2011). Furthermore, it has recently been proven that anthocyanin biosynthesis in grape flesh is independent of anthocyanin biosynthesis in the skin (Lu *et al.*, 2023). These investigations were carried out on new teinturier varieties, such as ‘ZhongShan-HongYu’ (Yang *et al.*, 2023), ‘Mio Red’ (Lu *et al.*, 2023) and the medicinal *Vitis vinifera* L. variety ‘SuoSuo’ (Wang *et al.*, 2023).

Immediately after synthesis, the aglycone forms of anthocyanins, anthocyanidins (Figure 2A), are unstable and undergo modifications, such as glycosylation, methylation and acylation (He *et al.*, 2010). Glycosylation increases the hydrophilicity and stability of anthocyanidins, resulting in their transformation into anthocyanins. In *Vitis vinifera* L. varieties, O-glycosylated anthocyanins are only present at the C3 position, with methylation of the hydroxyl groups occurring at C3’ or both C3’ and C5’ on the B rings of the anthocyanins (Grotewold, 2006) [Figure 2B]. Acylation is a modification that occurs on the C6” positions of the

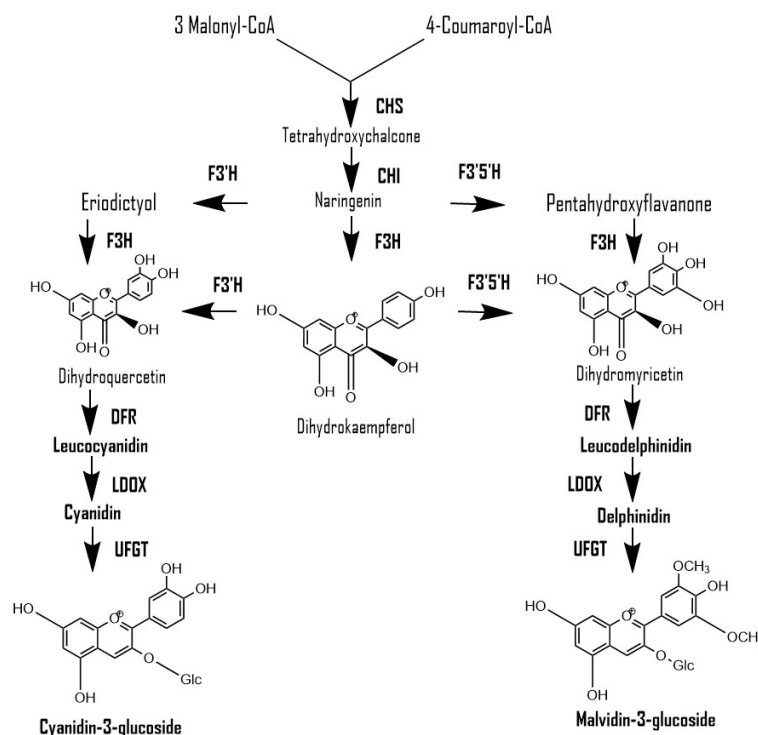


FIGURE 1. Biosynthesis pathway of anthocyanins in red grapes.

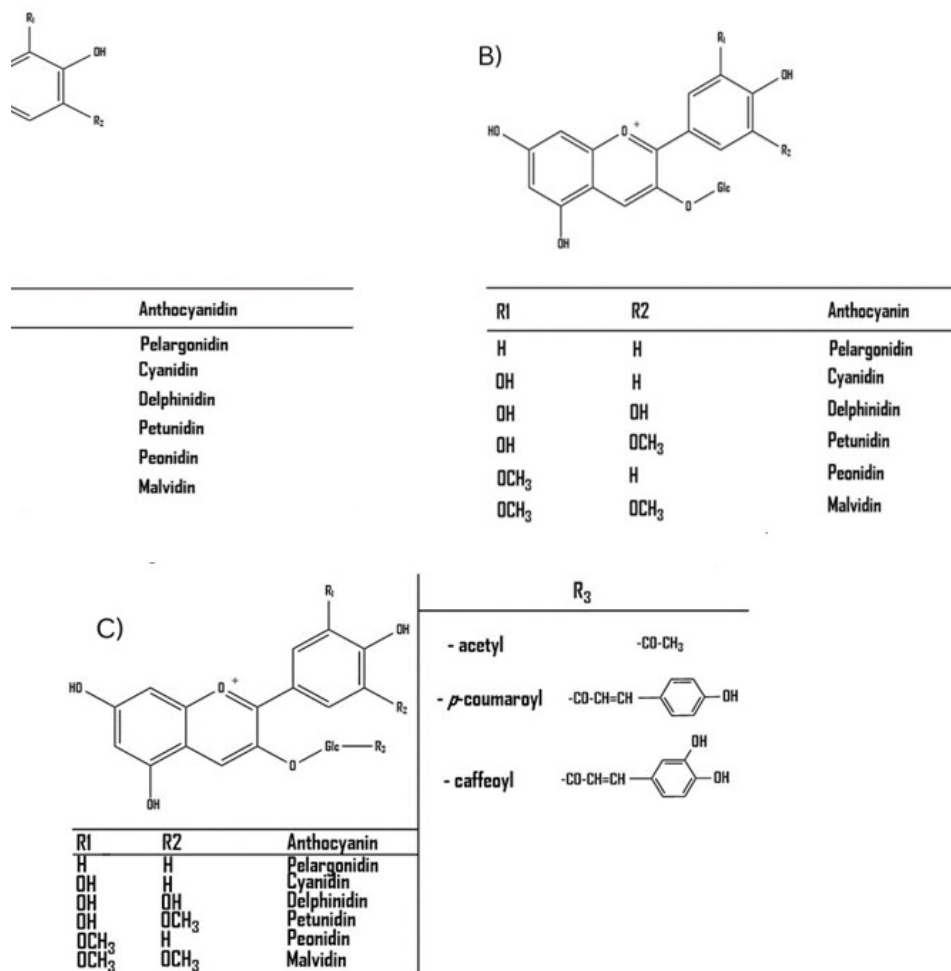


FIGURE 2. Structural formulas of A) Anthocyanidins, B) Anthocyanins, and C) Acylated anthocyanins.

glycosyl groups, involving the addition of the aromatic and/or aliphatic group (He *et al.*, 2010; Holton & Cornish, 1995; Grotewold, 2006) [Figure 2C]. All these modifications increase the stability of anthocyanins, which is particularly important during intracellular transport. Because anthocyanins are unstable in the cytosol, this is where modifications mostly occur (Wang *et al.*, 2024).

The biosynthesis of anthocyanins occurs in the endoplasmic reticulum (Alfenito *et al.*, 1998), after which they are transported to the vacuole (Grotewold, 2004). Two types of anthocyanin transport in the cells take place: ligandin transport (LT) and vesicular transport [VT] (Zhao & Dixon, 2010). The sequestration of anthocyanins inside the vacuoles is the last step of anthocyanin accumulation, where their colour will depend not only on their molecular structure but also on the vacuolar pH and the concentration of metal ions and co-pigments (Grotewold, 2006). These kinds of insights provide valuable information regarding the molecular structure and bioavailability of anthocyanins.

2. Anthocyanins in red grapes

The mature red grape berry has a complex anatomical structure characterised by distinct tissues and structures within the berry and the cluster. The outermost layer of the berry, known

as the exocarp or skin, is composed of multiple layers of epidermal cells rich in anthocyanins (Figure 3A). Under the exocarp is mesocarp or flesh, comprising parenchyma cells that store sugars, organic acids and water, which contribute to the berry's juiciness and flavour. The berries of teinturier varieties are morphologically different to other varieties, as their flesh is red and their mesocarp contains anthocyanins (Körösi *et al.*, 2022) [Figure 3B].

The exact role of anthocyanins in grapevine is still not clear (Roubelakis-Angelakis, 2001). Other than dispersing seed and being potential UV-protectants, red and black grape anthocyanins are responsible for the colour of the wine, which, their first most important role. The concentration, composition and distribution of anthocyanins and their derivatives significantly vary among red grape varieties of *Vitis vinifera* L. Moreover, these characteristics also differ among other grape north American and Asian species, including *Vitis labrusca*, *Vitis rotundifolia* and *Vitis amurensis*, and hybrid varieties resulting from crossing of *Vitis vinifera* L. cultivars with other species (Teissedre, 2018).

Much research has been conducted to identify and quantify the polyphenolic compounds of red grape varieties, including both *vinifera* and non-*vinifera* varieties. Important

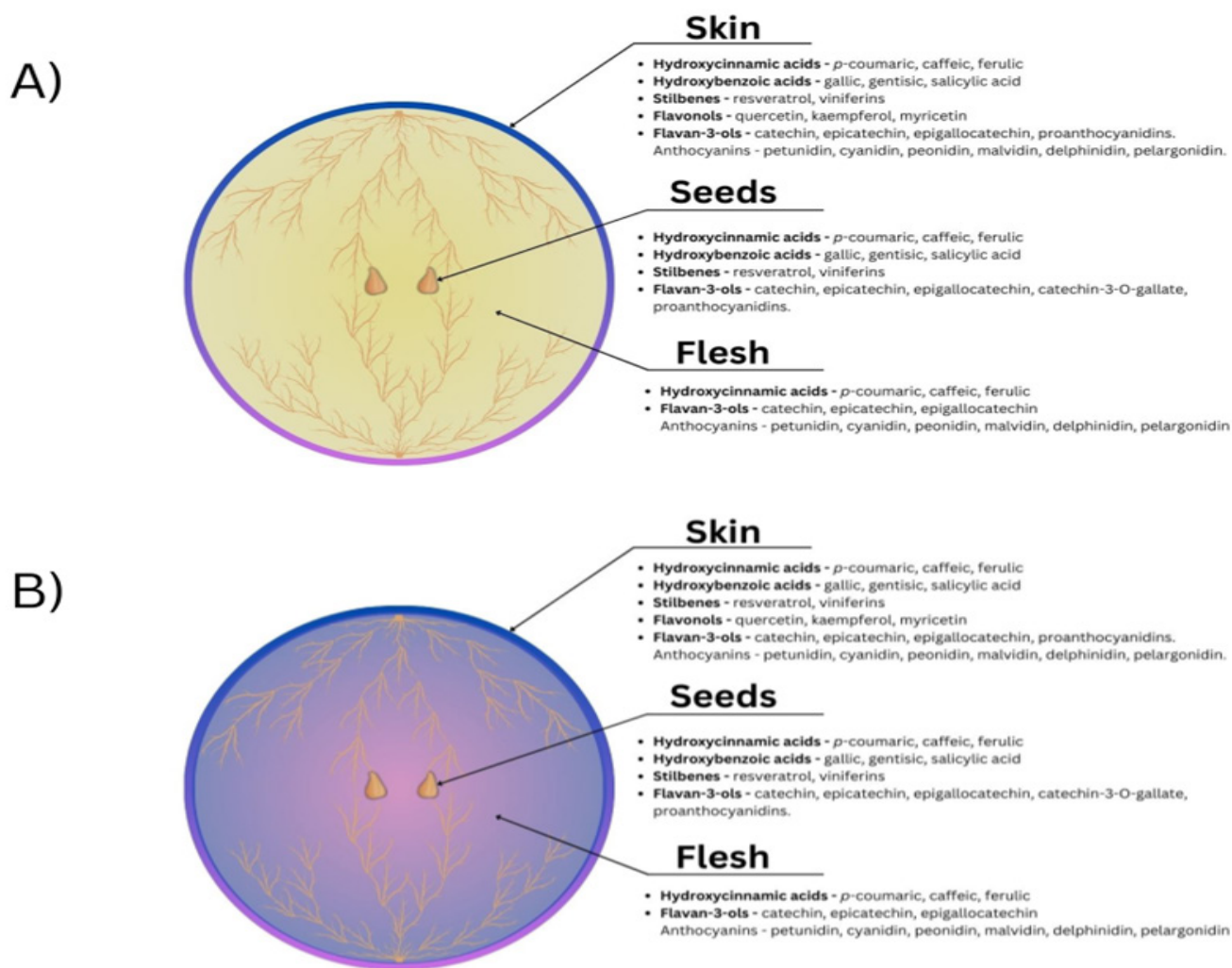


FIGURE 3. Anatomy of the red grapevine berry: A) varieties without anthocyanins in the mesocarp, and B) ‘teinturier’ varieties.

differences in their composition regarding anthocyanins have been reported, and they also vary in concentration. According to Lamikanra (1989), *Vitis vinifera* L. varieties contain mainly acylated and non-acylated anthocyanins, *Vitis rotundifolia* and its hybrids have non-acylated 3,5-*O*-diglucosides, and *Vitis labrusca* cultivars have a mixture of acylated and non-acylated mono- and di-glucosides of anthocyanidins, as previously reported by many studies (Ehrhardt *et al.*, 2014; Zhu *et al.*, 2012; Wojdyło *et al.*, 2018; Tassoni *et al.*, 2019; Milinčić *et al.*, 2021a; Forino *et al.*, 2022; Tampaktsi *et al.*, 2023; Milinčić *et al.*, 2023b).

The most analysed anthocyanin profiles are those of the French international varieties ‘Cabernet-Sauvignon’ and ‘Merlot’ (Chira *et al.*, 2011; Chira *et al.*, 2012; Zhao *et al.*, 2023), which have been investigated and compared in every winegrowing region worldwide. Many results have shown that ‘Cabernet-Sauvignon’ and ‘Merlot’ are predominant in malvidin-3-*O*-glucoside, which show potential for chemotaxonomic purposes (Garcia-Beneytez *et al.*, 2003). In addition, they are rich in acetylglucosides, with a 1.5- to 3-fold higher concentration of acetyl anthocyanins in ‘Cabernet-Sauvignon’ than in ‘Merlot’ (Lorrain *et al.*, 2011). Cinnamoyl derivatives

(*p*-coumaroylglucosides and caffeoylglucosides) have also been identified in these two varieties (Costa *et al.*, 2014). Other *V. vinifera* L. varieties have been investigated. Garcia-Beneytez *et al.* (2003) worked on the different red grapevine varieties mainly grown in Spain, namely Alicante Bouschet, Bobal, Carinena, Crujidera, Garnacha Peluda, Monastrell, Moristel, Morrastel-Bouschet, petit Bouschet, Prieto Picudo, Tempranillo and Vitidillo. HPLC-MS identification of anthocyanins in grape extracts has been done and the presence of 3-*O*-glucoside derivatives, acetyl glucosides, and cinnamoyl derivatives of delphinidin, cyanidin, petunidin, peonidin, and malvidin has been confirmed. For most of these varieties, malvidin-3-*O*-glucoside was the major anthocyanin, except for the teinturier varieties Alicante Bouschet, Morrastel Bouschet and petit Bouschet, in which the main anthocyanin was peonidin-3-*O*-glucoside, which is specific to teinturier cultivars. Portuguese varieties showed similar results in the research done by Costa *et al.* (2014). They investigated Portuguese *V. vinifera* L. varieties (Camarate, Monvedro, Moreto Boal, Negro Mole, Negro Mouro, Alfrocheiro, Alvarilhao, Bastardo, Cornifesto, Jean, Malvasia Petra, Rufete, Sausao, Tinta Amarela, Tinta Barca,

Tinta Barroca, Tinta Miuda and Tinto Cao) in comparison to some of the international varieties (Gewurztraminer, Aramon, Cabernet franc, Carignan noir, Gamay and Grenache). The main individual anthocyanin in the skin composition of these varieties was malvidin-3-*O*-glucoside, followed by the cinnamoyl derivative, malvidin-3-*p*-coumaroyl glucoside. These slight differences in composition and concentration of the most predominant individual anthocyanins in grape skin of *V. vinifera* L. cultivars are thought to be genetically determined, which has been confirmed by other authors (Obreque-Slier *et al.*, 2013; Perez-Navarro *et al.*, 2019; Guerrero *et al.*, 2009; Arozarena *et al.*, 2002; Lingua *et al.*, 2016; Sikuten *et al.*, 2021; Ivanova *et al.*, 2011a; Šuković *et al.*, 2020; Đorđević *et al.*, 2018; Milinčić *et al.*, 2021a; Ćirković *et al.*, 2022; Lakićević *et al.*, 2022; Milinčić *et al.*, 2023b). Table 1 lists the phenolic compounds found in different grape skins and their concentrations in mg/kg dry matter (DM).

Non-*vinifera* cultivars (hybrids or PIWI varieties), which are resistant to most grapevine pathogenic fungi and phylloxera, represent the future for sustainable viticulture and winemaking. They are known for having different phenolic compositions to the *V. vinifera* L. varieties. The potential significance of this fact is reflected in the need to increase the stability of the colour of grape juice and wine, due to the increasing industrial demand for natural colorants. Research carried out to date has resulted in the detection and identification of anthocyanins that are not specific to *V. vinifera* L., namely anthocyanidin-3,5-diglucosides (Lamikanra, 1989). Although some researchers have detected anthocyanidin-3,5-diglucosides in certain *V. vinifera* L. varieties (Pantelić *et al.*, 2016; Perez-Navarro *et al.*, 2019), they have been found to be different to those identified in non-*vinifera* cultivars and present in non-measurable concentrations. Through recent research on the different *Vitis amurensis* cultivars and hybrids, Zhu *et al.* (2021) discovered a new type of anthocyanin, which is believed to be 3,5,7-*O*-triglucosides. They also report the results of the anthocyanin composition of *V. amurensis* to show a high content of diglucoside anthocyanins and a low content of acylated anthocyanins, which is consistent with previous research (Zhao *et al.*, 2010). For further valorisation of the potential usage of non-*vinifera* cultivars in winemaking, and their contribution to human health, the red grape varieties ‘Rondo,’ ‘Regent,’ and ‘Cabernet-Cortis’ were investigated by different authors. Wojdylo *et al.* (2018) detected in ‘Rondo’ and ‘Regent’ the presence of 3,5-diglucosides of delphinidin, cyanidin, petunidin, peonidin and malvidin. Meanwhile, Ehrhardt *et al.* (2014) reported the presence of pelargonidin-3,5-diglucoside as well in both ‘Cabernet-Cortis’ and ‘Regent.’, which is mostly found in traces, and could therefore be an important distinguishing marker of the anthocyanin profiles of PIWIs. On a technological level, 3,5-diglucosides are largely preferable in winemaking and superior to monoglucosides, since they show greater stability to heat and light (Lamikanra, 1989); therefore, hybrids are quickly finding their place in grape juice production and winemaking.

ANTHOCYANINS IN RED WINE

Red winemaking is a complex process that involves the transformation of red grapes into red wine through a series of biochemical reactions. The production of red wine can be divided into several key stages, including harvesting and crushing the grapes, maceration, fermentation, aging and bottling. Each step contributes to the development of the wine’s colour, flavour and aroma. The most vital step is maceration, involving the contact of grape skins, seeds and pulp with the fermenting juice for a certain amount of time. This contact facilitates the extraction of anthocyanins, tannins and flavour compounds from the grape solids into the wine. The duration of maceration can vary depending on winemaker preference and grape variety. Anthocyanins are released into the wine, contributing to its characteristic red hue, while tannins affect its structure, astringency and aging potential (Razungles, 2022). As a result of fermentation, maceration, aging, and bottling, the wine develops its characteristic colour, flavour and aroma profile. The interaction between grape solids, phenolic compounds (anthocyanins), yeast and various winemaking techniques contributes to the complexity and diversity of red wines which influence their phenolic composition (Table 1). The anthocyanin composition of red wine is subject to various factors, primarily determined by the concentration and specific content of anthocyanins present in grape varieties. However, additional factors, such as environmental conditions, and viticultural and agrotechnical practices employed in the vineyard, can also impact the anthocyanin profile of red wine (Morgani *et al.*, 2023; Haselgrove *et al.*, 2000; Sivilotti *et al.*, 2020; Kyraleou *et al.*, 2016a; Ju *et al.*, 2021).

1. Effect of maceration time, skin contact, extraction methods and fermentation temperature on anthocyanins and their transformation

During winemaking, anthocyanins undergo different biochemical processes and transformations, changing their chemical and physio-chemical properties (Ribéreau-Gayon *et al.*, 2006). The extraction of anthocyanins and phenolic and non-phenolic compounds, as well as colour stability, depend on maceration time and fermentation conditions (Ribéreau-Gayon *et al.*, 2006). Maceration, as an extraction process, is a key step in red wine production, because it directly influences anthocyanin content. For this reason, different maceration lengths and techniques have been studied. Despite the importance of maceration as a step in red winemaking, there is a constant need to develop new methods to improve the sustainability and cost-effectiveness of winemaking. Reducing maceration time while ensuring appropriate wine quality would save energy use and reduce financial losses. Previous research on maceration length has shown that it influences the colour, and chemical and sensory properties of wines, since colour and anthocyanin concentration decrease with maceration length (Gil *et al.*, 2012). It has been recorded in the literature anthocyanin increases to its maximum concentration in the first five or six days of maceration (Ribéreau-Gayon, 1982).

TABLE 1. List of phenolic compounds found in different wines of different ages and their concentrations (in mg/L), as well as anthocyanins (% of total monomeric anthocyanins).

Phenolic compounds	Concentration	Wine age (months)	Wine variety	Winemaking technique	Source
Flavan-3-ol monomers	102.65–242.90	24	Syrah	Conventional winemaking	Garcia <i>et al.</i> , 2024
	130.9	3	Teran	Cold pre-fermentative maceration	Lukić <i>et al.</i> , 2017
	185.3			Pre-fermentative heating followed by traditional maceration	
	192.2			Post-fermentative maceration	
Flavan-3-ol dimers	75.29–156.90	24	Syrah	Conventional winemaking	Garcia <i>et al.</i> , 2024
	123.8	3	Teran	Cold pre-fermentative maceration	Lukić <i>et al.</i> , 2017
	157.3			Pre-fermentative heating followed by traditional maceration	
	163.1			Post-fermentative maceration	
Pyranoanthocyanins (vitisin A and B)	0.27–1.1	24	Syrah	Conventional winemaking	Garcia <i>et al.</i> , 2024
	Concentration (% of total anthocyanins)				
Glucosides	64.4–72.3	24	Syrah	Conventional winemaking	Garcia <i>et al.</i> , 2024
	83.13	3	Teran	Cold pre-fermentative maceration	Lukić <i>et al.</i> , 2017
	82.95			Pre-fermentative heating followed by traditional maceration	
	85.81			Post-fermentative maceration	
Anthocyanins	17.4–24.1	24	Syrah	Conventional winemaking	Garcia <i>et al.</i> , 2024
	9.26	3	Teran	Cold pre-fermentative maceration	Lukić <i>et al.</i> , 2017
	9.47			Pre-fermentative heating followed by traditional maceration	
	6.11			Post-fermentative maceration	
<i>p</i> -coumaroyl glucosides	9.05–14.4	24	Syrah	Conventional winemaking	Garcia <i>et al.</i> , 2024
	7.67	3	Teran	Cold pre-fermentative maceration	Lukić <i>et al.</i> , 2017
	7.66			Pre-fermentative heating followed by traditional maceration	
	8.11			Post-fermentative maceration	

As far as we know, to our knowledge, the kinetics of anthocyanin extraction and its result depend mostly on the berry skin properties, content and anthocyanin concentration of a given variety (Otteneder *et al.*, 2004). Numerous studies have reported an extended extraction duration to result in a decreased total anthocyanin content (Gil *et al.*, 2012; Sipiora *et al.*, 1998). Another has confirmed that maximum values for colour parameters and anthocyanin concentrations are obtained in wine that has been subjected to maceration for a duration of three to six days (Jagatić Korenika *et al.*, 2023). Opposite results were found by Alencar *et al.* (2017), who investigated the effect of maceration time on anthocyanin extraction in Syrah must and wine and showed that the extractions increased until the 20th day. The authors explained this phenomenon as being due to a markedly higher concentration of anthocyanins in the investigated Syrah grape skin because of its good adaptation to the agroecological conditions of the northeastern region of Brazil. The effectiveness of anthocyanin extraction during maceration, phenolic composition and colour characteristics might all be influenced by other factors, such as grape maturity, presence of seeds and grape solids, addition of oak and tannin, fermentation temperature or the implementation of different chemical and physical agents. Research has been conducted to ascertain new ways of speeding up extraction, of inducing better co-pigmentation and complexation of anthocyanins with other compounds, and of increasing colour stability, all while enhancing the sustainability of red winemaking. In this context, wines from very mature and ripe grapes contain a higher percentage of skin proanthocyanidins, and the extraction process is shorter (Gil *et al.*, 2012). Early seed removal caused a decrease in monomeric anthocyanin concentration, as well as gallic acid and flavan-3-ols (Jagatić Korenika *et al.*, 2023). The application of high hydrostatic pressure (HHP) at moderate pressure combined with oak chip maceration enhances the phenolic content and colour intensity of wine (Tao *et al.*, 2016). Giacosa *et al.* (2023) investigated seed impact on anthocyanin extraction kinetic of four Italian varieties, ‘Aglanico’, ‘Nebbiolo’, ‘Primitivo’, and ‘Sangiovese’. The results showed that the presence of seeds increased the polymerization rate, which is important for obtaining colour stability in mature wines. Concerning maceration and fermentation temperature, results in the literature differ in terms of temperature regimes and length of exposure to given temperatures. A study in California on three clones of Pinot noir showed that higher fermentation temperature (25 °C for 14 days) resulted in better anthocyanin extraction and higher anthocyanin concentrations (Reynolds *et al.*, 2022). The authors hypothesise that raising the fermentation temperature augments the extraction of phenolic compounds, and increases tannin extraction and the formation of polymeric pigments. These results are consistent with previous findings. Conversely, cold maceration has been shown through numerous studies to result in a decrease in anthocyanin concentration (Leong *et al.*, 2020; Kuchen *et al.*, 2018; Casassa *et al.*, 2019).

Techniques such as carbonic maceration, thermovinification, cold maceration (cryomaceration), pulsed electric field (PEF)

treatment, microwave treatment, ohmic heating and enzymatic treatment are among the winemaking methods that have been investigated (Tong *et al.*, 2023; Portu *et al.*, 2023; Zhang *et al.*, 2019; Pace *et al.*, 2014; Aleixandre-Tudo & Du Toit, 2018; Bianchi *et al.*, 2023; Fanzone *et al.*, 2022; López-Giral *et al.*, 2023; Gordillo *et al.*, 2021; Maza *et al.*, 2020; Wojdyło *et al.*, 2021; Río Segade *et al.*, 2015; Zhang *et al.*, 2021).

Being a water-soluble pigment, the extraction of anthocyanin occurs very quickly; meanwhile, the extraction of the other phenolic compounds starts with increased alcohol content during fermentation (Bautista-Ortín *et al.*, 2004). It has been previously reported that the concentration and content of anthocyanins extracted during maceration primarily depend on the characteristics of the grape variety. However, for other phenolic compounds and complex compounds co-pigmented with anthocyanins, the extraction method during winemaking plays a major role. It has been proven that different pre-treatment maceration techniques affect anthocyanin content (Wojdyło *et al.*, 2021).

Carbonic maceration is a winemaking process during which whole grape clusters are fermented in an anaerobic environment saturated with carbon dioxide. This technique initiates intracellular fermentation within the intact grapes, leading to unique biochemical transformations that influence the wine’s phenolic composition, aroma and flavour profile. Multiple studies on carbonic maceration have consistently demonstrated that this winemaking technique leads to a decrease in the concentration of individual anthocyanins, mostly monoglucosides and total phenols. Despite the reduced content of individual anthocyanins, research on carbonic maceration has indicated an increased potential for polymerization, which gives wines a higher hue (Chinnici *et al.*, 2009). Gonzalez-Arezana *et al.* (2020) have confirmed this by comparing the aromatic and phenolic composition of 84 commercial Tempranillo wines produced *via* carbonic maceration with wines produced by the standard (traditional) method of destemming and crushing. They reported the wines obtained *via* carbonic maceration to have higher colour intensity due to higher rates of ionization and polymerization. Additionally, the authors found coumaroyl derivatives and the vitisins A and B to be in higher concentrations in the wines made by carbonic maceration, which has also been confirmed by other researchers (Chinnici *et al.*, 2009; Portu *et al.*, 2023). Carbonic maceration results in wines with a lower average content in total phenolics, anthocyanins and resveratrol, but a higher concentration of catechins, as well as oligomeric and polymeric proanthocyanidins; therefore, the wines are less saturated but more brighter in colour due to higher chroma values (Zhang *et al.*, 2019). Wines made *via* carbonic maceration have greater colour intensity, more reddish hues and a high content in flavanols, hydroxycinnamic acids, ethyl-bridged anthocyanin isomers because of ionization of the pigments (Shmigelskaya *et al.*, 2021).

Thermovinification is a winemaking process that involves heating grape must to a temperature typically lower than 85 °C before fermentation for a certain controlled period

of time. This thermal treatment enhances the extraction of phenolic compounds, such as anthocyanins and tannins, from the grape skins and seeds, and it is thought to expeditiously result in wines with intensified colour, improved stability, and modified tannin structure. Research has been conducted to determine the validity of this theory. As a time- and temperature-dependent maceration method, thermomaceration has demonstrated its suitability for achieving higher concentrations of phenolic compounds compared to other methods (Aguilar *et al.*, 2016). However, in their review article, Maza *et al.* (2019) stated that thermovinification is associated with a range of issues stemming from the heating of the grapes, adversely affecting wine quality. Thermomaceration has shown different results depending on the grape variety. Some researchers have reported the use of thermal maceration to improve anthocyanin extraction, especially in Pinot noir (Girard *et al.*, 1997). Ntuli *et al.* (2023) investigated the impact of flash détente (FD) treatment on the chemical composition, colour stability and sensory profile of Merlot wines. The technique involved heating must to 85 °C, and the vacuum chamber was maintained at -0.94 bar. The temperature dropped to 32 °C, when the must entered the vacuum chamber. Results showed that the wine had significantly higher concentrations of caftaric acid, malvidin-3-*O*-glucoside and quercetin glycoside. While high temperatures caused the rapid pre-fermentation extraction of anthocyanins into must, the authors found that approximately 40 % of the colour was lost during fermentation. They also suggested that FD can be used to improve the body and astringency of the wine by extracting the polysaccharides and proanthocyanidins.

Ohmic heating is a relatively new pre-fermentative maceration technique based on the application of an electric field. Also known as the Moderate Electric Fields method, it induces electroporation of plant cell membranes. Ohmic heating generates an electric field of less than 1 kV/cm and the little research that has been conducted to date has shown it to be a promising method for shortening the maceration period and thereby conserving energy (Junqua *et al.*, 2021). The aforementioned authors report that, in comparison to conventional heating and no treatment of Aglianico and Barbera musts, ohmic heating resulted in a phenolic compound content twice as high as that before treatment. This indicates that the pre-fermentation process could be significantly shortened and simplified. The same research demonstrated that the total polyphenolic content in the finished wines was 17 % higher than in conventionally heated samples and 30 % higher than in untreated samples. Research conducted by Pereira *et al.* (2020) yielded similar results, postulating ohmic heating to be an environmentally sustainable technology for food processing. Their investigation of aqueous extracts from grape skin winemaking residues of the 'Vinho Verde' variety demonstrated that, irrespective of the temperature applied, ohmic heating increases the concentration of total phenolic compounds, soluble solids and colour intensity. Notably, total anthocyanin concentration increased from 756 to 1349 µg/g, primarily comprising malvidin-3-*O*-glucoside. These authors consider the internal heating generated by

ohmic heating to be advantageous, as it means that chemical solvents are not required, the treatment duration and water usage are reduced, and energy consumption is lower than during thermovinification.

Pulsed electric field (PEF) winemaking is a non-thermal processing technique that involves the application of short bursts of high-voltage electric fields to grape must. This process disrupts the cell membranes through electroporation, enhancing the extraction of intracellular compounds, such as phenolics and anthocyanins. Studies have shown that PEF treatment improves juice yield, accelerates maceration and preserves the sensory and nutritional qualities of wine, providing an energy-efficient alternative to conventional thermal methods (López *et al.*, 2008; López-Giral *et al.*, 2023). El Darra *et al.* (2016) has reported a 56 % increase in colour intensity of Cabernet-Sauvignon wine treated by PEF compared to that treated using other methods (thermovinification and enzymatic treatment). Additionally, total phenolic content was 18 % higher and total flavonol content 48 % higher. They also suggest that the higher intensity and difference in colour composition between the control and pretreated freshly-fermented model wines are not solely due to a higher content of residual native anthocyanins but also to co-pigmentation and the formation of derived pigments. Leong *et al.* (2020) investigated the effect of a continuous PEF system operated at high-intensity electric field strengths exceeding 30 kV/cm during a 4-day cold maceration of Merlot. This study is the first in the literature to apply such high-intensity electric fields. The results indicate that malvidin derivatives were the most significant anthocyanins to contribute to changes in colour intensity. Furthermore, it was noted that PEF at high-intensity electric field strengths and under continuous operation caused sufficient damage to grape cells, leading to the release of various types of anthocyanins. This technique could be particularly useful for the maceration of grape varieties with impermeable skins.

The use of the microwave technique in winemaking involves the application of microwave radiation to grape must or wine. This method generates heat through the dielectric heating of water molecules, leading to rapid and uniform heating. The microwave technique enhances the extraction of phenolic compounds and other desirable constituents from grape skins, improves colour stability, and accelerates the maceration process. It is a time-efficient and energy-saving alternative to conventional thermal methods, with the potential benefits of enhancing the sensory attributes and quality of the final wine. Carew *et al.* (2014) studied the influence of microwave maceration on phenolic extraction and fermentation kinetics of Pinot Noir in combination with early pressing. They reported the microwave pre-treatment to show significant cellular and phenolic integrity degradation and to accelerate the anthocyanin extraction. The results indicate that this technique could be an alternative solution to fermenting Pinot noir for seven days. After six months, the microwave wine had a higher anthocyanin content, which allowed colour to develop. Furthermore, other authors have reported the physical force employed by this method

to cause cell wall disruption, resulting in faster extraction kinetics. Additionally, this method can be advantageous when extracting acetylated anthocyanins, as they can become trapped within the matrix or form hydrogen bonds with polysaccharides (Sommer & Cohen, 2018). The same research found similar results for the application of the ultrasound technique. The use of ultrasound technique in winemaking involves applying high-frequency sound waves to grape must or wine. This process induces cavitation, which creates microbubbles that collapse and generate localized high temperatures and pressures. These effects enhance the extraction of phenolic compounds, improve maceration efficiency and facilitate the homogenization of the wine. Like the microwave treatment, ultrasound treatment can reduce processing times, improve the extraction of desirable compounds and preserve the sensory and nutritional qualities of the wine, making it an efficient and environmentally friendly alternative to traditional winemaking methods. Bautista-Ortín *et al.* (2017) investigated the application of high-power ultrasound in Monastrell red winemaking. The study examined maceration periods of 3, 6, and 8 days, and compared the results to a control vinification process involving 8 days of skin maceration without pre-treatment: in the initial must, the sonicated samples exhibited significantly higher levels of total phenols, total anthocyanins and colour intensity. These elevated levels persisted throughout the entire winemaking process, with the sample subjected to just three days of skin contact showing a higher concentration of anthocyanins than the control. After fermentation, the sonicated sample that underwent 3-day maceration maintained the highest amounts of total and polymeric anthocyanins and the highest colour intensity. Regarding the influence of sonication on anthocyanin composition, the anthocyanins present in the highest concentrations initially were dihydroxylated, cyanidin-3-*O*-glucoside and peonidin-3-*O*-glucoside. Following skin contact and fermentation, malvidin-3-*O*-glucoside was found in the greatest quantity. The authors reported that high-power ultrasound treatment was favourable for the extraction of skin tannins, as evidenced by the high concentrations of total tannins, the mean degree of polymerization, the percentage of galloylation and the concentration of epigallocatechin. This has been corroborated by recent studies, which investigated the effects of high-power ultrasound on the colour and aroma of Monastrell rosé wines: the sonicated wines exhibited higher colour intensity and increased total polyphenol and anthocyanin content (Labrador Fernandez *et al.*, 2023).

The use of high hydrostatic pressure (HHP) in winemaking involves subjecting grape must or wine to extremely high pressures, typically ranging from 100 to 600 MPa, in a liquid medium. This non-thermal technique deactivates spoilage microorganisms and enzymes, enhances the extraction of phenolic compounds and preserves the sensory and nutritional qualities of the wine. HHP treatment can improve wine stability, colour and the flavour profile while maintaining the integrity of heat-sensitive compounds, providing an innovative and efficient alternative to traditional winemaking processes. Given its potential applications in winemaking, the high

hydrostatic pressure (HHP) technique has been investigated to improve winemaking sustainably. Tao *et al.* (2012) conducted a study to investigate the influence of high hydrostatic pressure (HHP) on the physicochemical and sensorial properties of the 2010 vintage of the Nero d'Avola Syrah red wine. The treatment involved subjecting the wine to 650 MPa for durations of 0.25, 0.5, 1 and 2 hours. The results showed a shift in the equilibrium of chemical reactions within the wine, with an enhancement of its organoleptic properties and acceleration of the aging process. In terms of phenolic composition, the treatment resulted in a decrease in content of total phenols, total anthocyanins, tartaric esters, flavonols and tannins, mirroring changes that occur during natural aging. Regarding anthocyanin composition, the levels of monomeric and polymerized anthocyanins decreased post-pressurization, while the level of co-pigmented anthocyanins showed a slight increase, likely due to condensation reactions induced by HHP. Another study demonstrated the influence of high hydrostatic pressure (HHP) processing combined with oak chip maceration on the physicochemical and sensory properties of young red Sangiovese and Merlot blend (Tao *et al.*, 2016). The results showed that young wine subjected to 250 MPa, 450 MPa and 650 MPa for 45 min and macerated with oak chips had higher total phenolics, tartaric esters and flavonols in comparison to the untreated control wine. The concentration of total monomeric and polymeric anthocyanins increased, probably due to HHP influence on potential desorption of anthocyanins from the oak chips, and the tannic compounds extracted from the oak chips reacted with anthocyanins to form new polymeric compounds. This technique is noted as potentially beneficial for both red wine quality and the costs associated with its production, since it increases colour stability by influencing the formation of polymeric pigments and accelerates wine aging without the need for the use of oak barrels. Other researchers have also reported that HHP can influence long-term red wine physicochemical and sensorial characteristics, increasing the condensation reactions of phenolics (Santos *et al.*, 2013; Christofi *et al.*, 2020).

Enzymatic treatment in red winemaking involves the adding specific enzymes, such as pectinases, cellulases and hemicellulases, to the grape must. These enzymes catalyze the breakdown of complex polysaccharides in the grape cell walls, facilitating the release of phenolic compounds, anthocyanins and other desirable constituents. This process enhances colour extraction, improves juice yield, accelerates clarification and stabilizes the wine. Enzymatic treatment is a controlled and efficient method for optimizing the maceration process, improving sensory attributes, and increasing the overall quality of the red wine. Enzymatic maceration depends highly on choice of enzymes (Wang *et al.*, 2016) and the thickness and structure of the berry skin of the 'Syrah' and 'Cabernet-Sauvignon' grape (Apolinar-Valiente *et al.*, 2016). Scientific research has demonstrated that the addition of commercial enzymes, including polygalacturonase and cellulase, does not increase the concentration of anthocyanins relative to other winemaking techniques. In the comparative study of enzymatic maceration with thermomaceration in 'Lachryma

Christi' and 'Pais' winemaking, thermomaceration resulted in the extraction of higher concentrations of phenolic compounds. The low anthocyanin content after the enzymatic treatment might be due to the formation of an enzyme polyphenol complex as a result of hydrophobic interactions (Aguilar *et al.*, 2016; Wang *et al.*, 2016). However, it is noted that the addition of pectolytic enzymes might influence the greater concentration of iron, calcium and magnesium in wines (Soto Vázquez *et al.*, 2013).

Cryomaceration in winemaking is a cold maceration technique in which grape must is subjected to low temperatures, typically around 5-10 °C, before fermentation. This process slows down enzymatic activity and microbial growth, resulting in a prolonged extraction of aromatic compounds, phenolics and colour from the grape skins without the risk of oxidation. However, Aleixandre-Tudo and Du Toit (2017) have found that the efficiency of this method depends on the variety, maturity of the grapes, the temperature applied, the maceration length and potential combination with other techniques, such as enzyme or dry ice addition. In a comparative study of the effect of different alternative winemaking techniques on the pigment profile of red Tannat wines, Gonzalez-Neves *et al.* (2010) did not find cold pre-fermentation maceration to significantly increase anthocyanin content. The reason for these results might be the low extractability of the pigments in the Tannat grapes. Casassa *et al.* (2016) reported that a cold prefermentative soak affects colour and wine phenolics of Barbera D'asti and Malbec wine only with the addition of SO₂ of 100 mg/L. This method positively influenced colour intensity and violet hue. In their study on thermal pre-fermentative treatment of André wine, Ševcech *et al.* (2015) found that the cold soak gave the best results regarding anthocyanin concentrations.

2. Wine aging biochemical transformation and colour changes

During winemaking and aging, anthocyanins undergo different chemical reactions and transformations, being influenced by various factors (grape variety, maceration time, yeasts, winemaking technique, amount of SO₂, wine aging, fining agents and micro-oxygenation). More complex polyphenolic compounds are formed through co-pigmentation, condensation and polymerization reactions (Boulton *et al.*, 2001). The compounds formed in these reactions are condensed tannin (flavan-3-ol) products, better known as polymeric pigments, and different groups of pyranoanthocyanins (vitisin A, vitisin B, vinylphenolic pyranoanthocyanins, vitisin A derivatives, and other types of pyranoanthocyanins-oxovitisins, methylpyranoanthocyanins and pyranoanthocyanin dimers). Vinylphenolic pyranoanthocyanins include pinotins and flavanol-pyranoanthocyanins, while portisins are a distinct class of derived pigments usually found in Port wine (Waterhouse & Zhu, 2019). Numerous studies have demonstrated that these biochemical transformations affect the phenolic composition, colour intensity and hue, colour stability and sensory properties of red wine, which have been thoroughly reviewed by Qualgieri *et al.* (2017). The

complexation of anthocyanins with various molecules (other anthocyanins [self-association], tannins, proteins and ions) during fermentation leads to an increase in colour intensity due to the co-pigmentation phenomenon (Moreno-Arribas & Polo, 2009). Co-pigmentation causes hyperchromic effect and bathochromic shift, leading to higher colour density and a richer purple hue of the wine (Zhang *et al.*, 2022). The percentage of the contribution of intermolecular co-pigmentation to colour has been estimated to be up to 30-50 % in young red wines (Boulton *et al.*, 2001), while that of monomeric anthocyanins, in the form of flavylium cation, is 30-70 % (Brouillard *et al.*, 1982). Polymerized pigments are the main contributors of colour to aged wines, contributing to 35-63 % of the wine colour (Han *et al.*, 2008). Many authors have used well-known CIELAB parameters to define the colour of wine, namely lightness (L^*), redness (a^*), blueness (b^*), chroma (C^*), hue angle (H^*) and ΔE^* (colour difference). Young wines are dark, with a high colour density, and they are predominantly violet-red. Han *et al.* (2008) applied principal component analysis of the relationship between CIELAB colour and monomeric anthocyanins in young Cabernet-Sauvignon wines: various monomeric anthocyanins exhibited negative correlations with the L^* , b^* , and H^* values, and positive correlations with the a^* and C^* values. During wine aging the value of L^* , b^* , and H^* parameters increases due to the decrease in colour density, loss of violet hue and accumulation of tawny tones (McRae *et al.*, 2012). Chromatically, the colour of red wine during wine aging changes from bright red and deep purple to pale red (Apolinar-Valiente *et al.*, 2016), and thus red wine colour can be used as an indicator of age (Wang *et al.*, 2023). Furthermore, it has been observed by (Zhang *et al.*, 2021) that the tawny characteristics of aged wines are attributable to pyranoanthocyanins, excluding vitisin B, with a particular emphasis on pinotins. Being more stable phenolic compounds that are formed during the aging process through direct cycloaddition of malvidin and caffeic acid, pinotins maintain colour intensity and add to the complexity of wine's hue. Chromatically, they are connected to the tawny and brick-red colours of the well-aged red wines. The colour density associated with aging is closely linked to the concentration of vitisin A and flavanyl-pyranoanthocyanins (Zhang *et al.*, 2021). The same group of authors noted that evolution and stability patterns differ among anthocyanin derivative classes, with pinotins as the most stable anthocyanin derivative compound, followed by flavanyl-pyranoanthocyanins, vitisin A, monomeric anthocyanin and direct anthocyanin-flavan-3-ol condensation products, and then vitisin B and anthocyanin ethyl-linked flavan-3-ols products as the least stable. They were investigating the anthocyanin derivatives and chromatic characteristics of 234 different vintage red wines (of the varieties Cabernet-Sauvignon, Syrah, Merlot, Cabernet franc, Tempranillo, Zinfandel, Pinotage, Carmenere, and Marselan) from 13 countries. Chromatic changes in wine correspond to biochemical changes, including alterations in the ratio between non-acylated and acylated anthocyanins. Acylation, a process that occurs during wine aging, significantly impacts

anthocyanins—which are crucial sensory constituents of red wine—and plays an important role in the formation of pyranoanthocyanins and polymeric pigments, leading to colour changes as acylated and non-acylated anthocyanins are lost (Wang *et al.*, 2023). The same authors reported that the younger wines had higher concentrations of acylated anthocyanins than non-acylated anthocyanins due to the slow degradation of acylated ones. Moreover, the decrease in both acylated and non-acylated anthocyanins was accompanied by the development of pyranoanthocyanins and polymeric pigments, with a reduction in the a^* value and an increase in the b^* and H^* values.

Generally, after fermentation, anthocyanins continue to react with other polyphenolic compounds, including the derived pigments and tannic compounds extracted from the grapes (Boulton, 2001). Each step after fermentation can cause a different reaction and result in the formation of different phenolic compounds; therefore the treatments and wine aging vessels should be chosen appropriately. Numerous researchers have investigated the effects of aging wine in oak barrels, adding oak chips and micro-oxygenation on red wine's phenolic composition and colour. Red wines typically undergo a maturation process, primarily in wooden barrels or bottles, which constitutes a critical stage in their production (Teissedre & Jourdes, 2013). Consequently, monitoring the physicochemical properties of wine throughout the aging period is essential. Watrelot and Waterhouse (2018) investigated the percentage of the degradation and loss of monomeric anthocyanins and the percentage of the formation of pigmented tannins in 'Cabernet-Sauvignon' red wines, aged 8 and 12 months in barrels of different toasting levels (low LTP, medium MTP, and high HTP). Being the most reactive monomeric anthocyanin, malvidin coumaroyl glucoside was found, and its loss was significant. The loss of coumaroylated anthocyanins is a result of the reaction of ester hydrolysis with the acyl group or its precipitation, and is not a result of oxidation. Red wine aged in barrels toasted at lower temperatures exhibited higher levels of ellagitannins, as higher temperatures lead to their degradation. Meanwhile, the wines that contained lower concentrations of ellagitannins showed a higher percentage in loss of monomeric anthocyanins. These results indicate that ellagitannins protect anthocyanins and stabilize wine colour by forming new compounds with them, as was also suggested by Chassaing *et al.* (2010). The aging process in oak barrels imposes significant financial costs on wineries due to both the expense of the barrels and the extended duration required. Substantial efforts have been made to develop alternative, cost-effective methods to reduce the aging period while minimizing financial expenditures and maintaining the high quality of the wines (Ferreiro-Gonzales *et al.*, 2019). Some of these efforts include the addition of oak chips, micro-oxygenation treatments and high hydrostatic pressure (HHP) treatment, the latter of which is still undergoing validation.

The addition of oak chips can influence wine characteristics, depending on the timing of their addition. When added

during fermentation, oak chips do not favorably affect ellagitannin extraction and anthocyanin stabilization, but the wines can contain wood-extracted volatiles, such as lactones, ramified ethyl esters, and acetates. The addition of the chips after fermentation may give wines greater aging potential and increase the condensation reactions of tannins and anthocyanins (Kyraleou *et al.*, 2016b). Comparative studies have been conducted to evaluate the differences between the influence of oak barrels, the addition of oak chips, micro-oxygenation and high hydrostatic pressure (HHP) on the polyphenolic complexation and colour characteristics of wine. Cano-Lopez *et al.* (2010) reported that, like oak barrel aging in the same aging period (3 months), micro-oxygenation improves quality and colour of Monastrell wine. Nevertheless, when analyzed after aging for six months in a bottle, the wines showed different chromatic characteristics, with a higher percentage of yellow tint. The assumption is that ellagitannins, phenolic acids and wood aldehydes extracted from wood during barrel aging form anthocyanin-tannin complexes which stabilize the colour. Gonzales-Sais *et al.* (2014) carried out similar research on the Spanish variety Tempranillo to determine the effect of different treatments that advance the aging process. Namely, they used different oxygen doses, oak chip dose, wood origin (French and American oak), different toasting degree and maceration time. The objective was to investigate the influence of controlled doses of oxygen and the addition of oak chips to wine by mimicking the conditions of oak barrel aging. The results showed that changes in anthocyanin content during aging are associated with numerous processes that compete for the same substrates: monomeric anthocyanins. The final state of anthocyanin content results from a complex equilibrium of these reactions and mechanisms, which are highly sensitive to variations in experimental conditions. More specifically, the higher degree of toasting had an advantage over the maceration time, due to higher extraction and formation of complex polymeric anthocyanin with flavan-3-ols and proanthocyanidins, and the formation of pyranoanthocyanidins. Meanwhile, high oxygen doses in combination with medium to medium-plus toasting levels showed the best results, with the formation of anthocyanin derivative complex compounds; however, when medium to high oxygen doses were combined with heavy-plus toasted chips a relatively high level of anthocyanins remained. According to Ćurko *et al.* (2021), micro-oxygenation enhances polymerization reactions between flavanols and anthocyanins *via* acetaldehyde mediation.

As already mentioned, high hydrostatic pressure (HHP) is one of the new sustainable winemaking techniques for accelerating the extraction of anthocyanins and other polyphenolic compounds during maceration. Like the other techniques described, its influence on the physicochemical and sensorial properties of red wine has been investigated. It has been employed for producing high-quality wines and reducing manufacturing costs with effective inhibition of spoilage microorganisms. Tao *et al.* (2012) investigated the influence of HHP processing of 650 MPa at an ambient

temperature of around 18 °C, for 0.25, 0.5, 1, and 2 h, respectively, on the phenolic complex of Nero D'Avola Syrah wine. The aim was to investigate the possibility of using HHP as a technique for accelerating the wine aging process. The best results were obtained for the 2 h treatment, with a reduction in the intensity of the wine colour and in phenolic compound content; it also enhanced the sourness, astringency, alcoholic and bitter taste of the wine. The authors suggest this method is useful for the aging of wines with low aging potential (low anthocyanin and tannin content).

3. Influence of anthocyanins on red wine astringency

Wine phenolics influence the flavour and mouthfeel of the wines (Setford *et al.*, 2017). While astringency might be defined as a tactile sensation (Breslin *et al.*, 1993) of the oral cavity triggered by harsh food compounds [*i.e.*, tannins] (Gibbins *et al.*, 2013), it is perceived more as a feeling (Pires *et al.*, 2020). To characterize astringency as a complex sensation, Gawel *et al.* (2000) developed terminology linked to a “mouth-feel wheel” to systematically describe the sensory attributes of red wine. A total of 33 astringency characteristics, are classified into 7 classes: particulate, surface smoothness, complex, drying, dynamic, harshness and unripeness. During red wine tasting, astringency occurs due to the tannins-salivary protein interaction (Breslin *et al.*, 1993), which involves the precipitation of proteins, leaving a puckering sensation and oral dryness as an aftertaste (Green, 1993). The most studied mechanism of astringency is the interaction between salivary proteins rich in proline (PRPs) and astringent compounds (Pires *et al.*, 2020). Proline-rich proteins are very reactive and are divided into three groups: acidic (aPRPs), basic (bPRPs) and glycosylated (gPRPs). It has been previously reported that their presence in mammals' saliva is connected to the consumption of food rich in tannins (Bennick, 1982). In biochemical terms, astringency might occur because of noncovalent binding interactions; for example, the hydrophobic effect and hydrogen bonds that occur between PRPs and tannins (García-Estévez *et al.*, 2018; Delić *et al.*, 2023). Because anthocyanins interact with tannins during the production of red wine to form anthocyanin-tannin complexes, the question arises of whether there is an interaction mechanism between anthocyanins and salivary proteins (Paissoni *et al.*, 2018; Ferrer-Gallego *et al.*, 2014; Ferrer-Gallego *et al.*, 2015; Paissoni *et al.*, 2020; Mattioli *et al.*, 2020). Some studies have shown that there are anthocyanin-salivary protein complexes, but the interaction between them has not yet been explained (Ferrer-Gallego *et al.*, 2015). Many recent studies have worked on determining the influence of anthocyanins on enhancing or reducing the astringency of red wine (Paissoni *et al.*, 2018). Previous studies have shown that, during wine aging, anthocyanins undergo reactions with tannins to form polymeric pigments. When their sensorial effect was investigated, these polymeric pigments were found to interact with salivary proteins and show decreased

astringency levels (Villamor *et al.*, 2009). Additionally, polysaccharides in aged wines can bind to anthocyanins and tannins, decreasing the perception of astringency (Escot *et al.*, 2001). Therefore, molecular investigations are needed in order to better understand the interaction between anthocyanins and salivary proteins necessitates.

The affinity between plant pigments and salivary proteins has been investigated and found to probably depend on the functional groups and molecular weight of pigments (Yao *et al.*, 2011). The effect of different anthocyanin fractions (glucoside, acetylated and cinnamoylated fractions) on oral sensory properties and astringency have also been studied, with the conclusion that cinnamoylated anthocyanins are the most reactive anthocyanins to salivary proteins rich in proline (Paissoni *et al.*, 2018). Soares *et al.* (2019) carried out research to determine whether the co-pigmentation of malvidin-3-*O*-glucoside and epicatechin could affect the ability of flavonols to interact with PRPs, *via* saturation-transfer difference (STD)-NMR and isothermal titration calorimetry (ITC). The epicatechin-malvidin-3-*O*-glucoside mixture showed the same affinity for PRPs as individual compounds; in addition, epicatechin was found to involve hydrophobic and hydrophilic interactions, while malvidin-3-*O*-glucoside involved electrostatic interactions. Meanwhile, recent studies have investigated the role of anthocyanins in the interaction between salivary mucins and wine astringent compounds (Torres-Rochera *et al.*, 2023), with the results showing that, when isolated, anthocyanin (namely malvidin-3-*O*-glucoside) had the strongest affinity for salivary mucins compared to catechin, epicatechin and quercetin-3- β -glucopyranoside. Additionally, according to the authors, the co-pigmentation phenomenon might have a larger purpose, since the presence of malvidin-3-*O*-glucoside modified the intensity and characteristics of the interactions between the mucins and other phenolic compounds. Similarly, Mao *et al.* (2024) studied the interactions between oral mucins and cyanidin-3-*O*-glucoside, emphasizing the effect of oxidized quinone. They explain that when the anthocyanins oxidize into quinones they covalently bind with mucins in the oral cavity and form a tighter cross-linkage, enhancing oral astringency. Some researchers went further by developing different oral models and investigating the interactions of phenolic compounds in conditions similar to the human oral cavity. Soares *et al.* (2020) developed oral epithelia comprising buccal mucosa and tongue, human saliva and the mucosal pellicle in order to investigate the interactions with anthocyanin red wine extract and green tea flavanol extract. They reported that anthocyanins had higher interaction with oral cells only, but the studied anthocyanins (delphinidin-3-*O*-glucoside, peonidin-3-*O*-glucoside, petunidin-3-*O*-glucoside and malvidin-3-*O*-glucoside) all showed the same level of interaction ability. Additionally, it was shown that various oral constituents can perform distinct functions at different phases of phenolic compound intake. These findings may serve as a useful starting point for future investigations into the influence of anthocyanins on red wine astringency.

APPLICATION OF ANTHOCYANINS IN THE FOOD INDUSTRY

The antioxidant properties of anthocyanins make them essential in food technology. They can serve as natural food colorants or specific indicators for food quality control. The estimated daily intake (ADI) for humans is 2.5 mg/kg body weight per day (Clifford, 2000). Enhancing the stability of anthocyanins would increase their use in food technology (Mattioli *et al.*, 2020). To improve the stability and ingestion of these compounds, encapsulation techniques have been developed. This method is also widely used to enhance the quality of food in terms of colour, aroma, smell and taste. Various studies have explored different types of encapsulations, materials, matrices and carriers for polyphenol-loaded microparticles (Popović *et al.*, 2019; Milinčić *et al.*, 2019; Milinčić *et al.*, 2022b; Milinčić *et al.*, 2023a; Đorđević *et al.*, 2015; Popović *et al.*, 2019). Micro- and nanoencapsulation are among the most effective methods for incorporating anthocyanins within bioactive compounds and facilitating their introduction into the human body. Researchers are addressing challenges such as low stability, low bioavailability and controlled release of anthocyanins *via* new micro- and nanoencapsulation techniques (Milinčić *et al.*, 2019; Milinčić *et al.*, 2022b; Milinčić *et al.*, 2023a). For example, Lavelli *et al.* (2019) investigated the microencapsulation of grape skin phenolics and its controlled release, utilising calcium chloride as a hardening agent. Their findings indicate that alginate is effective as a pH-controlled release system for grape skin phenolics. Additionally, seed/skin extracts are frequently added to various products, such as meat, cereal-based infant formula (Pešić *et al.*, 2019), goat milk yogurt (Milinčić *et al.*, 2021b; Milinčić *et al.*, 2022b; Milinčić *et al.*, 2024) and biscuits (Kammerer *et al.*, 2005) to enhance techno-functional and functional properties, or to examine the bioavailability of phenolic compounds in these matrices.

1. Anthocyanins as natural food colorants and additives

Food colour is constantly being improved, especially with the development of new technologies for the use of natural colours. Increased awareness of the harmful health effects of synthetic colorants can be achieved by utilising anthocyanins as natural food colorants. For these purposes, different encapsulation techniques have been used (Lavelli & Sri Harsha, 2019; Ghosh *et al.*, 2021) and studied, namely coacervation, spray drying, freeze drying, liposomal systems, electro spraying and electrospinning, inclusion complexation, emulsification, ionic gelation, and extrusion (Shaddel *et al.*, 2018; Kanha *et al.*, 2020; Adali *et al.*, 2020; Zhang *et al.*, 2020; Atay *et al.*, 2018; Forghani *et al.*, 2021; Fernandes *et al.*, 2018; Norcino *et al.*, 2022; De Moura *et al.*, 2018; Mohammadalnejhad *et al.*, 2023). In micro- (particle size between 1 µm and 1,000 µm per unit) and nanoencapsulation (particles of 50 nm and 500 nm in size), the most common encapsulation techniques are spray drying and electrospinning, which have been thoroughly reviewed by Sharif *et al.* (2020). Recent research on the

microencapsulation of anthocyanins combined different wall materials and different encapsulation techniques to achieve higher stability of encapsulates, better preservation of microcapsule colour, a longer storage period, higher antioxidant capacity, sustained release of the bioactive compound, and high encapsulation efficiency. The use of different wall materials, such as alginate (Lević *et al.*, 2015), gelatin (Gao *et al.*, 2022), inulin (Enache *et al.*, 2022), pectin (Pereira Souza *et al.*, 2017), chitosan (Wang *et al.*, 2016) and whey protein isolate (Wang *et al.*, 2022), has improved existing encapsulation techniques and provided new solutions for the development of strategies and methods for producing microparticles containing anthocyanins. Two different paths of scientific research are associated with the use of anthocyanin nanoencapsulation (Milinčić *et al.*, 2019): one focussing on the fabrication of nanofibers or films for developing new, smart/intelligent food packaging using anthocyanins as biosensors of food spoilage and microbiological activity (Almasi *et al.*, 2022); and the other to find new ways of producing anthocyanin-loaded nanoparticles increase their use in food technology and potentially give a new perspective on their use (Nivetha *et al.*, 2022). The encapsulating base material is very important since the concentration and molecular structure of the anthocyanins will depend on it. Grapes, pomace and wine have been actively investigated as sources of anthocyanins (Tikhonova *et al.*, 2021; Lavelli & Sri Harsha, 2019; Brezoiu *et al.*, 2019), using varieties such as Ancellota and Aspirant Bouchet wines to produce encapsulated natural colorant powders because of their high anthocyanin content. The encapsulate properties of the colouring powder was evaluated, and the results showed that the increase from 135 to 145 °C in inlet air temperature did not influence the anthocyanin profile in the wine powder; thus indicating that the encapsulates were appropriate for potential use as natural food colorants (Alvarez Gaona *et al.*, 2022). Other authors have reported that using spray drying to produce powders resulted in encapsulates with low moisture content, low hygroscopicity, high solubility and stable colour (De Souza *et al.*, 2015). A widely-used anthocyanin-rich extract obtained from grape pomace called Enocianina is one of the most important colorants in the food industry, being rich in polyphenols and anthocyanins (mostly malvidin derivatives) and also showing anti-inflammatory properties (Della Vedova *et al.*, 2022).

2. Grape pomace as source of anthocyanins

Economically, grapevine production is one of the most important productions in the world. According to the statistical data reported by the Food and Agriculture Organization (FAO) for 2022, grape production was almost 75 million tons (FAO, 2022). The International Organization for Vine and Wine (OIV) reported that in 2023 world wine production was the lowest it had ever been in 60 years, with a decrease of 7 % compared to 2022 (OIV, 2024). They stated that extreme climatic conditions have significantly impacted vineyard output worldwide. Overcoming meteorological problems that occur during grapevine production is often costly, which further increases the costs of wine production in general.

Many wineries have turned to sustainable winemaking, trying to use as much waste as possible from their production, namely pomace. Not only is the practice of using different by-products cost-effective, but it is also environmentally friendly and contributes to the fight against climate change, and air, soil and water pollution. Grape pomace is a by-product of vinification and winemaking. Soceanu *et al.* (2021) state that as a by-product of winemaking, pomace should be considered as a starting point for creating new products that are nutritionally and industrially valuable. At the same time, this would decrease the harmful ecological impacts of winemaking related to inconsistent waste disposal practices. Since it is rich in phenolic compounds (listed in Table 2), grape pomace can be used in many different industries. The composition and concentration of anthocyanins within grape pomace vary depending on grape variety, growth conditions and winemaking methodology. To extract anthocyanins from grape pomace, several techniques such as solvent extraction, enzymatic hydrolysis, microwave-assisted extraction and supercritical fluid extraction have been employed (Milinčić *et al.*, 2021a; Valls *et al.*, 2017; Castellanos-Gallo *et al.*, 2022; Monteiro *et al.*, 2021; Muñoz *et al.*, 2021; Peixoto *et al.*, 2018; Milinčić *et al.*, 2023b). Extraction parameters, such as solvent selection, temperature, pH and extraction duration, play a critical role in influencing the yield of anthocyanins (Peixoto *et al.*, 2018). Hoss *et al.* (2021) have noted that grape pomace contains a high content of bioactive molecules, which, once extracted, can be used in cosmetic products due to the antioxidant, antiaging, anti-hyperpigmentation and photoprotective effects of the phenolic compounds. According to Gomez-Brandon *et al.* (2021) grape pomace can potentially be used for the production of vermicompost, which increases the content of macro- and micronutrients in the soil. Another group of scientists studied the potential use of winemaking pomace as a new source of compounds for antibacterial agents: due to their bacteriostatic and/or bactericidal properties, phenolic compounds that have synergistic action with antibiotics might prevent bacterial resistance to the antibiotic (Silva *et al.*, 2021).

Anthocyanins from grape pomace can be successfully used in the formulation of different nutraceuticals and functional foods. Nutraceutical products (*i.e.*, dietary supplements, capsules and powders) can be formulated to deliver concentrated anthocyanin extracts. The enrichment of food products with grape pomace powder or extracts has been shown to improve the functional properties of the food and to have health benefits (Troilo *et al.*, 2022). The incorporation of grape pomace into these products not only increases their anthocyanin content, but also imparts desirable attributes related to, for example, colour, flavour and texture (Monteiro *et al.*, 2021, Tikhonova *et al.*, 2021). A comparative study of white wine Pinot noir pomace flour and white wine Pinot noir pomace extracted in hot water, showed the higher potential of the extracts for use as an antioxidant dietary ingredient. The extracts had higher mineral and soluble fibre content, and human digestion simulation showed that digested fractions had higher bioactive value, with gallic, vanillic and seringic acid as the main bio-accessible phenolic compounds

(Beres *et al.*, 2019). Adding grape pomace (specifically, 5 % of grape seed flour) as an ingredient to bread gave interesting results regarding sensorial analysis and customer acceptance. The panelists found it to be very similar to black bread (Oprea & Gaceu, 2020). Troilo *et al.* (2022) studied the addition of different particle sized fractions of pomace powder to flour and their influence on the chemical, technological and sensorial characteristics of functional muffins. They reported that particle size was not the decisive factor when 15 % of grape pomace powder was added. The functional muffins were valuable sources of fiber with high content of antioxidants. The utilization of grape pomace as a source of anthocyanins and dietary fibre promotes sustainability within the agri-food industry (Milinčić *et al.*, 2020; Milinčić *et al.* 2021a). Finally, the valorization of grape waste by using it for obtaining highly valuable bioactive compounds is environmentally friendly and is one of the principles of circular economy (Rajković *et al.*, 2020).

3. Anthocyanins as smart/intelligent food packaging bio-compounds

Being bioactive compounds, anthocyanins can be used as incorporated agents for the fabrication of nanofibers or films and the development of new, smart/intelligent food packaging (Forghani *et al.*, 2021; De Silva *et al.*, 2022). For this purpose, anthocyanins are used as biosensors of food spoilage and microbiological activity, because they easily change colour when pH changes (Almasi *et al.*, 2022). There are two different types of intelligent food packaging: food freshness indicators (FFI) and time-temperature indicators [TTI] (Forghani *et al.*, 2024; Almasi *et al.*, 2022). There are different methods of preparation of food freshness indicators, as well as different intelligent food packaging. Recent advances have been made in research on 3D printing technology combined with incorporating anthocyanins in intelligent packaging. Bao *et al.* (2024) conducted research on a starch based 3D printed intelligent colorimetric film with blueberry anthocyanin-phycoerythrin incorporated (with chondroitin sulfate as a co-pigment). Colorimetric film exhibited high sensitivity to ammonia response, high antioxidant activity, biosafety and degradability, showing efficacy in maintaining the freshness of salmon and beef. Li *et al.* (2024) developed a composite film made of soybean protein isolate/carboxymethyl cellulose sodium/blueberry anthocyanin by *in situ* incorporation of anthocyanin, which showed UV-shielding, gas barrier and water resistance performance. Developing an intelligent edible electrospun nanofiber film for shrimp preservation, Wu *et al.* (2024) incorporated anthocyanin and thymol into polymer fibers of gelatin and zein. The results showed that the film had high mechanical properties and decreased water vapour permeability, excellent sensitivity to change of the pH solutions, high antioxidant properties and antibacterial responses. Additionally, it extended shelf life for packaged shrimps by 11 days when at 4 °C. Qin *et al.* (2024) developed highly pH-sensitive film for quantifying fish freshness in real-time based on a chitosan/gelatin matrix and incorporating *Zingiber striolatum* Diels anthocyanin extract. When tested, the film showed greater melting temperature

TABLE 2. List of phenolic compounds found in grape pomace (skins, seed stems) and their concentrations expressed as mg/g dry weight (DW).

Phenolic compounds	Concentration	Grape variety/Pomace type	Source
Gallic acid	0.07	Prokupac seed, skin and stem pomace	Milinčić <i>et al.</i> , 2021a
	0.01–0.03	Grenache, Syrah, Carignan noir and Mouvedre skin pomace	Ky <i>et al.</i> , 2014
Catechin	0.19	Prokupac seed, skin and stem pomace	Milinčić <i>et al.</i> 2021a
	0.02–0.08	Syrah, Merlot and Cabernet-Sauvignon pomace	Lingua <i>et al.</i> , 2016
	0.003–0.01	Grenache, Syrah, Carignan noir, Mouvedre skin pomace	Ky <i>et al.</i> , 2014
Epicatechin	0.12	Prokupac seed, skin and stem pomace	Milinčić <i>et al.</i> , 2021a
	0.02–0.1	Syrah, Merlot and Cabernet-Sauvignon pomace	Lingua <i>et al.</i> , 2016
Quercetin	0.01	Prokupac seed, skin and stem pomace	Milinčić <i>et al.</i> , 2021a
Resveratrol	0.04	Prokupac skin and stem pomace	Milinčić <i>et al.</i> , 2021a
	2.9–9.0	Grenache, Syrah, Carignan noir and Mouvedre skin pomace	Ky <i>et al.</i> , 2014
Anthocyanin glucosides	1.35	Prokupac skin and stem pomace	Milinčić <i>et al.</i> , 2021a
	0.05–0.14	Syrah, Merlot and Cabernet-Sauvignon pomace	Lingua <i>et al.</i> , 2016
Anthocyanins acylated	0.05	Prokupac skin pomace	Milinčić <i>et al.</i> , 2021a
	0.2–0.8	Grenache, Syrah, Carignan noir and Mouvedre skin pomace	Ky <i>et al.</i> , 2014
	0.02–0.2	Syrah, Merlot and Cabernet-Sauvignon pomace	Lingua <i>et al.</i> , 2016
Anthocyanins <i>p</i> -coumaroylated	0.15	Prokupac skin pomace	Milinčić <i>et al.</i> , 2021a
	0.7–5.2	Grenache, Syrah, Carignan noir and Mouvedre skin pomace	Ky <i>et al.</i> , 2014
	0.07–0.4	Syrah, Merlot and Cabernet-Sauvignon pomace	Lingua <i>et al.</i> , 2016
Total Tannins	31.8 – 56.1	Grenache, Syrah, Carignan Noir and Mouvedre skin pomace	Ky <i>et al.</i> , 2014

and a lower weight loss at melting temperature, with a visible colour change from red to yellow green in a pH 1–14 buffer. Teixeira Gomez *et al.* (2024) reported that the sensitivity of the films and their behaviour depends on the source of the extract of anthocyanins. They developed gelatin/polyvinyl alcohol films incorporated with different blueberry extracts (raw blueberry extract, purified fractions of phenolic extract and anthocyanin extract). The film with anthocyanin extract was more sensitive to acidic vapours, changing the colour from green to purple, while the film with raw blueberry extract was more sensitive to basic conditions. This might be because of the presence of other phenolic compounds in the raw blueberry extract, which are less susceptible to acidic conditions in comparison to anthocyanins. Therefore, the phenolic composition of the extract is of high importance when developing the food indicator film. The use of anthocyanin extracts in the development of novel and intelligent indicator food film and packaging is vast and still to be thoroughly explored, with high potential in food industry.

BENEFITS OF GRAPE AND WINE ANTHOCYANINS FOR HUMAN HEALTH

Anthocyanins have multiple protective and therapeutic properties for human health (Zia ul Haq *et al.*, 2016). Being bioactive compounds, anthocyanins have a wide range of properties; they are, for example, antioxidants (Monteiro *et al.*, 2021) and anti-inflammatories (Della Vedova *et al.*, 2022), and they also regulate blood lipids (Bayram *et al.*, 2024). Furthermore, they show insulin resistance, and anti-mutation and anti-tumour

properties (Li *et al.*, 2021). Many studies have confirmed that anthocyanins can also improve visual (Liu *et al.*, 2024) and brain functions (Baek *et al.*, 2023; Hasan *et al.*, 2023). Their role in the prevention of diseases is based on their antioxidant capacity and regulatory function in the human immune system.

1. *In vitro* studies for evaluation functionality of anthocyanin

In vitro testing of anthocyanin health benefits for humans is a scientific approach that investigates the biological effects and potential therapeutic properties of anthocyanin compounds isolated from different sources (grapes or pomace) through experiments conducted in a laboratory setting. During *in vitro* testing, anthocyanins are isolated from their sources and exposed to controlled conditions to evaluate their interactions with various cellular and molecular components (Corrêa *et al.*, 2017; Lingua *et al.*, 2018; Pešić *et al.*, 2019). The health benefits of the grape and the mechanisms of anthocyanin activity are analysed, improving the understanding of the correlation between the grape, anthocyanins and human health (Zhou *et al.*, 2022). As anthocyanins are sensitive to oxidation and susceptible to isomerisation, their bio-efficacy will depend on their bioavailability and bioaccessibility (Pešić *et al.*, 2019). Various factors, such as temperature, pH, light, food and gastrointestinal conditions (temperature, pH of digestive fluid and enzymes), influence the ability of anthocyanins to absorb, convert and metabolise (Moreno-Arribas & Polo, 2009; Milinčić *et al.*, 2022a; Milinčić *et al.*, 2023a). Several researchers have studied the effect of simulated digestion on the phenolic components of red grapes and

wines. Corrêa *et al.* (2017) investigated the in vitro-simulated digestion and bioactivity of bioactive compounds from Merlot variety grape pomace. Anthocyanin content decreased after gastrointestinal digestion. After cologne digestion, it did not change. However, characteristic metabolites originated from anthocyanins in the colon phase (Corrêa *et al.*, 2017). According to some researchers, anthocyanins from grapes and wine are less affected by the human gastrointestinal tract, which explains their high antioxidant capacity (Lingua *et al.*, 2018). These results are consistent with previous studies by the same group of authors, who reported that changes that occur from grape to wine during the winemaking process are responsible for the differences in phenolic content, especially anthocyanin concentration and antioxidative capacity. Therefore, grape will have higher antioxidative capacity due to its content in kaempferol-3-O-glucoside and ferulic acid, and wine and pomace will have lower antioxidative capacity due to the presence of ethyl-gallate (Lingua *et al.*, 2016).

2. *In vivo* studies for evaluation functionality of anthocyanin

The *in vivo* evaluation of the effects of anthocyanins on human health has very often been conducted on a living organism, whether animal or human, to study physiological effects, mechanisms of action and any health benefits associated with the consumption of anthocyanins. Grimes *et al.* (2018) investigated the effect of anthocyanins from table grape on cancer cell inhibition, and the results showed inhibition of cancer cells in three human cancer cell lines. Anthocyanins from both grape and red wine have beneficial effects on human health (Grimes *et al.*, 2018). In their review regarding the health benefits of grape bioactive molecules, Sabra *et al.* (2021) noted that whole grapes and their derivatives may have blood pressure-lowering potential. Meanwhile, in their review of the chemical properties of anthocyanins and their health effects on cardiovascular and neurodegenerative diseases, Mattioli *et al.* (2020) pointed out their protective effects on CVDs and neurodegenerative diseases. The protective effects of anthocyanins for both diseases are related to their antioxidant and anti-inflammatory properties (Kyrleou *et al.*, 2016b). In a recent study, Radeka *et al.* (2022) investigated the bioactive compounds of Croatian white and red wines and the effect of their consumption on human health. The results showed that regular moderate wine consumption (200 mL per day) decreases systolic and diastolic blood pressure, total cholesterol and LDL. Ferrer-Gallego and Silva (2022) suggested that food enriched with wine by-products can prevent chronic human diseases, since they are rich in anthocyanins, phenolic acids, flavonols, proanthocyanidins and stilbenes. Many researchers have worked on the estimation and valorization of the health benefit potentials of new functional foods as detailed in a comprehensive review by Iuga and Mironeasa (2020). According to Teissedre *et al.* (2018) in their review, the moderate and regular consumption of wine has cardioprotective effects due to anthocyanins and other polyphenolic compounds. It is thus necessary to find the best way to preserve the stability of anthocyanins and to increase their bioavailability so that they can be used to the maximum against various human diseases.

CONCLUSION

Although anthocyanins from red grape and wine have been widely investigated, this is the first comprehensive overview on their biosynthesis and transformations during winemaking and wine aging, their effects on sensorial properties and application in the food industry, and the valorization in order to benefit human health.—Anthocyanins are red grape and wine pigments that are responsible for wine colour. In our in-depth analysis of winemaking processes, we have unravelled the complex biochemical transformations undergone by anthocyanins during fermentation and maceration, offering insights into factors influencing their extraction and stability, and co-pigmentation and polymerization in wines. Being very susceptible to changes, they undergo numerous transformations during wine aging that influence colour intensity, colour density and hue. They influence the sensorial properties of wine by interacting with salivary proteins during wine tasting. The increasing application of anthocyanins in the food industry as natural colorants and functional ingredients highlight their versatility and potential for enhancing the aesthetic and nutritional qualities of various food products. Our investigation into the health benefits of anthocyanins reveals their promising therapeutic potential due to their antioxidant, anti-inflammatory and cardioprotective properties. This not only substantiates the significance of the role of anthocyanins in preventive healthcare but also paves the way to novel interventions that address a spectrum of human health conditions. In summary, this comprehensive review synthesises current knowledge on grape, wine and pomace anthocyanins in the fields of plant physiology, winemaking science, food technology and human health. The findings discussed here serve as a foundation for future research endeavours, inspiring further exploration into the intricate interplay of anthocyanins across disciplines.

ACKNOWLEDGEMENTS

This work was supported by the Ministry of Science, Technological Development and Innovation of the Republic of Serbia, Grant No. 451-03-65/2024-03/200116 and the Science Fund of the Republic of Serbia, #GRANT No. 7744714, FUNPRO.

The PhD of K. Delić is supported by a scholarship from the French Embassy in Serbia/French Institute (French Ministry of Foreign Affairs) 2021–2024.

REFERENCES

- Adali, M. B., Barresi, A. A., Boccardo, G., & Pisano, R. (2020). Spray Freeze-Drying as a Solution to Continuous Manufacturing of Pharmaceutical Products in Bulk. *Processes*, 8, 709. <https://doi.org/10.3390/pr8060709>
- Aguilar, T., Loyola C, De Bruijn J, Bustamante L, Vergara C, Von Baer D, Mardones C, & Serra I. (2016). Effect of thermomaceration and enzymatic maceration on phenolic compounds of grape must enriched by grape pomace, vine leaves, and canes. *Eur Food Res Technol*, 242, 1149–1158. <https://doi.org/10.1007/s00217-015-2619-3>

- Aleixandre-Tudo, J. L., & Du Toit, W. (2018). Cold maceration application in red wine production and its effects on phenolic compounds: A review. *LWT*, 95, 200–208. <https://doi.org/10.1016/j.lwt.2018.04.096>
- Alencar, N. M. M., Cazarin, C. B. B., Correa, L. C., Marostica Junior, M. R., Telles Biasoto, A. C., & Herman Behrens, J. (2017). Influence of maceration time on phenolic compounds and antioxidant activity of the Syrah must and wine. *Journal of Food Biochemistry*, 42, e12471. <https://doi.org/10.1111/jfbc.12471>
- Alfenito, M. R., Souer, E., Goodman, C. D., Buell, R., Mol, J., Koes, R., & Walbot, V. (1998). Functional Complementation of Anthocyanin Sequestration in the Vacuole by Widely Divergent Glutathione S-Transferases. *The Plant Cell*, 10(7), 1135–1149. <https://doi.org/10.1105/tpc.10.7.1135>
- Almasi, H., Forghani, S., & Moradi, M. (2022). Recent advances on intelligent food freshness indicators; an update on natural colorants and methods of preparation. *Food Packaging and Shelf Life*, 32, 100839. <https://doi.org/10.1016/j.foodchem.2022.100839>
- Alvarez Gaona, I. J., Fanzone, M. L., Galmarini, M. V., Chirife, J., Ferreras-Charro, R., García-Estévez, I., & Escribano-Bailón, M. T. (2022). Encapsulation of phenolic compounds by spray drying of Ancellotta and Aspirant Bouchet wines to produce powders with potential use as natural food colorants. *Food Bioscience*, 50 (102093). <https://doi.org/10.1016/j.fbio.2022.102093>
- Apolinar-Valiente, R., Romero-Cascales, I., Gomez-Plaza, E., & Ros-Garcia, J. M. (2016). Degradation of Syrah and Cabernet Sauvignon grapes skin: application of different enzymatic activities: a preliminary study. *European Food Research and Technology*, 242, 2041–2049. <https://doi.org/10.1007/s00217-016-2702-4>
- Arozarena, I., Ayestaran, B., Jesus-Castalejo, M., Navarro, M., Vera, M., Abril, I., & Casp, A. (2002). Anthocyanin composition of Tempranillo, Garnacha and Cabernet Sauvignon grapes from high- and low-quality vineyards over two years. *Eur Food Res Technol*, 214, 303–309. <https://doi.org/10.1007/s00217-001-0471-0>
- Atay, E., Fabra, M. J., Martinez-Sanz, M., Gomez-Mascaraque, L. G., Altan, A., & Lopez-Rubio, A. (2018). Development and characterization of chitosan/gelatin electrosprayed microparticles as food grade delivery vehicles for anthocyanin extracts. *Food Hydrocolloids*, 77, 699–710. <https://doi.org/10.1016/j.foodhyd.2017.11.011>
- Azuma, A. (2018). Genetic and Environmental Impacts on the Biosynthesis of Anthocyanins in Grapes. *The Horticulture Journal*, 87, 1–17. <https://doi.org/10.2503/hortj.OKD-IR02>
- Baek, H., Sanjays, Park, M., & Lee, H.-J. (2023). Cyanidin-3-O-glucoside protects the brain and improves cognitive function in APPswe/PS1ΔE9 transgenic mice model. *J Neuroinflammation* 20, (268). <https://doi.org/10.1186/s12974-023-02950-3>
- Bao, Y., Wang, M., Li, J., Yang, X., Wen, B., Wang, L., He, Y., Si, X., & Li, B. (2024). A starch-based 3D printed intelligent colorimetric film co-loaded natural pigments for visualizing food freshness: Effect of nozzle size on gel structure formation. *Food Hydrocolloids*, 155, 110218. <https://doi.org/10.1016/j.foodhyd.2024.110218>
- Bautista-Ortín, A. B., Fernández-Fernández, J. I., López-Roca, J. M., & Gómez-Plaza, E. (2004). Wine-making of High Coloured Wines: Extended Pomace Contact and Run-off of Juice Prior to Fermentation. *Food sci technol int*, 10, 287–295. <https://doi.org/10.1177/1082013204047565>
- Bautista-Ortín, A. B., Jimenez-Martinez, M. D., Jurado, R., Iniesta, J. A., Terrades, S., Andres, A., & Gomez-Plaza, E. (2017). Application of high-power ultrasounds during red wine vinification. *International Journal of Food Science and Technology*, 52, 1314–1323. <https://doi.org/10.1111/ijfs.13411>
- Bayram, H. M., Iliaz, R., & Gunes, F. E. (2024). Effects of Cornus mas L. on anthropometric and biochemical parameters among metabolic associated fatty liver disease patients: Randomized clinical trial. *Journal of Ethnopharmacology*, 318, 117068. <https://doi.org/10.1016/j.jep.2023.117068>
- Bennick, A. (1982). Salivary proline-rich proteins. *Mol Cell Biochem.*, 45, 83–99. <https://doi.org/10.1007/bf00223503>
- Beres, C., Pereira Freitas, S., de Oliveira Godoy, R., Rodrigues de Oliveira, D. C., Deliza, R., Iacomini, M., Mellinger-Silva, C., & Correa Cabral, L. (2019). Antioxidant dietary fibre from grape pomace flour or extract: Does it make any difference on the nutritional and functional value? *Journal of Functional Foods*, 56, 276–285. <https://doi.org/10.1016/j.jff.2019.03.014>
- Bianchi, A., Santini, G., Piombino, P., Pittari, E., Sanmartin, C., Moio, L., Modesti, M., Bellincontro, A., & Mencarelli, F. (2023). Nitrogen maceration of wine grape: An alternative and sustainable technique to carbonic maceration. *Food Chemistry*, 404, 134138. <https://doi.org/10.1016/j.foodchem.2022.134138>
- Boulton, R. (2001). The Copigmentation of Anthocyanins and Its Role in the Color of Red Wine: A Critical Review. *Am J Enol Vitic*, 52, 67–87. <https://doi.org/10.5344/ajev.2001.52.2.67>
- Breslin, P. A. S., Gilmore, M. M., Beauchamp, G. K., & Green, B. G. (1993). Psychophysical evidence that oral astringency is a tactile sensation. *Chem Senses*, 18, 405–417. <https://doi.org/10.1093/chemse/18.4.405>
- Brezoiu, A.-M., Matei, C., Deaconu, M., Stanciuc, A.-M., Trifan, A., Gaspar-Pintilie, A., & Berger, D. (2019). Polyphenols extract from grape pomace. Characterization and valorisation through encapsulation into mesoporous silica-type matrices. *Food and Chemical Toxicology*, 133, 110787. <https://doi.org/10.1016/j.fct.2019.110787>
- Brouillard, R., Iacobucci, G. A., & Sweeny, J. G. (1982). Chemistry of anthocyanin pigments. 9. UV-visible spectrophotometric determination of the acidity constants of apigeninidin and three related 3-deoxyflavylium salts. *J Am Chem Soc*, 104, 7585–7590. <https://doi.org/10.1021/ja00390a033>
- Cano-Lopez, M., Lopez-Roca, J. M., Pardo-Minguez, F., & Gomez Plaza, E. (2010). Oak barrel maturation vs. micro-oxygenation: Effect on the formation of anthocyanin-derived pigments and wine colour. *Food Chemistry*, 119, 191–195. <https://doi.org/10.1016/j.foodchem.2009.06.018>
- Carew, A. L., Gill, W., Close, D. C., & Damberg, R. G. (2014). Microwave Maceration with Early Pressing Improves Phenolics and Fermentation Kinetics in Pinot noir. *Am. J. Enol. Vitic.*, 65, 401–406. <https://doi.org/10.5344/ajev.2014.13089>
- Casassa, L. F., Bolcato, E. A., Sari, S. E., Fanzone, M. L., & Jofre, V. P. (2016). Combined effect of prefermentative cold soak and SO₂ additions in Barbera D’Asti and Malbec wines: Anthocyanin composition, chromatic and sensory properties. *LWT – Food Science and Technology*, 66, 134–142. <https://doi.org/10.1016/j.lwt.2015.10.026>
- Casassa, L. F., Sari, S., Bolcato, E., Diaz-Sambueza, M. A., Catania, A. A., Fanzone, M. L., Raco, F., & Barda, N. (2019). Chemical and Sensory Effect of Cold Soak, Whole Cluster Fermentation, and Stem Additions in Pinot noir Wines. *Am. J. Enol. Vitic.*, 70, <https://doi.org/10.5344/ajev.2018.18014>
- Castellanos-Gallo, L., Ballinas-Casarrubias, L., Espinoza-Hicks, J. C., Hernández-Ochoa, L. R., Muñoz-Castellanos, L. N., Zermeño-Ortega, M. R., Borrego-Loya, A., & Salas, E. (2022). Grape Pomace Valorization by Extraction of Phenolic Polymeric Pigments: A Review. *Processes*, 10, 469. <https://doi.org/10.3390/pr10030469>
- Chassaing, S., Lefeure, D., Jacquet, R., Jourdes, M., Ducasse, L., Galland, S., Grelard, A., Saucier, C., Teissedre, P.-L., Dangles, O., & Quideau, S. (2010). Physicochemical Studies of New Anthocyanano-Ellagitannin Hybrid Pigments: About the Origin of the Influence of Oak C-Glycosidic Ellagitannins on Wine Color. *European Journal of Organic Chemistry*, 29, 1680. <https://doi.org/10.1002/ejoc.200901133>

- Chinnici, F, Sonni, F, Natali, N, Galassi, S, & Riponi, C. (2009). Colour features and pigment composition of Italian carbonic macerated red wines. *Food Chemistry*, 113, 651–657. <https://doi.org/10.1016/j.foodchem.2008.07.055>
- Chira, K, Pacella, N, Jourdes, M, & Teissedre, P-L. (2011). Chemical and sensory evaluation of Bordeaux wines (Cabernet-Sauvignon and Merlot) and correlation with wine age. *Food Chemistry*, 126, 1971–1977. <https://doi.org/10.1016/j.foodchem.2010.12.056>
- Chira, K, Jourdes, M, & Teissedre, P-L. (2012). Cabernet sauvignon red wine astringency quality control by tannin characterization and polymerization during storage. *Eur Food Res Technol*, 234, 25. <https://doi.org/10.1007/s00217-011-1627-1>
- Christofi, S, Malliaris, D, Katsaros, G, Panagou, E, & Kallithraka, S. (2020). Limit SO₂ content of wines by applying High Hydrostatic Pressure. *Innovative Food Science and Emerging Technologies*, 62, 102342. <https://doi.org/10.1016/j.ifset.2020.102342>
- Ćirković, D, Matijašević, S, Ćirković, B, Laketić, D, Jovanović, Z, Kostić, B, Bešlić, Z, Sredojević, M, Tešić, Ž, Banjanac, T, & Gašić, U. (2022). Influence of Different Defoliation Timings on Quality and Phenolic Composition of the Wines Produced from the Serbian Autochthonous Variety Prokupac (*Vitis vinifera* L.). *Horticulturae*, 8, 296. <https://doi.org/10.3390/horticulturae80402963-261>
- Clifford, M. N. (2000). Anthocyanins – nature, occurrence and dietary burden. *Journal of the Science of Food and Agriculture*, 80, 1063–1072. [https://doi.org/10.1002/\(SICI\)1097-0010\(20000515\)80:7<1063::AID-JSFA605>3.0.CO;2-Q](https://doi.org/10.1002/(SICI)1097-0010(20000515)80:7<1063::AID-JSFA605>3.0.CO;2-Q)
- Corrêa, R. C. G, Haminiuk, C. W. I, Barros, L, Dias, M. I, Calhelha, R. C, Kato, C. G, Correa, V. G, Peralta, R. M, & Ferreira, I. C. F. R. (2017). Stability and biological activity of Merlot (*Vitis vinifera*) grape pomace phytochemicals after simulated in vitro gastrointestinal digestion and colonic fermentation. *Journal of Functional Foods*, 36, 410–417. <https://doi.org/10.1016/j.jff.2017.07.030>
- Costa, E, Cosme, F, Jordão, A. M, & Mendes-Faia, A. (2014). Anthocyanin profile and antioxidant activity from 24 grape varieties cultivated in two Portuguese wine regions. *OENO One*, 48, 51. <https://doi.org/10.20870/oeno-one.2014.48.1.1661>
- Crang, R, Lyons-Sobaski, S, & Wise, R. (2018). Plant Anatomy: A Concept-Based Approach to the Structure of Seed Plants. *Springer International Publishing, Cham*, XVI, 725. <https://doi.org/10.1007/978-3-319-77315-5>
- Ćurko, N, Ganić, K. K, Tomašević, M, Gracin, L, Jourdes, M, & Teissedre, P-L. (2021). Effect of enological treatments on phenolic and sensory characteristics of red wine during aging: Micro-oxygenation, sulfur dioxide, iron with copper and gelatin fining. *Food Chemistry*, 339, 127848. <https://doi.org/10.1016/j.foodchem.2020.127848>
- Cutanda-Perez, M-C, Ageorges, A, Gomez, C, Violet, S, Terrier, N, Romieu, C, & Torregrosa, L. (2009). Ectopic expression of VmybA1 in grapevine activates a narrow set of genes involved in anthocyanin synthesis and transport. *Plant Mol Biol*, 69, 633–648. <https://doi.org/10.1007/s11103-008-9446-x>
- Da Silva, D. J, de Oliveira, M. M, Wang, S. H, Carastan, D. J, & Rosa, D. S. (2022). Designing antimicrobial polypropylene films with grape pomace extract for food packaging. *Food Packaging and Shelf Life*, 34, 100929. <https://doi.org/10.1016/j.foodchem.2022.100929>
- De Souza, V. B, Thomazini, M, Balieiro, J. C. de C, & Fávoro-Trindade, C. S. (2015). Effect of spray drying on the physicochemical properties and color stability of the powdered pigment obtained from vinification byproducts of the Bordo grape (*Vitis labrusca*). *Food and Bioprocess Technology*, 93, 39–50. <https://doi.org/10.1016/j.fbp.2013.11.001>
- De Moura, S. C. S. R, Berling, C. L, Germer, S. P. M, Alvim, I. D, & Hubinger, M. D. (2018). Encapsulating anthocyanins from *Hibiscus sabdariffa* L. Calyces by ionic gelation: Pigment stability during storage of microparticles. *Food Chemistry*, 241, 317–327. <http://dx.doi.org/10.1016/j.foodchem.2017.08.095>
- Delić, K, Payan, C, Aleksovych, V, Jouin, A, Vignault, A, Chira, K, Jourdes, M, & Teissedre, P-L. (2023). Wine Phenolic Compounds: Chemistry and Biological Properties. Natural Products in Beverages. *Springer International Publishing, Cham*, 1–47. https://doi.org/10.1007/978-3-031-04195-2_218-1
- Della Vedova, L, Ferrario, G, Gado, F, Altomare, A, Carini, M, Morazzoni, P, Aldini, G, & Baron, G. (2022). Liquid Chromatography–High-Resolution Mass Spectrometry (LC–HRMS) Profiling of Commercial Enocianina and Evaluation of Their Antioxidant and Anti-Inflammatory Activity. *Antioxidants*, 11, 1187. <https://doi.org/10.3390/antiox11061187>
- Đorđević, V, Balanč, B, Belščak-Cvitanović, A, Lević, S, Trifković, K, Kalušević, A, Kostić, I, Komes, D, Bugarski, B, & Nedović, V. (2015). Trends in Encapsulation Technologies for Delivery of Food Bioactive Compounds. *Food Eng Rev*, 7, 452–490. <https://doi.org/10.1007/s12393-014-9106-7>
- Đorđević, N, Novaković, M, Pejini, B, Živković, M, Savić, A, Mutić, J, & Tešević, V. (2018). An insight into chemical composition and biological activity of Montenegrin Vranac red wine. *Scientia Horticulturae*, 230, 142–148. <https://doi.org/10.1016/j.scienta.2017.11.033>
- Ehrhardt, C, Arapitsas, P, Stefanini, M, Flick, G, & Mattivi, F. (2014). Analysis of the phenolic composition of fungus-resistant varieties cultivated in Italy and Germany using UHPLC-MS/MS. *Journal of Mass Spectrometry*, 49, 860–869. <https://doi.org/10.1002/jms.3440>
- El Darra, N, Turk, M. F, Ducasse, M-A, Grimi, N, Maroun, R. G, Louka, N, & Vorobiev, E. (2016). Changes in polyphenol profiles and color composition of freshly fermented model wine due to pulsed electric field, enzymes and thermovinification pretreatments. *Food Chemistry*, 194, 944–950. <https://doi.org/10.1016/j.foodchem.2015.08.059>
- Enache, M. I, Vasile, M. A, Craciunescu, O, Prelipcean, A. M, Oancea, A, Enachi, E, Barbu, V. V, Stanciu, N, & Vizireanu, C. (2022). Co-Microencapsulation of Anthocyanins from Cornelian Cherry (*Cornus mas* L.) Fruits and Lactic Acid Bacteria into Antioxidant and Anti-Proliferative Derivative Powders. *Nutrients*, 14, 3458. <https://doi.org/10.3390/nu14173458>
- Escot, S, Feuillat, M, Dulau, L, & Charpentier, C. (2001). Release of polysaccharides by yeasts and the influence of released polysaccharides on colour stability and wine astringency. *Australian Journal of Grape and Wine Research*, 7, 153–159. <https://doi.org/10.1111/j.1755-0238.2001.tb00204.x>
- FAO. (2022). Food and Agriculture Organization Corporate Statistical Database.
- Fanzone, M, Coronado, I, Sari, S, Catania, A, Gil I Cortiella, M, Assof, M, Jofré, V, Ubeda, C, & Peña-Neira, A. (2022). Microwave-assisted maceration and stems addition in Bonarda grapes: Effects on wine chemical composition over two vintages. *Food Research International*, 156, 111169. <https://doi.org/10.1016/j.foodres.2022.111169>
- Fernandes, A, Rocha, M. A. A, Santos, L. M. N. B. F, Bras, J, Oliveira, J, Mateus, N, & de Freitas, V. (2018). Blackberry anthocyanins: - β-Cyclodextrin fortification for thermal and gastrointestinal stabilization. *Food Chemistry*, 245, 426–431. <http://dx.doi.org/10.1016/j.foodchem.2017.10.109>
- Ferreiro-González, M, Ruiz-Rodríguez, A, Barbero, G. F, Ayuso, J, Álvarez, J. A, Palma, M, & Barros, C. G. (2019). FT-IR, Vis spectroscopy, color and multivariate analysis for the control of ageing processes in distinctive Spanish wines. *Food Chemistry*, 277, 6–11. <https://doi.org/10.1016/j.foodchem.2018.10.087>
- Ferrer-Gallego, R, Hernández-Hierro, J. M, Rivas-Gonzalo, J. C, & Escribano-Bailón, M. T. (2014). Sensory evaluation of bitterness and astringency sub-qualities of wine phenolic compounds: synergistic effect and modulation by aromas. *Food Research International*, 62, 1100–1107. <https://doi.org/10.1016/j.foodres.2014.05.049>

- Ferrer-Gallego, R., Soares, S., Mateus, N., Rivas-Gonzalo, J., Escribano-Bailón, M. T., Freitas, V. de (2015). New Anthocyanin–Human Salivary Protein Complexes. *Langmuir*, 31, 8392–8401. <https://doi.org/10.1021/acs.langmuir.5b01122>
- Ferrer-Gallego, R., & Silva, P. (2022). The Wine Industry By-Products: Applications for Food Industry and Health Benefits. *Antioxidants*, 11, 2025. <https://doi.org/10.3390/antiox11102025>
- Forino, M., Cassiano, C., Gambuti, A., Picariello, L., Aversano, R., Villano, C., Basile, B., Moio, L., & Frusciante, L. (2022). *ASC Food Sci. Technol.*, 2, 638–646. <https://doi.org/10.1021/acfoodsctech.1c00437>
- Forghani, S., Almasi, H., & Moradi, M. (2021). Electrospun nanofibers as food freshness and time-temperature indicators: A new approach in food intelligent packaging. *Innovative Food Science & Emerging Technologies*, 73, 102804. <https://doi.org/10.1016/j.ifset.2021.102804>
- Forghani, S., & Almasi, H. (2024). Characterization and performance evaluation of colorimetric pH-sensitive indicator based on K-carrageenan/quince seed mucilage hydrogel as freshness/spoilage monitoring of rainbow trout fillet. *Food Chemistry*, 457, 140072. <https://doi.org/10.1016/j.foodchem.2024.140072>
- Gao, R., Hu, H., Shi, T., Bao, Y., Sun, Q., Wang, L., Ren, Y., Jin, W., & Yuan, L. (2022). Incorporation of gelatin and Fe²⁺ increases the pH-sensitivity of zein-anthocyanin complex films used for milk spoilage detection. *Current Research in Food Science*, 5, 677–686. <https://doi.org/10.1016/j.crf.2022.03.016>
- Garcia, L., Martet, E., Suc, L., Garcia, F., & Saucier, C. (2024). Analysis of targeted phenolic ageing markers in Syrah red wines during bottle ageing: Influence of cork oxygen transfer rate. *Food Chemistry*, 434, 138491. <https://doi.org/10.1016/j.foodchem.2024.138491>
- García-Beneytez, E., Cabello, F., & Revilla, E. (2003). Analysis of Grape and Wine Anthocyanins by HPLC-MS. *J Agric Food Chem*, 51, 5622–5629. <https://doi.org/10.1021/jf0302207>
- García-Estévez, I., Ramos-Pineda, A. M., & Escribano-Bailón, M. T. (2018). Interactions between wine phenolic compounds and human saliva in astringency perception. *Food Funct*, 9, 1294–1309. <https://doi.org/10.1039/C7FO02030A>
- García-Lomillo, J., & González-SanJosé, M. L. (2017). Applications of Wine Pomace in the Food Industry: Approaches and Functions. *Comprehensive Reviews in Food Science and Food Safety*, 16, 3–22. <https://doi.org/10.1111/1541-4337.12238>
- Gawel, R., Oberholster, A., & Francis, I. L. (2000). A ‘Mouth-feel Wheel’ terminology for communicating the mouth-feel characteristics of red wine. *Australian Journal of Grape and Wine Research*, 6, 203–207. <https://doi.org/10.1111/j.1755-0238.2000.tb00180.x>
- Ghosh, S., Sarkar, T., Das, A., & Chakraborty, R. (2021). Micro and Nanoencapsulation of Natural Colors: a Holistic View. *Appl Biochem Biotechnol*, 193, 3787–3811. <https://doi.org/10.1007/s12010-021-03631-8>
- Giacosa, S., Ferrero, L., Pissoni, M. A., Rio Segade, S., Gerbi, V., & Rolle, L. (2023). Grape skin anthocyanin extraction from red varieties during stimulated maceration: Influence of grape seeds and pigments adsorption on their surface. *Food Chemistry*, 424, 136463. <https://doi.org/10.1016/j.foodchem.2023.136463>
- Gibbins, H. L., & Carpenter, G. H. (2013). Alternative Mechanisms of Astringency – What is the Role of Saliva? *J Texture Stud*, 44, 364–375. <https://doi.org/10.1111/jtxs.12022>
- Gil, M., Kontoudakis, N., Gonzalez, E., Esteruelas, M., Fort, F., Canals, J. M., & Zamora, F. (2012). Influence of Grape Maturity and Maceration Length on Color, Polyphenolic Composition, and Polysaccharide Content of Cabernet Sauvignon and Tempranillo Wines. *Journal of Agricultural and Food Chemistry*, 60, 7988–8001. <https://doi.org/10.1021/jf302064n>
- Girard, B., Kopp, T. G., Reynolds, A. G., & Cliff, M. (1997). Influence of Vinification Treatments on Aroma Constituents and Sensory Descriptors of Pinot noir Wines. *Am J Enol Vitic*, 48, 198–206. <https://doi.org/10.5344/ajev.1997.48.2.198>
- Gomez-Brandon, M., Martinez-Cordeiro, H., & Dominguez, J. (2021). Changes in the nutrient dynamics and microbiological properties of grape marc in a continuous-feeding vermicomposting system. *Waste Management*, 135, 1–10. <https://doi.org/10.1016/j.wasman.2021.08.004>
- González-Neves, G., Gil, G., Barreiro, L., & Favre, G. (2010). Pigment profile of red wines cv. Tannat made with alternative winemaking techniques. *Journal of Food Composition and Analysis*, 23, 447–454. <https://doi.org/10.1016/j.jfca.2009.08.021>
- Gonzalez-Saiz, J. M., Esteban-Diez, I., Rodriguez-Tecedor, S., Perez-del-Notario, N., Arezana-Ramila, & Pizarro, C. (2014). Modulation of the phenolic composition and colour of red wines subjected to accelerated ageing by controlling process variables. *Food Chemistry*, 165, 271–281. <http://dx.doi.org/10.1016/j.foodchem.2014.05.016>
- Gordillo, B., Rivero, F. J., Jara-Palacios, M. J., González-Miret, M. L., & Heredia, F. J. (2021). Impact of a double post-fermentative maceration with ripe and overripe seeds on the phenolic composition and color stability of Syrah red wines from warm climate. *Food Chemistry*, 346, 128919. <https://doi.org/10.1016/j.foodchem.2020.128919>
- Guerrero, F. R., Liazid, A., Palma, M., Puertas, B., Gonzalez-Barrio, R., Gil-Izquierdo, A., Garcia-Barroso, C., & Cantos-Villar, E. (2009). Phenolic characterisation of red grapes autochthonous to Andalusia. *Food Chemistry*, 112, 949–955. <https://doi.org/10.1016/j.foodchem.2008.07.014>
- Green, B. G. (1993). Oral astringency: A tactile component of flavor. *Acta Psychologica*, 84, 119–125. [https://doi.org/10.1016/0001-6918\(93\)90078-6](https://doi.org/10.1016/0001-6918(93)90078-6)
- Grimes, K. L., Stuart, C. M., McCarthy, J. J., Kaur, B., Cantu, E. J., & Forester, S. C. (2018). Enhancing the Cancer Cell Growth Inhibitory Effects of Table Grape Anthocyanins. *Journal of Food Science*, 83, 2369–2374. <https://doi.org/10.1111/1750-3841.14294>
- Grotewold, E. (2004). The challenges of moving chemicals within and out of cells: insights into the transport of plant natural products. *Planta*, 219. <https://doi.org/10.1007/s00425-004-1336-0>
- Grotewold, E. (2006). The science of flavonoids. *Springer*, 1, VIII, 274. <https://doi.org/10.1007/978-0-387-28822-2>
- Han, F-L, Zhang, W-N, Pan, Q-H, Zheng, C-R, Chen, H-Y, & Duan, C-Q. (2008). Principal Component Regression Analysis of the Relation Between CIELAB Color and Monomeric Anthocyanins in Young Cabernet Sauvignon Wines. *Molecules*, 13, 2859–2870. <https://doi.org/10.3390/molecules13112859>
- Hasan, S., Khatri, N., Rahman, Z. N., Menezes, A. A., Martini, J., Shehjar, F., Mujeeb, N., & Shah, Z. A. (2023). Neuroprotective Potential of Flavonoids in Brain Disorders. *Brain Sciences*, 13, 1258. <https://doi.org/10.3390/brainsci13091258>
- Haselgrove, L., Botting, D., van HEESWIJCK, R., Høj, P. B., Dry, P. R., Ford, C., & Land, P. G. I. (2000). Canopy microclimate and berry composition: The effect of bunch exposure on the phenolic composition of *Vitis vinifera* L cv. Shiraz grape berries. *Australian Journal of Grape and Wine Research* 6, 141–149. <https://doi.org/10.1111/j.1755-0238.2000.tb00173.x>
- He, F., Mu, L., Yan, G-L, Liang, N-N, Pan, Q-H, Wang, J., Reeves, M. J., & Duan, C-Q. (2010). Biosynthesis of Anthocyanins and Their Regulation in Colored Grapes. *Molecules*, 15, 9058–9091. <https://doi.org/10.3390/molecules15129057>

- Holton, T., & Cornish, E. (1995). Genetics and Biochemistry of Anthocyanin Biosynthesis. *Plant Cell* 7, 1071–1083. <https://doi.org/10.1105/tpc.7.7.1071>
- Iuga, M., & Mironeasa, S. (2020). Potential of grape byproducts as functional ingredients in baked goods and pasta. *Compr Rev Food Sci Food Saf.*, 19, 2473–2505. <https://doi.org/10.1111/1541-4337.12597>
- Hoss, I., Rajha, H., El Khoury, R., Youssef, S., Manca, M. L., Manconi, M., Louka, N., & Maroun, R. G. (2021). Valorization of Wine-Making By-Products' Extracts in Cosmetics. *Cosmetics*, 8, 109. <https://doi.org/10.3390/cosmetics8040109>
- Ivanova, V., Dörnyei, Á., Márk, L., Vojnoski, B., Stafilov, T., Stefova, M., & Kilár, F. (2011a). Polyphenolic content of Vranec wines produced by different vinification conditions. *Food Chemistry*, 124, 316–325. <https://doi.org/10.1016/j.foodchem.2010.06.039>
- Ivanova, V., Stefova, M., Vojnoski, B., Dörnyei, Á., Márk, L., Dimovska, V., Stafilov, T., & Kilár, F. (2011b). Identification of polyphenolic compounds in red and white grape varieties grown in R. Macedonia and changes of their content during ripening. *Food Research International*, 44, 2851–2860. <https://doi.org/10.1016/j.foodres.2011.06.046>
- OIV. (2017). Distribution of the world's grapevine varieties. *Focus OIV 2017*.
- OIV. (2024). State of the world vine and wine sector in 2023.
- Jagatić Korenika, A-M, Kozina, B, Preiner, D, Tomaz, I, Volarević, J, & Jeromel, A. (2023). The Effect of Seed Removal and Extraction Time on the Phenolic Profile of Plavac Mali Wine. *Applied Sciences*, 13, 5411. <https://doi.org/10.3390/app13095411>
- Ju, Y, Yang, L, Yue, X, Li, Y, He, R, Deng, S, Yang, X, & Fang, Y. (2021). Anthocyanin profiles and color properties of red wines made from *Vitis davidii* and *Vitis vinifera* grapes. *Food Science and Human Wellness*, 10, 335–344. <https://doi.org/10.1016/j.fshw.2021.02.025>
- Junqua, R, Carullo, D, Ferrari, G, Pataro, G, & Ghidossi, R. (2021). Ohmic heating for polyphenol extraction from grape berries: an innovative prefermentary process. *OENO One*, 3, 39-51. <https://doi.org/10.20870/oeno-one.2021.55.3.4647>
- Kammerer, D, Claus, A, Schieber, A, & Carle, R. (2005). A Novel Process for the Recovery of Polyphenols from Grape (*Vitis vinifera* L.) Pomace. *Journal of Food Science*, 70, 157–163. <https://doi.org/10.1111/j.1365-2621.2005.tb07077.x>
- Kanha, N, Surawang, S, Pitchakarn, P, & Laokuldilok, T. (2020). Microencapsulation of copigmented anthocyanins using double emulsion followed by complex coacervation: Preparation, characterization, and stability. *LWT*, 133, 110154. <https://doi.org/10.1016/j.lwt.2020.110154>
- Körösi, L, Molnár, S, Teszlák, P, Dörnyei, Á, Maul, E, Töpfer, R, Marosvölgyi, T, Szabó, É, & Röckel, F. (2022). Comparative Study on Grape Berry Anthocyanins of Various Teinturier Varieties. *Foods*, 11, 3668. <https://doi.org/10.3390/foods11223668>
- Kuchen, B, Vasquez, F, Mestre, M. V, Toro, M. E, & Maturano, Y. P. (2018). Influence of Cold Maceration Time on Chromatic and Microbiological Characteristics of Cabernet Sauvignon Wines. *Afr. J. Enol. Vitic.*, 39, 89-99. <http://dx.doi.org/10.21548/39-1-2460>
- Ky, I, Lorrain, B, Kolbas, N, Crozier, A, & Teissedre, P. L. (2014). Wine by-products: phenolic characterization and antioxidant activity evaluation of grapes and grape pomaces from six different French grape varieties. *Molecules (Basel, Switzerland)*, 19(1), 482–506. <https://doi.org/10.3390/molecules19010482>
- Kyraleou, M, Kotseridis, Y, Koundouras, S, Chira, K, Teissedre, P-L, & Kallithraka, S. (2016a). Effect of irrigation regime on perceived astringency and proanthocyanidin composition of skins and seeds of *Vitis vinifera* L. cv. Syrah grapes under semiarid conditions. *Food Chemistry*, 203, 292–300. <https://doi.org/10.1016/j.foodchem.2016.02.052>
- Kyraleou, M, Tzanakouli, E, Kotseridis, Y, Chira, K, Ligas, I, Kallithraka, S, & Teissedre, P-L. (2016b). Addition of wood chips in red wine during and after alcoholic fermentation: differences in color parameters, phenolic content and volatile composition. *OENO One*, 50. <https://doi.org/10.20870/oeno-one.2016.50.4.885>
- Labrador Fernandez, L, Perez-Porras, P, Diaz-Maroto, M. C, Gomez-Plaza, E, Perez-Coello, M. S, & Bautista-Ortin, A. B. (2023). The technology of high-power ultrasound and its effect on the color and aroma of rose wines. *Journal of The Science of Food and Agriculture*, 103, 6616-6624. <https://doi.org/10.1002/jsfa.12757>
- Lakićević, S. H, Karabegović, I. T, Cvetković, D. J, Lazić, M. L, Jančić, R, & Popović-Djordjević, J. B. (2022). Insight into the Aroma Profile and Sensory Characteristics of 'Prokupac' Red Wine Aromatised with Medicinal Herbs. *Horticulturae*, 8, 277. <https://doi.org/10.3390/horticulturae8040277>
- Lamikanra, O. (1989). Anthocyanins of *Vitis rotundifolia* Hybrid Grapes. *Food Chemistry*, 33, 225–237. [https://doi.org/10.1016/0308-8146\(89\)90016-2](https://doi.org/10.1016/0308-8146(89)90016-2)
- Lavelli, V, & Sri Harsha, P. S. C. (2019). Microencapsulation of grape skin phenolics for pH controlled release of antiglycation agents. *Food Research International*, 119, 822–828. <https://doi.org/10.1016/j.foodres.2018.10.065>
- Leong, S. Y, Treadwell, M, Liu, T, Hochberg, M, Sack, M, Mueller, G, Sigler, J, Silcock, P, & Oey, I. (2020). Influence of Pulsed Electric Fields processing at high-intensity electric field strength on the relationship between anthocyanin composition and colour intensity of Merlot (*Vitis vinifera* L.) musts during cold maceration. *Innovative Food Science and Emerging Technologies*, 59, 102243. <https://doi.org/10.1016/j.ifset.2019.102243>
- Li, B, Wang, L, Bai, W, Chen, W, Chen, F, & Shu, C. (2021). Anthocyanins: Chemistry, Processing & Bioactivity. *Springer Nature Singapore*, XVIII, 336. <https://doi.org/10.1007/978-0-387-77335-3>
- Li, L, Xu, Z, Yang, H, Zhao, W, Tao, Y, Lu, J, Xia, X, Tan, M, Du, J, & Wang, H. (2024). *In situ* encapsulation of anthocyanin within soybean protein isolate chains driven by pH for detecting the freshness of food. *Food Bioscience*, 61, 104688. <https://doi.org/10.1016/j.fbio.2024.104688>
- Lingua, M. S, Fabani, M. P, Wunderlin, D. A, & Baroni, M. V. (2016). From grape to wine: Changes in phenolic composition and its influence on antioxidant activity. *Food Chemistry*, 208, 228–238. <https://doi.org/10.1016/j.foodchem.2016.04.009>
- Lingua, M. S, Wunderlin, D. A, & Baroni, M. V. (2018). Effect of simulated digestion on the phenolic components of red grapes and their corresponding wines. *Journal of Functional Foods*, 44:86–94. <https://doi.org/10.1016/j.jff.2018.02.034>
- Liu, T, Qi, W, Peng, W, Zhang, J, & Wang, Y. (2024). Cyanidin-3-glucoside protects the photooxidative damage of retinal pigment epithelium cells by regulating sphingolipid signaling and inhibiting MAPK pathway. *Food Science and Human Wellness*, 13, 621–632. <https://doi.org/10.26599/FSHW.2022.9250053>
- Lević, S, Pajić Lijaković, I, Đorđević, V, Rac, V, Rakić, V, Šolević Knudsen, T, Pavlović, V, Bugarski, B, & Nedović, V. (2015). Characterization of sodium alginate/D-limonene emulsions and respective calcium alginate/D-limonene beads produced by electrostatic extrusion. *Food Hydrocolloids*, 45, 111-123. <http://dx.doi.org/10.1016/j.foodhyd.2014.10.001>
- López-Giral, N, López, R, Santamaria, P, Gonzalez-Areñana, L, & Garde-Cerdan, T. (2023). Phenolic and colour characteristics of must and wine obtained from red grapes treated by pulsed electric fields. Efficacy of PEF to reduce maceration time in elaboration of red wines. *European Food Research and Technology*, 249, 273-282. <https://doi.org/10.1007/s00217-022-04114-8>

- López, N, Puértolas, E, Condón, S, Álvarez, I, & Raso, J. (2008). Effects of pulsed electric fields on the extraction of phenolic compounds during the fermentation of must of Tempranillo grapes. *Innovative Food Science & Emerging Technologies*, 9, 477–482.
- Lorrain, B, Chira, K, & Teissedre, P-L. (2011). Phenolic composition of Merlot and Cabernet-Sauvignon grapes from Bordeaux vineyard for the 2009-vintage: Comparison to 2006, 2007 and 2008 vintages. *Food Chemistry*, 126, 1991–1999. <https://doi.org/10.1016/j.foodchem.2010.12.062>
- Lu, R, Song, M, Wang, Z, Zhai, Y, Hu, C, Perl, A, & Ma, H. (2023). Independent flavonoid and anthocyanin biosynthesis in the flesh of a red-fleshed table grape revealed by metabolome and transcriptome co-analysis. *BMC Plant Biol*, 23, 361. <https://doi.org/10.1186/s12870-023-04368-8>
- Lukić, I., Budić-Leto, I., Bubola, M., Damijanić, K., & Staver, M. (2017). Pre-fermentative cold mac-eration, saignée, and various thermal treatments as options for modulating volatile aroma and phenol profiles of red wine. *Food chemistry*, 224, 251–261. <https://doi.org/10.1016/j.foodchem.2016.12.077>
- Mao, M, Li, K, Chen, F, Hu, X, Ma, L, & Ji, J. (2024). Study on the interactions between oral mucin and cyanidin 3-O-glucoside: the effect of oxidized quinone. *Food Bioprocess Technol*, 17, 1335–1345. <https://doi.org/10.1007/s11947-023-03207-w>
- Mattioli, R, Francioso, A, Mosca, L, & Silva, P. (2020). Anthocyanins: A Comprehensive Review of Their Chemical Properties and Health Effects on Cardiovascular and Neurodegenerative Diseases. *Molecules*, 25, 3809. <https://doi.org/10.3390/molecules25173809>
- Maza, M, Alvarez, I, & Raso, J. (2019). Thermal and Non-Thermal Physical Methods for Improving Polyphenol Extraction in Red Winemaking. *Beverages*, 5, 47. <https://doi.org/10.3390/beverages5030047>
- Maza, M. A, Pereira, C, Martínez, J. M, Camargo, A, Álvarez, I, & Raso, J. (2020). PEF treatments of high specific energy permit the reduction of maceration time during vinification of Caladoc and Grenache grapes. *Innovative Food Science & Emerging Technologies*, 63, 102375. <https://doi.org/10.1016/j.ifset.2020.102375>
- Milinić, D.D., Popović, D.A., Lević, S.M., Kostić, A.Ž., Tešić, Ž.Lj., Nedović, V. A., Pešić, M.B. (2019). Application of polyphenol-loaded nanoparticles in food industry. *Nanomaterials*, 9, 1629. <https://doi.org/10.3390/nano9111629>
- Milinić, D.D., Kostić, A.Ž., Špirović-Trifunović, B.D., Tešić, Ž.Lj., Tosti, T.B., Dramićanin A.M., Barać, M.B., Pešić, M.B. (2020). Grape seed flour of different grape pomaces: Fatty acid profile, soluble sugar profile and nutritional value. *Journal of the Serbian Chemical Society*, 85(3), 305–319. <https://doi.org/10.2298/JSC190713117M>
- Milinić, D. D, Stanisavljević, N. S, Kostić, A. Ž, Soković Bajić, S, Kojić, M. O, Gašić, U. M, Barać, M. B, Stanojević, S. P, Lj Tešić, Ž, & Pešić, M. B. (2021a). Phenolic compounds and biopotential of grape pomace extracts from Prokupac red grape variety. *LWT*, 138, 110739. <https://doi.org/10.1016/j.lwt.2020.110739>
- Milinić, D. D, Kostić, A. Ž, Gašić, U. M, Lević, S, Stanojević, S. P, Barać, M. B, Tešić, Ž. Lj, Nedović, V, & Pešić M. B. (2021b). Skimmed Goat's Milk Powder Enriched with Grape Pomace Seed Extract: Phenolics and Protein Characterization and Antioxidant Properties. *Biomolecules*, 11, 965. <https://doi.org/10.3390/biom11070965>
- Milinić, D.D., Stanisavljević, N.S., Kostić, A.Ž., Gašić, U.M., Stanojević, S.P., Tešić, Ž.Lj., Pešić, M.B. (2022a). Bioaccessibility of phenolic compounds and antioxidant properties of goat-milk powder fortified with grape-pomace-seed extract after in vitro gastrointestinal digestion. *Antioxidants*, 11, 2164. <https://doi.org/10.3390/antiox11112164>
- Milinić, D.D., Salević, A., Kostić, A.Ž., Nedović, V.A., Pešić, M.B., (2022b). Improvement of physicochemical properties of food, functionality, quality, and safety by phytochemical-loaded nanoemulsions. In: Bio-based nanoemulsions for agri-food applications, Eds: Kamel A. Abd-Elsalam, Kasi Murugan, Chapter 17: 279–296. <https://doi.org/10.1016/B978-0-323-89846-1.00007-3>
- Milinić, D.D., Salević-Jelić, A.S., Kostić, A.Ž., Stanojević, S.P., Nedović, V., Pešić, M.B. (2023a). Food nanoemulsions: how simulated gastrointestinal digestion models, nanoemulsion, and food matrix properties affect bioaccessibility of encapsulated bioactive compounds. *Critical Reviews in Food Science and Nutrition*, 1–23. <https://doi.org/10.1080/10408398.2023.2195519>
- Milinić, D.D., Gašić, U., Kostić, A.Ž., Stanojević, S.P., Tešić, Ž.Lj., Pešić, M.B. (2023b). Grape seed powders as source of phenolic compounds: UHPLC Orbitrap MS4 characterisation. 14th European Nutrition Conference FENS 2023, 14–17 November 2023, Belgrade, Serbia. *Proceedings*, 91, 376. <https://doi.org/10.3390/proceedings2023091376>
- Milinić, D.D., Kostić, A.Ž., Kolašinac, S., Rac, V., Banjac, N., Ladarević, J., Lević, S., Pavlović, V.B., Stanojević, S.P., Nedović, V.A., Pešić, M.B. (2024). Goat milk powders enriched with grape pomace seed extract: Physical and techno-functional properties. *Food Hydrocolloids*, 146, 109293. <https://doi.org/10.1016/j.foodhyd.2023.109293>
- Mohammadalinejad, S, Almonaityte, A, Jensen, I-J, Kurek, M, & Lerfall, J. (2023). Alginate microbeads incorporated with anthocyanins from purple corn (*Zean mays L.*) using electrostatic extrusion: Microencapsulation optimization, characterization, and stability studies. *International Journal of Biological Macromolecules*, 246, 125684. <https://doi.org/10.1016/j.ijbiomac.2023.125684>
- Monteiro, G. C, Minatel, I. O, Junior, A. P, Gomez-Gomez, H. A, De Camargo, J. P. C, Diamante, M. S, Pereira Basílio, L. S, Tecchio, M. A, & Pereira Lima, G. P. (2021). Bioactive compounds and antioxidant capacity of grape pomace flours. *LWT*, 135, 110053. <https://doi.org/10.1016/j.lwt.2020.110053>
- Moreno-Arribas, M. V, & Polo, M. C. (2009). Wine Chemistry and Biochemistry. *Springer New York*, XV, 735, <https://doi.org/10.1007/978-0-387-74118-5>
- Morgani, M. B, Fanzone, M, Peña, J. E. P, Sari, S, Gallo, A. E, Tournier, M. G, & Prieto, J. A. (2023). Late pruning modifies leaf to fruit ratio and shifts maturity period, affecting berry and wine composition in *Vitis vinifera L.* cv. ‘Malbec’ in Mendoza, Argentina. *Scientia Horticulturae*, 313, 111861. <https://doi.org/10.1016/j.scienta.2023.111861>
- Muñoz, P, Pérez, K, Cassano, A, & Ruby-Figueroa, R. (2021). Recovery of Anthocyanins and Monosaccharides from Grape Marc Extract by Nanofiltration Membranes. *Molecules*, 26, 2003. <https://doi.org/10.3390/molecules26072003>
- McRae, J. M, Dambergs, R. G, Kassara, S, Parker, M, Jeffery, D. W, Herderich, M. J, & Smith, P. A. (2012). Phenolic Compositions of 50 and 30 Year Sequence of Australian Red Wines: The Impact of Wine Age. *Journal of Agricultural and Food Chemistry*, 60, 10093–10102. <https://dx.doi.org/10.1021/jf301571q>
- Nivetha, K, Vinotha, V, Albeshr, M. F, Mahboob, S, Manzoor, I, Govindarajan, M, & Vaseeharan, B. (2022). Synthesis and characterization of *Vitis vinifera* exocarp-mediated ZnO nanoparticles: An evaluation of biological potential and ecotoxicity. *Journal of Drug Delivery Science and Technology*, 77, 103846. <https://doi.org/10.1016/j.jddst.2022.103846>
- Norcino, L. B, Farinassi Mendes, J, de Abreu Figueiredo, J, Leite Oliveira, N, Alvarenga Botrel, D, & Capparelli Mattoso, L. H. (2022). Development of alginate/pectin microcapsules by a dual process combining emulsification and ultrasonic gelation for encapsulation and controlled release of anthocyanins from grapes (*Vitis labrusca L.*). *Food Chemistry*, 391, 133256. <https://doi.org/10.1016/j.foodchem.2022.133256>
- Ntuli, R. G, Saltman, Y, Ponangi, R, Jeffery, D. W, Bindon, K, Wilkinson, K. L. (2023). Impact of skin contact time, oak and tannin addition on the chemical composition, color stability and sensory profile of Merlot wines made from flash détente treatment. *Food Chemistry*, 405, 134849. <https://doi.org/10.1016/j.foodchem.2022.134849>

- Oberque-Slier, E, Pena-Neira, A, Lopez-Solis, R, Caceres-Mella, A, Toledo-Araya, H, & Lopez-Rivera, A. (2013). Phenolic composition of skins from four Carmenet grape varieties (*Vitis vinifera* L.) during ripening. *LWT – Food Science and Technology*, 54, 404–413. <https://doi.org/10.1016/j.lwt.2013.06.009>
- Oprea, O. B, & Gaceu, L. (2020). Consumer acceptance and sensorial analysis of bread with grape seed flour. *E3S Web of Conferences*, 215, 01005. <https://doi.org/10.1051/e3sconf/202021501005>
- Otteneder, H, Marx, R, & Zimmer, M. (2004). Analysis of the anthocyanin composition of Cabernet Sauvignon and Portugieser wines provides an objective assessment of grape varieties. *Australian Journal of Grape and Wine Research*, 10, 3-7. <https://doi.org/10.1111/j.1755-0238.2004.tb00002.x>
- Otto, T, Botelho, R, Biasi, L, Miljić, U. C, Correia, A, & M. Jordão, A. (2023). Adaptability of Different International Grape Varieties in Diverse Terroirs: Impact on Grape and Wine Composition. In: M. Jordão A, Botelho R, Miljić U (eds) Recent Advances in Grapes and Wine Production - New Perspectives for Quality Improvement. *IntechOpen*, <https://doi.org/10.5772/intechopen.100883>
- Pace, C, Giacosa, S, Torchio, F, Río Segade, S, Cagnasso, E, & Rolle, L. (2014). Extraction kinetics of anthocyanins from skin to pulp during carbonic maceration of winegrape berries with different ripeness levels. *Food Chemistry*, 165, 77–84. <https://doi.org/10.1016/j.foodchem.2014.05.070>
- Paissoni, M. A, Waffo-Teguo, P, Ma, W, Jourdes, M, Rolle, L, & Teissedre, P-L. (2018). Chemical and sensorial investigation of in-mouth sensory properties of grape anthocyanins. *Sci Rep*, 8, 17098. <https://doi.org/10.1038/s41598-018-35355-x>
- Paissoni, M. A, Waffo-Teguo, P, Ma, W, Jourdes, M, Giacosa, S, Río Segade, S, Rolle, L, & Teissedre, P-L. (2020). Sensory assessment of grape polyphenolic fractions: an insight into the effects of anthocyanins on in-mouth perceptions. *OENO One*, 54, 1059–1075. <https://doi.org/10.20870/oeno-one.2020.54.4.4142>
- Pantelić, M. M, Dabić Zagorac, D. Č, Davidović, S. M, Todić, S. R, Bešlić, Z. S, Gašić, U. M, Tešić, Ž. Lj, & Natić, M. M. (2016). Identification and quantification of phenolic compounds in berry skin, pulp, and seeds in 13 grapevine varieties grown in Serbia. *Food Chemistry*, 211, 243–252. <https://doi.org/10.1016/j.foodchem.2016.05.051>
- Papoušková, B, Bednář, P, Hron, K, Stávek, J, Balík, J, Myjavcová, R, Barták, P, Tománková, E, & Lemr, K. (2011). Advanced liquid chromatography/mass spectrometry profiling of anthocyanins in relation to set of red wine varieties certified in Czech Republic. *Journal of Chromatography A*, 1218, 7581–7591. <https://doi.org/10.1016/j.chroma.2011.07.027>
- Pereira, R. N, Coelho, M. I, Genisheva, Z, Fernandes, J. M, Vicente, A. A, Pintado, M. E, & Teixeira, J. A. (2020). Using Ohmic Heating effect on grape skin as a pretreatment for anthocyanins extraction. *Food and Bioprocess Technology*, 124, 320-328. <https://doi.org/10.1016/j.fbp.2020.09.009>
- Pereira Souza, A. C, Deyse Gurak, P, & Damasceno Ferreira Marczak, L. (2017). Maltodextrin, pectin, and soy protein isolate as carrier agents in the encapsulation of anthocyanins-rich extract from jaboticaba pomace. *Food and Bioprocess Technology*, 102, 186-194. <http://dx.doi.org/10.1016/j.fbp.2016.12.012>
- Peixoto, C. M, Dias, M. I, Alves, M. J, Calhelha, R. C, Barros, L, Pinho, S. P, & Ferreira, I. C. F. R. (2018). Grape pomace as a source of phenolic compounds and diverse bioactive properties. *Food Chemistry*, 253, 132–138. <https://doi.org/10.1016/j.foodchem.2018.01.163>
- Pérez-Navarro, J, Izquierdo-Cañas, P. M, Mena-Morales, A, Martínez-Gascuña, J, Chacón-Vozmediano, J. L, García-Romero, E, Hermosín-Gutiérrez, I, & Gómez-Alonso, S. (2019). Phenolic compounds profile of different berry parts from novel *Vitis vinifera* L. red grape genotypes and Tempranillo using HPLC-DAD-ESI-MS/MS: A varietal differentiation tool. *Food Chemistry*, 295, 350–360. <https://doi.org/10.1016/j.foodchem.2019.05.137>
- Pešić, M. B, Milinčić, D. D, Kostić, A. Ž, Stanisavljević, N. S, Vukotić, G. N, Kojić, M. O, Gašić, U. M, Barać, M. B, Stanojević, S. P, Popović, D. A, Banjac, N. R, & Tešić, Ž. Lj. (2019). In vitro digestion of meat- and cereal-based food matrix enriched with grape extracts: How are polyphenol composition, bioaccessibility and antioxidant activity affected? *Food Chemistry*, 284, 28–44. <https://doi.org/10.1016/j.foodchem.2019.01.107>
- Pires, M. A, Pastrana, L. M, Fuciños, P, Abreu, C. S, & Oliveira, S. M. (2020). Sensorial Perception of Astringency: Oral Mechanisms and Current Analysis Methods. *Foods*, 9, 1124. <https://doi.org/10.3390/foods9081124>
- Popović, D. A, Milinčić, D. D, Pešić, M. B, Kalušević, A. M, Tešić, Ž. Lj, & Nedović, V. A. (2019). Encapsulation technologies for polyphenol-loaded microparticles in food industry. In: Green Food Processing Techniques. *Elsevier*, 335–367. <https://doi.org/10.1016/B978-0-12-815353-6.00012-4>
- Portu, J, Rosa Gutiérrez-Viguera, A, González-Arenzana, L, & Santamaría, P. (2023). Characterization of the color parameters and monomeric phenolic composition of ‘Tempranillo’ and ‘Graciano’ wines made by carbonic maceration. *Food Chemistry*, 406, 134327. <https://doi.org/10.1016/j.foodchem.2022.134327>
- Qin, Y, Wang, Y, Tang, Z, Chen, K, Wang, Z, Cheng, G, Chi, H, & Soteyome, T. (2024). A pH-sensitive film based on chitosan/gelatin and anthocyanin from *Zingiber striolatum* Diels for monitoring fish freshness. *Food Chemistry: X*, 23, 101639. <https://doi.org/10.1016/j.fochx.2024.101639>
- Quaglieri, C, Prieto-Perea, N, Berrueta, L. A, Gallo, B, Rasines-Perea, Z, Jourdes, M, & Teissedre, P-L. (2017). Comparison of Aquitaine and Rioja Red Wines: Characterization of Their Phenolic Composition and Evolution from 2000 to 2013. *Molecules*, 22, 192. <https://doi.org/10.3390/molecules22020192>
- Radeka, S, Rossi, S, Bestulić, E, Budić-Leto, I, Kovačević Ganić, K, Horvat, I, Lukić, I, Orbanić, F, Zaninović Jurjević, T, & Dvornik, Š. (2022). Bioactive Compounds and Antioxidant Activity of Red and White Wines Produced from Autochthonous Croatian Varieties: Effect of Moderate Consumption on Human Health. *Foods*, 11, 1804. <https://doi.org/10.3390/foods11121804>
- Rajković, M, Popović-Minić, D, Milinčić, D, & Zdravković, M. (2020). Circular economy in food industry. *Zaštita materijala*, 61, 229–250. <https://doi.org/10.5937/zasmat2003229R6>
- Razungles, A. (2022). Extraction technologies and wine quality. In: Managing Wine Quality. *Elsevier*, 3–41. <https://doi.org/10.1016/B978-0-08-102065-4.00009-2>
- Reynolds, J. C, Meusel, R. C, Catania, A. A, & Casassa, L. F. (2022). Chemical and Chromatic Effects of Fermentation Temperature on Three Clones of Pinot noir over Two Consecutive Vintages. *Am. J. Enol. Vitic.*, 73, 75-92. <https://doi.org/10.5344/ajev.2021.21035>
- Ribéreau-Gayon, P. (1982). The anthocyanins of grapes and wines. In: Markakis, P, Ed., *Anthocyanins as Food Colors*, Academic Press, New York, 209-244.
- Ribéreau-Gayon, P, Dubourdieu, D, & Donèche, B. (2006). Handbook of enology, 2nd ed. *John Wiley*
- Río Segade, S, Pace, C, Torchio, F, Giacosa, S, Gerbi, V, & Rolle, L. (2015). Impact of maceration enzymes on skin softening and relationship with anthocyanin extraction in wine grapes with different anthocyanin profiles. *Food Research International*, 71, 50–57. <https://doi.org/10.1016/j.foodres.2015.02.012>
- Roubelakis-Angelakis, K. A. (2001). Molecular Biology & Biotechnology of the Grapevine. *Springer*, XXIII, 474. <https://doi.org/10.1007/978-94-017-2308-4>
- Roubelakis-Angelakis, K. A. (2009). Grapevine Molecular Physiology & Biotechnology. *Springer*, XXXVI, 610. <https://doi.org/10.1007/978-90-481-2305-6>

- Sabra, A., Neticadan, T., & Wijekoon, C. (2021). Grape bioactive molecules, and the potential health benefits in reducing the risk of heart diseases. *Food Chemistry*, 12, 100149. <https://doi.org/10.1016/j.fochx.2021.100149>
- Santos, M. C., Nunes, C., Cappelle, J., Goncalves, F. J., Rodrigues, A., Saraiva, J. A., & Coimbra, M. A. (2013). Effect of high pressure treatments on the physicochemical properties of a sulphur dioxide-free red wine. *Food Chemistry*, 141, 2558-2566. <http://dx.doi.org/10.1016/j.foodchem.2013.05.022>
- Setford, P. C., Jeffery, D. W., Grbin, P. R., & Muhlack, R. A. (2017). Factors affecting extraction and evolution of phenolic compounds during red wine maceration and the role of process modelling. *Trends in Food Science & Technology*, 69, 106–117. <https://doi.org/10.1016/j.tifs.2017.09.005>
- Silva, A., Silva, V., Igrejas, G., Gaivao, I., Aires, A., Klibi, N., de Lurdes Enes Dapkevicius, M., Valentao, P., Falco, V., & Poeta, P. (2021). Valorization of Winemaking By-Products as a Novel Source of Antibacterial Properties: New Strategies to Fight Antibiotic Resistance. *Molecules*, 26, 2331. <https://doi.org/10.3390/molecules26082331>
- Sikuten, I., Stambuk, P., Tomaz, I., Marchal C., Karoglan Kontic, J., Lacombe, T., Maletic, E., & Preiner, D. (2021). Discrimination of genetic and geographical groups of grape varieties (*Vitis vinifera* L.) based on their polyphenolic profiles. *Journal of Food Composition and Analysis*, 102, 104062. <https://doi.org/10.1016/j.jfca.2021.104062>
- Sipiora, M. J., & Gutierrez-Granda, M.-J. (1998). Effect of Pre-Veraison Irrigation Cutoff and Skin Contact Time on the Composition, Color, and Phenolic Content of Young Cabernet Sauvignon Wines in Spain. *Am. J. Enol. Vitic.*, 49, 152-162, <https://doi.org/10.5344/ajev.1998.49.2.152>
- Sivilotti, P., Falchi, R., Vanderweide, J., Sabbatini, P., Bubola, M., Vanzo, A., Lisjak, K., Peterlunger, E., & Herrera, J. C. (2020). Yield reduction through cluster or selective berry thinning similarly modulates anthocyanins and proanthocyanidins composition in Refosco dal peduncolo rosso (*Vitis vinifera* L.) grapes. *Scientia Horticulturae*, 264, 109166. <https://doi.org/10.1016/j.scienta.2019.109166>
- Soares, S., Santos Silva, M., Garcia-Estevez, I., Brandao, E., Fonseca, F., Ferreira-da-Silva, F., Escribano-Bailon, M. T., Mateus, N., & de Freitas, V. (2019). Effect of malvidin-3-glucoside and epicatechin interaction on their ability to interact with salivary proline-rich proteins. *Food Chemistry*, 276, 33-42. <https://doi.org/10.1016/j.foodchem.2018.09.167>
- Soares, S., Brandao, E., Guerreiro, C., Soares, S., Mateus, N., & de Freitas, V. (2020). Tannins in Food: Insights into the Molecular Perception of Astringency and Bitter Taste. *Molecules*, 25, 2590. <https://doi.org/10.3390/molecules25112590>
- Soceanu, A., Dobrinas, S., Sibru, A., Manea, N., & Popescu, V. (2021). Economic aspects of waste recovery in the wine industry. A multidisciplinary approach. *Science of the Total Environment*, 759, 143543. <https://doi.org/10.1016/j.scitotenv.2020.143543>
- Sommer, S., & Cohen, S. D. (2018). Comparison of Different Extraction Methods to Predict Anthocyanin Concentration and Color Characteristics of Red Wines. *Fermentation*, 4, 39. <https://doi.org/10.3390/fermentation4020039>
- Soto Vasquez, E., Rio Segade, S., & Fernandez Gomez, E. (2013). Incidence of the Winemaking Technique on Metal Content and Phenolic Composition of Red Wines. *International Journal of Food Properties*, 16, 622-633. <https://doi.org/10.1080/10942912.2011.558228>
- Shaddel, R., Hesari, J., Azadmard-Damirchi, S., Hamishehkar, H., Fathi-Achachlouei, B., & Huang, Q. (2018). Double emulsion followed by complex coacervation as a promising method for protection of black raspberry anthocyanins. *Food Hydrocolloids*, 77, 803-816. <https://doi.org/10.1016/j.foodhyd.2017.11.024>
- Sharif, N., Khoshnoudi-Nia, S., & Mahdi Jafari, S. (2020). Nano/microencapsulation of anthocyanins; a systematic review and meta-analysis. *Food Research International*, 132, 109077. <https://doi.org/10.1016/j.foodres.2020.109077>
- Shmigelskaya, N., Cherviak, S., Makarov, A., Sivochoub, G., & Vesytova, A. (2021). The influence of extraction methods on the quality characteristics of red wines. *E3S Web Conf*, 316, 03019. <https://doi.org/10.1051/e3sconf/202131603019>
- Ševcech, J., Vicenova, L., Furdikova, K., & Malik, F. (2015). Influence of Thermal Treatment on Polyphenol Extraction of Wine cv. Andre. *Czech J. Food Sci*, 33, 91-96. <https://doi.org/10.17221/286/2014-CJFS>
- Šuković, D., Knežević, B., Gašić, U., Sredojević, M., Ćirić, I., Todić, S., Mutić, J., & Tešić, Ž. (2020). Phenolic Profiles of Leaves, Grapes and Wine of Grapevine Variety Vranac (*Vitis vinifera* L.) from Montenegro. *Foods*, 9, 138. <https://doi.org/10.3390/foods9020138>
- Tampaktsi, C., Gancel, A. L., Escudier, J-L., Samson, A., Ojeda, H., Pic, L., Rousseau, J., Gauthier, P., Viguier, D., Furet, M. I., & Teissedre, P-L. (2023). Phenolic potential of new red hybrid grape varieties to produce quality wines and identification by the malvin. *BIO Web Conf*, 56, 02012. <https://doi.org/10.1051/bioconf/20235602012>
- Tao, Y., Sun, D-W, Gorecki, A., Blaszcak, W., Lamparski, G., Amarowicz, R., Fornal, J., & Jelinski, T. (2012). Effects of high hydrostatic pressure processi on the physicochemical and sensorial properties of a red wine. *Innovative Food Science and Emerging Technologies*, 16, 409-416. <http://dx.doi.org/10.1016/j.ifset.2012.09.005>
- Tao, Y., Sun, D-W, Gorecki, A., Blaszcak, W., Lamparski, G., Amarowicz, R., Fornal, J., & Jelinski, T. (2016). A preliminary study about the influence of high hydrostatic pressure processing in parallel with oak chip maceration on the physicochemical and sensory properties of a young red wine. *Food Chemistry*, 194, 545-554. <http://dx.doi.org/10.1016/j.foodchem.2015.07.041>
- Tassoni, A., Zappi, A., Melucci, D., Reisch, I. B., & Davies, J. P. (2019). Seasonal changes in amino acids and phenolic compounds in fruits from hybrid cross populations of American grapes differing in disease resistance. *Plant Physiology and Biochemistry*, 135, 182–193. <https://doi.org/10.1016/j.plaphy.2018.11.034>
- Teissedre, P-L, & Jourdes, M. (2013). Tannins and Anthocyanins of Wine: Phytochemistry and Organoleptic Properties. *Natural Products, Springer*, 2255-2271. https://doi.org/10.1007/978-3-642-22144-6_73
- Teissedre, P-L. (2018). Composition of grape and wine from resistant vines varieties. *OENO One*, 52, 211–217. <https://doi.org/10.20870/oeno-one.2018.52.3.2223>
- Teissedre, P-L, Stockley, C., Boban, M., Gambert, P., Ortiz Alba, M., Flesh, M., & Ruf, J-C. (2018). The effect of wine consumption on cardiovascular disease and associated risk factors: a narrative review. *OENO One*, 52, 62-79. <https://doi.org/10.20870/oeno-one.2018.52.1.2129>
- Teixeira Gomez, B., Rodrigez Borges, L. L., Mitterhofer Eiterer, N., de Leon da Costa, P., Rodrigues Arruda, T., Costa Ribeiro, A. R., Suprani Marques, C., Stringheta, C., Veloso de Oliveira, T., & De Fatima Ferreira Soares, N. (2024). Gelatin/polyvinyl alcohol films incorporated with different blueberry extracts as potential colorimetric indicators to detect acidic and basic vapor. *Food Control*, 165, 110648. <https://doi.org/10.1016/j.foodcont.2024.110648>
- Tikhonova, A., Ageeva, N., & Globa, E. (2021). Grape pomace as a promising source of biologically. *BIO Web of Conferences*, 34, 06002. <https://doi.org/10.1051/bioconf/20213406002>
- Tong, W, Sun, B, Ling, M, Zhang, X, Yang, W, Shi, Y, Pan, Q, Duan, C, & Lan, Y. (2023). Influence of modified carbonic maceration technique on the chemical and sensory characteristics of Cabernet Sauvignon wines. *Food Chemistry*, 403, 134341. <https://doi.org/10.1016/j.foodchem.2022.134341>

- Torres-Rochera, B., Manjon, E., Escribano-Bailon, M., & Garcia-Estevéz, I. (2023). Role of Anthocyanins in the Interaction between Salivary Mucins and Wine Astringent Compounds. *Foods*, 12, 3623. <https://doi.org/10.3390/foods1219362>
- Troilo, M., Difonzo, G., Paradiso, V. M., Pasqualone, A., & Caponio, F. (2022). Grape Pomace as Innovative Flour for the Formulation of Functional Muffins: How Particle Size Affects the Nutritional, Textural and Sensory Properties. *Foods*, 11, 1799. <https://doi.org/10.3390/foods11121799>
- Valls, J., Agnolet, S., Haas, F., Struffi, I., Ciesa, F., Robatscher, P., & Oberhuber, M. (2017). Valorization of Lagrein grape pomace as a source of phenolic compounds: analysis of the contents of anthocyanins, flavanols and antioxidant activity. *Eur Food Res Technol*, 243, 2211–2224. <https://doi.org/10.1007/s00217-017-2923-1>
- Villamor, R. R., Harbertson, J. F., & Ross, C. F. (2009). Influence of Tannin Concentration, Storage Temperature and Time on Chemical and Sensory Properties of Cabernet Sauvignon and Merlot Wines. *Am. J. Enol. Vitic.*, 60, 442–449. <https://doi.org/10.5344/ajev.2009.60.4.442>
- Vermerris, W., Nicholson, R. L. (2006). Phenolic compound biochemistry. *Springer*, XII, 276. <https://doi.org/10.1007/978-1-4020-5164-7>
- Wang, L., Zhou, W., Liu, C., Chen, P., & Zhou, L. (2024). Study on the accumulation pattern of anthocyanins, sugars and organic acids in medicinal *Vitis vinifera* ‘SuoSuo’ during ripening. *Food Chemistry*, 433, 137294. <https://doi.org/10.1016/j.foodchem.2023.137294>
- Wang, J., Huo, S., Zhang, Y., Liu, Y., & Fan, W. (2016). Effect of different pre-fermentation treatments on polyphenols, color, and volatile compounds of three wine varieties. *Food Sci Biotechnol*, 25, 735–743. <https://doi.org/10.1007/s10068-016-0127-2>
- Wang, Y., Yang, C., Zhang, J., & Zhang, L. (2022). Interaction between whey protein isolate and rose anthocyanin extracts at different pHs: Structure, emulsification and digestibility of complexes. *Food Bioscience*, 49, 101888. <https://doi.org/10.1016/j.fbio.2022.101888>
- Wang, Z., Zhang, L., Li, Y., Liu, Q., & Yuan, C. (2023). Non-acylated and acylated anthocyanins in red wines of different ages: Color contribution and evaluation. *Journal of Food Composition and Analysis*, 115, 104951. <https://doi.org/10.1016/j.jfca.2022.104951>
- Waterhouse, A., & Zhu, J. (2019). A quarter century of wine pigment discovery. *Society of Chemical Industry*, 100, 5093-5101. <https://doi.org/10.1002/jsfa.9840>
- Watrelet, A. A., & Waterhouse, A. L. (2018). Oak barrel tannin and toasting temperature: Effects on red wine anthocyanin chemistry. *LWT*, 98, 444–450. <https://doi.org/10.1016/j.lwt.2018.09.025>
- Winefield, C., Davies, K., & Gould, K. (2009). Anthocyanins. *Springer New York*, XVIII, 336. <https://doi.org/10.1007/978-0-387-77335-3>
- Wojdyło, A., Samoticha, J., Nowicka, P., & Chmielewska, J. (2018). Characterisation of (poly)phenolic constituents of two interspecific red hybrids of Rondo and Regent (*Vitis vinifera*) by LC-PDA-ESI-MS QToF. *Food Chemistry*, 239, 94–101. <https://doi.org/10.1016/j.foodchem.2017.06.077>
- Wojdyło, A., Samoticha, J., & Chmielewska, J. (2021). Effect of different pre-treatment maceration techniques on the content of phenolic compounds and color of Dornfelder wines elaborated in cold climate. *Food Chemistry*, 339, 127888. <https://doi.org/10.1016/j.foodchem.2020.127888>
- Wu, X., Liu, Z., Liu, J., He, S., & Shao, W. (2024). Development of novel intelligent and edible dual-functional electrospun nanofiber films for shrimp preservation and freshness monitoring in real-time. *Food Bioscience*, 61, 104828. <https://doi.org/10.1016/j.fbio.2024.104828>
- Yao, J-W, Lin, F, Tao, T., & Lin, C-J. (2011). Affinity interactions between natural pigments and human whole saliva. *Archives of Oral Biology*, 56, 285-293. <https://doi.org/10.1016/j.archoralbio.2010.10.003>
- Yang, Y, Li, H, Li, H, Zheng, H, & Tao, J. (2023). Transcriptome analysis reveals the mechanism of ‘ZhongShan-HongYu’ grape anthocyanin accumulation. *Scientia Horticulturae*, 321, 112331. <https://doi.org/10.1016/j.scienta.2023.112331>
- Zhang, Y-S, Du, G, Gao, Y-T, Wang, L-W, Meng, D, Li, B-J, Brennan, C, Wang, M-Y, Zhao, H, Wang, S-Y, Guan, W-Q. (2019). The Effect of Carbonic Maceration during Winemaking on the Color, Aroma and Sensory Properties of ‘Muscat Hamburg’ Wine. *Molecules*, 24, 3120. <https://doi.org/10.3390/molecules24173120>
- Zhang, R, Zhou, L, Li, J, Oliveira, H, Yang, N, Jin, W, Zhu, Z, Li, S, & He, J. (2020). Microencapsulation of anthocyanins extracted from grape skin by emulsification/internal gelation followed by spray/freeze-drying techniques: Characterization, stability and bioaccessibility. *LWT – Food Science and Technology*, 123, 109097. <https://doi.org/10.1016/j.lwt.2020.109097>
- Zhang, X-K, Lan, Y-B, Huang, Y, Zhao, Y, & Duan, C-Q. (2021). Targeted metabolomics of anthocyanin derivatives during prolonged wine aging: Evolution, color contribution and aging prediction. *Food Chemistry*, 339, 127795. <https://doi.org/10.1016/j.foodchem.2020.127795>
- Zhang, X-K, Jeffery, D. W, Li, D-M, Lan, Y-B, Zhao, X, & Duan, C-Q. (2022). Red wine coloration: A review of pigmented molecules, reactions, and applications. *Comprehensive Reviews in Food Science and Food Safety*, 21, 3834-3866. <https://doi.org/10.1111/1541-4337.13010>
- Zhao, Q, Duan, C-Q, & Wang, J. (2010). Anthocyanins Profile of Grape Berries of *Vitis amurensis*, Its Hybrids and Their Wines. *IJMS*, 11, 2212–2228. <https://doi.org/10.3390/ijms11052212>
- Zhao, J, & Dixon, R. A. (2010). The ‘ins’ and ‘outs’ of flavonoid transport. *Trends in Plant Science*, 15, 72–80. <https://doi.org/10.1016/j.tplants.2009.11.006>
- Zhao, C. L., Chen, Z. J, Bai, X. S, Ding, C, Long, T. J, Wei, F. G, & Miao, K. R. (2014). Structure-activity relationships of anthocyanidin glycosylation. *Mol Divers*, 18, 687–700. <https://doi.org/10.1007/s11030-014-9520-z>
- Zhao, Q, Du, G, Zhao, P, Guo, A, Cao, X, Cheng, C, Liu, H, Wang, F, Zhao, Y, Liu, Y, & Wang, X. (2023). Investigating wine astringency profiles by characterizing tannin fractions in Cabernet Sauvignon wines and model wines. *Food Chemistry*, 414, 135673. <https://doi.org/10.1016/j.foodchem.2023.135673>
- Zhou, D-D, Li, J, Xiong, R-G, Saimaiti, A, Huang, S-Y, Wu, S-X, Yang, Z-J, Shang, A, Zhao, C-N, Gan, R-Y, & Li, H-B. (2022). Bioactive Compounds, Health Benefits and Food Applications of Grape. *Foods*, 11, 2755. <https://doi.org/10.3390/foods11182755>
- Zhu, L, Zhang, Y, Deng, J, Li, H, & Lu, J. (2012). Phenolic Concentrations and Antioxidant Properties of Wines Made from North American Grapes Grown in China. *Molecules*, 17, 3304–3323. <https://doi.org/10.3390/molecules17033304>
- Zhu, L, Li, X, Hu, X, Wu, X, Liu, Y, Yang, Y, Zang, Y, Tang, H, Wang, C, & Xu, J. (2021). Quality Characteristics and Anthocyanin Profiles of Different *Vitis amurensis* Grape Cultivars and Hybrids from Chinese Germplasm. *Molecules*, 26, 6696. <https://doi.org/10.3390/molecules26216696>
- Zia ul Haq, M, Riaz, M, & Saad, B. (2016). Anthocyanins and Human Health: Biomolecular and therapeutic aspects. *Springer International Publishing, Cham*, XII, 138. <https://doi.org/10.1007/978-3-319-26456-1>
- Zuidam, N. J, & Nedovic, V. (2010). Encapsulation Technologies for Active Food Ingredients and Food Processing. *Springer*, XII, 400. <https://doi.org/10.1007/978-1-4419-1008-0>